

A fluorescence microscopy image showing several kidney organoids. The organoids are stained with various fluorescent markers, appearing in shades of blue, green, and red. They have a complex, branching structure with a central core and outer layers of cells. The background is dark, making the brightly stained organoids stand out.

# Generation of Kidney Organoids from human iPS cells

Minoru Takasato, PhD

RIKEN BDR

# What is Organoid?

Organoid = Organ + oid (something similar to a real organ)

A common definition: Self-organizing tissues from stem cells

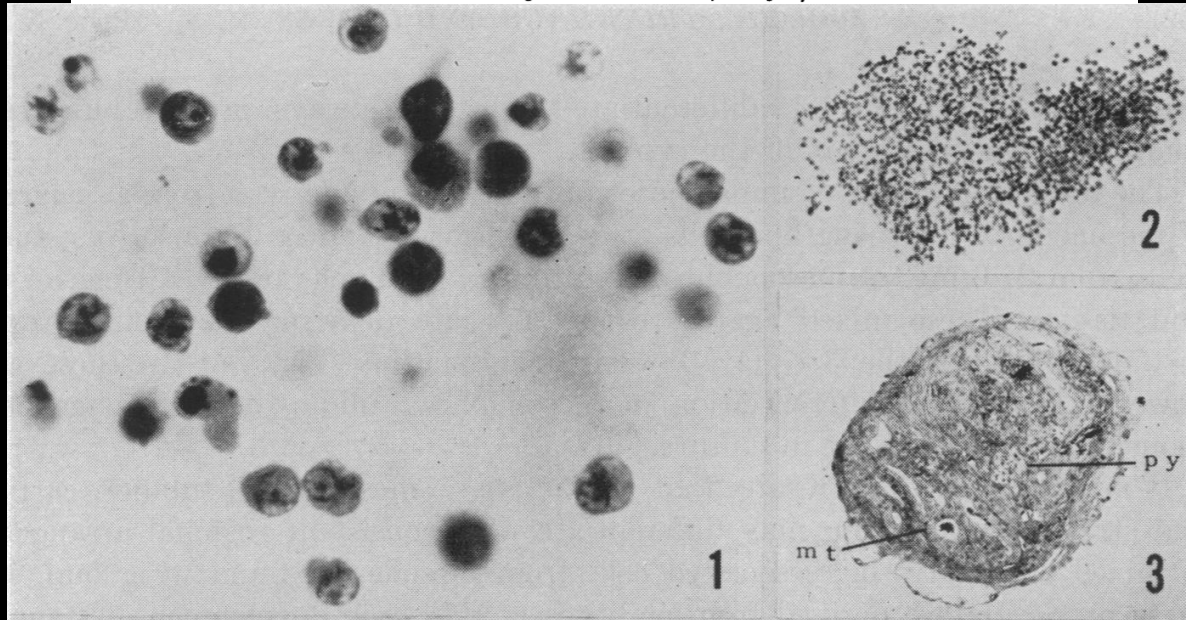
Originally, it was an aggregate of embryonic cells.

*DIFFERENTIATION IN CULTURE OF MIXED AGGREGATES OF  
DISSOCIATED TISSUE CELLS\**

BY J. P. TRINKAUS AND PEGGY W. GROVES

OSBORN ZOOLOGICAL LABORATORY, YALE UNIVERSITY

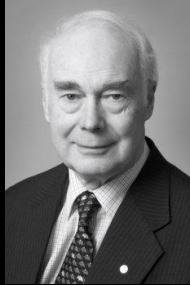
*Communicated by J. S. Nicholas, July 1, 1955*



(Trinkaus et al., *PNAS* 1955)



## Embryoid body



Sir Martin J. Evans

### Establishment in culture of pluripotential cells from mouse embryos

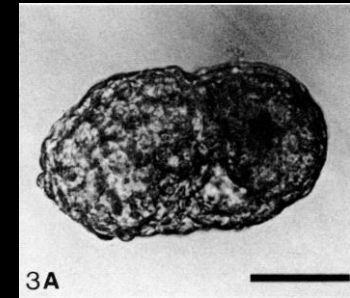
M. J. Evans\* & M. H. Kaufman†

Departments of Genetics\* and Anatomy†, University of Cambridge, Downing Street, Cambridge CB2 3EH, UK

Pluripotential cells are present in a mouse embryo until at least an early post-implantation stage, as shown by their ability to take part in the formation of chimaeric animals<sup>1</sup> and to form teratocarcinomas<sup>2</sup>. Until now it has not been possible to establish progressively growing cultures of these cells *in vitro*, and cell lines have only been obtained after teratocarcinoma formation *in vivo*. We report here the establishment in tissue culture of pluripotent cell lines which have been isolated directly from *in vitro* cultures of mouse blastocysts. These cells are able to differentiate either *in vitro* or after inoculation into a mouse as a tumour *in vivo*. They have a normal karyotype.

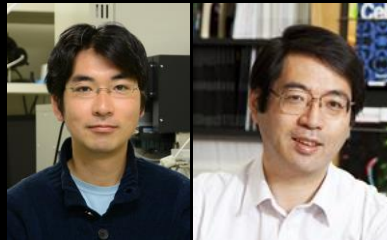
(Evans et al., *Nature* 1981)

### Embryoid body



(Doetschman et al., *Development* 1985)

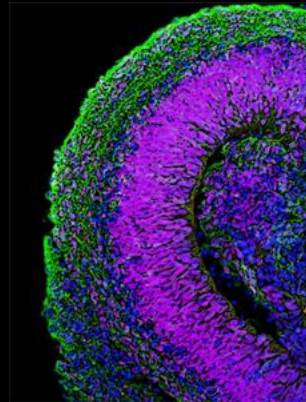
## Organoid from stem cells



M. Eiraku

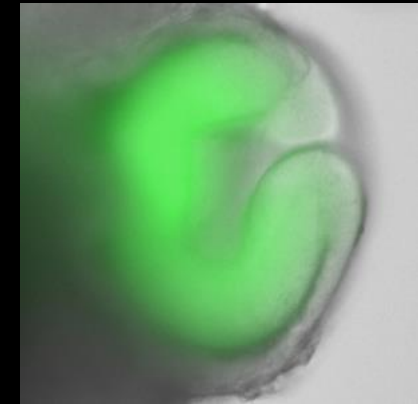
Y. Sasai

### Self-Organizing Polarized Cortical Tissues



(Eiraku et al., *Cell Stem Cell* 2008)

### Self-Organizing Optic-cup



(Eiraku et al., *Nature* 2011)

Organoids are not only interesting tools for studying organogenesis but also could be useful platforms for medical applications

# Potential of organoid technologies

CellPress

Cell Stem Cell  
Short Article

## Zika Virus Depletes Neural Progenitors in Human Cerebral Organoids through Activation of the Innate Immune Receptor TLR3

Jason Dang,<sup>1,2,6</sup> Shashi Kant Tiwari,<sup>1,6</sup> Gianluigi Lichinchi,<sup>1,3</sup> Yue Qin,<sup>1</sup> Veena S. Patil,<sup>1</sup> Alexey M. Eroshkin,<sup>4</sup> and Tariq M. Rana<sup>1,5,\*</sup>

<sup>1</sup>Department of Pediatrics, University of California San Diego, 9500 Gilman Drive, La Jolla, CA 92093, USA

<sup>2</sup>Department of Bioengineering, University of California San Diego, La Jolla, CA 92093, USA

<sup>3</sup>Graduate School of Biomedical Sciences

<sup>4</sup>Bioinformatics core

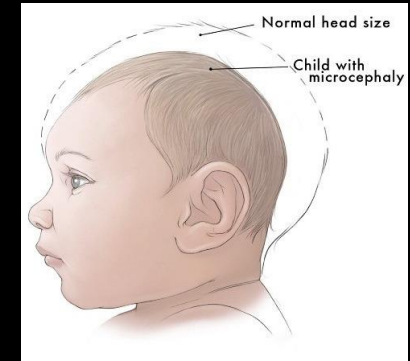
Sanford Burnham Prebys Medical Discovery Institute, 10901 North Torrey Pines Road, La Jolla, CA 92037, USA

<sup>5</sup>Institute for Genomic Medicine, University of California San Diego, La Jolla, CA 92093, USA

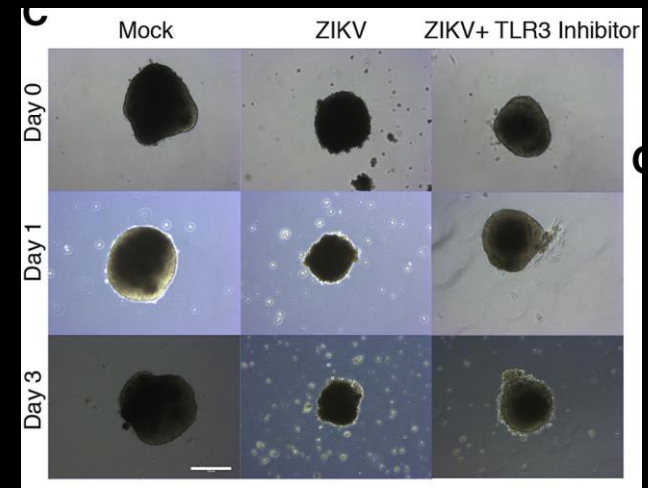
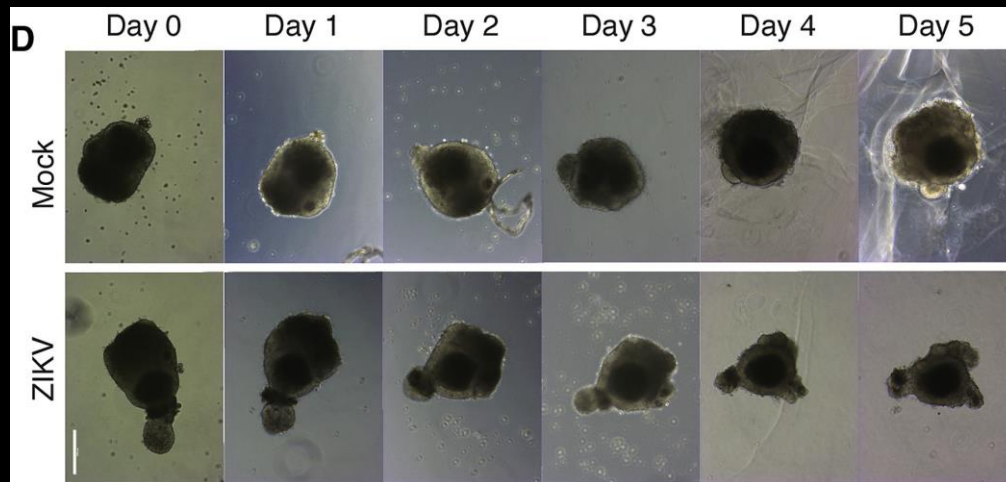
<sup>6</sup>Co-first author

\*Correspondence: [trana@ucsd.edu](mailto:trana@ucsd.edu)

<http://dx.doi.org/10.1016/j.stem.2016.04.014>

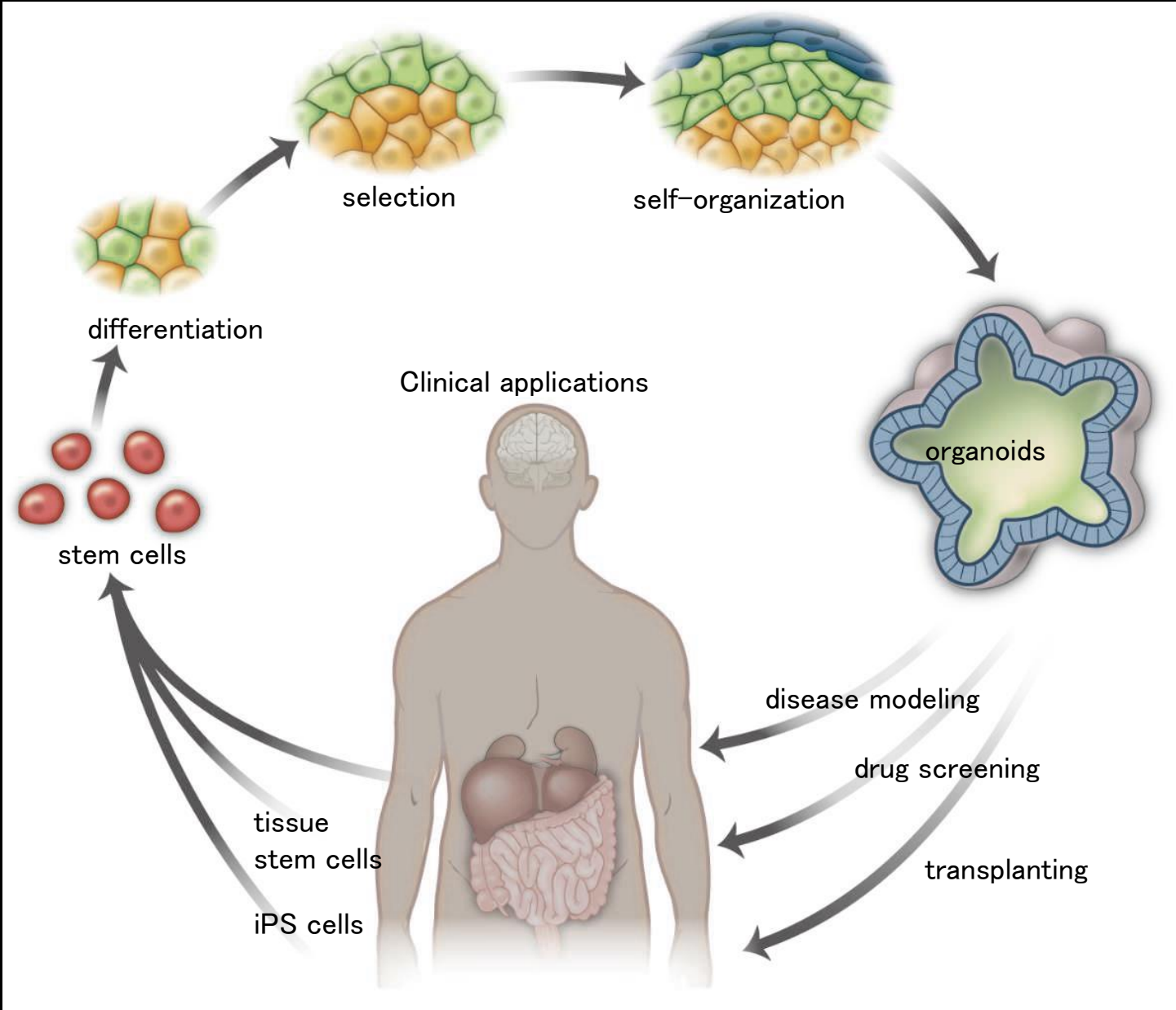


Zika virus infection during pregnancy causes microcephaly in a baby



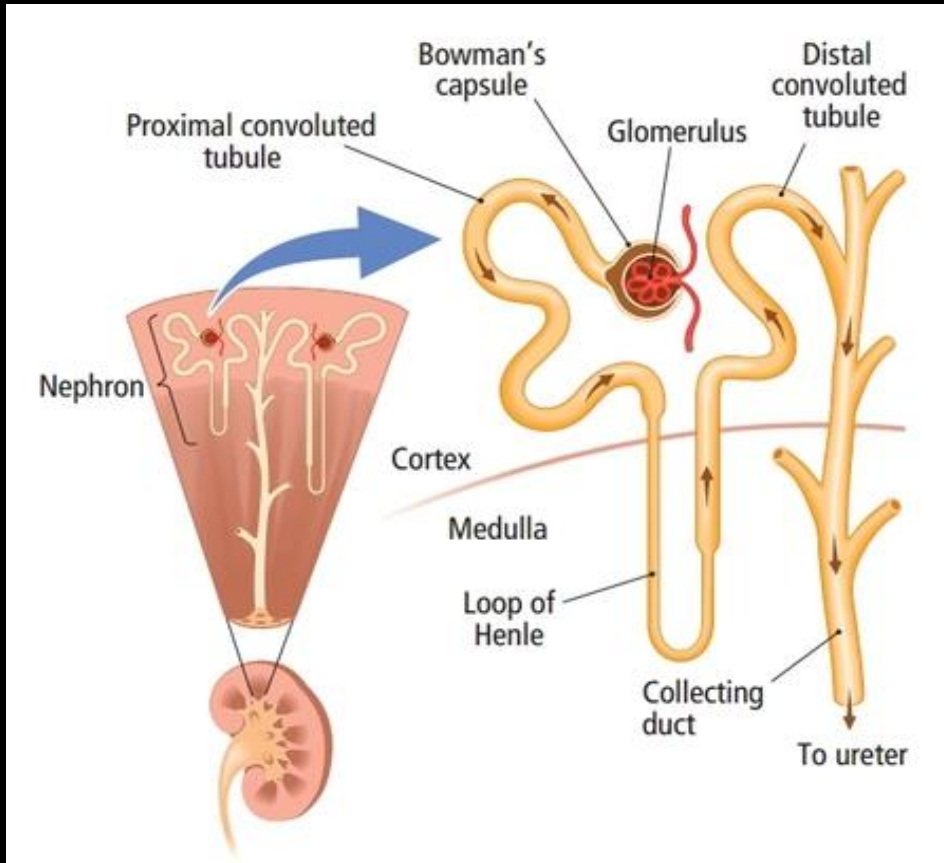
(J. Dong et al., *Cell Stem Cell* 2016)

# Potential of organoid technologies



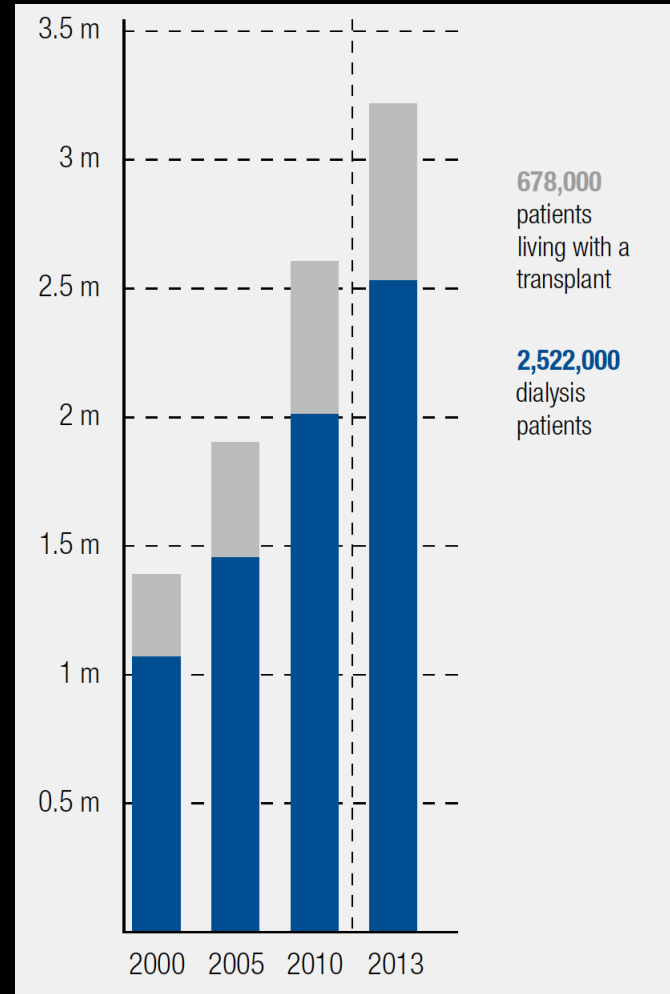
# Kidney anatomy

## Structure



leavingcertbiology.net (Chapter 37: The Human Urinary System)

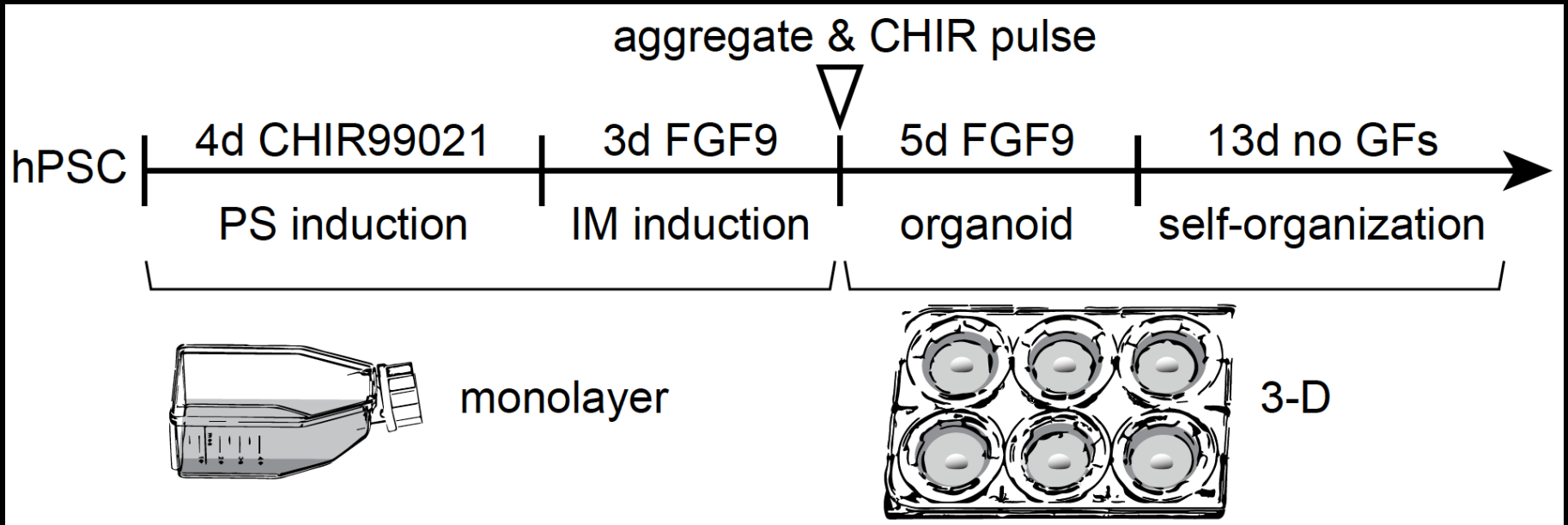
## # of ESKD patients in the world



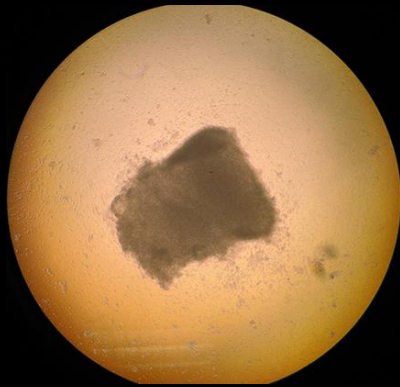
(Fresenius Medical Care Deutschland. 2014)



# A mixture of AI + PI generated kidney organoids



pellet



3 days



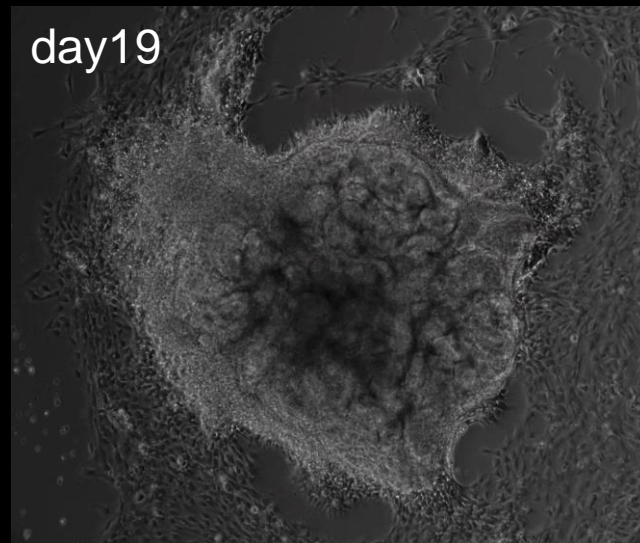
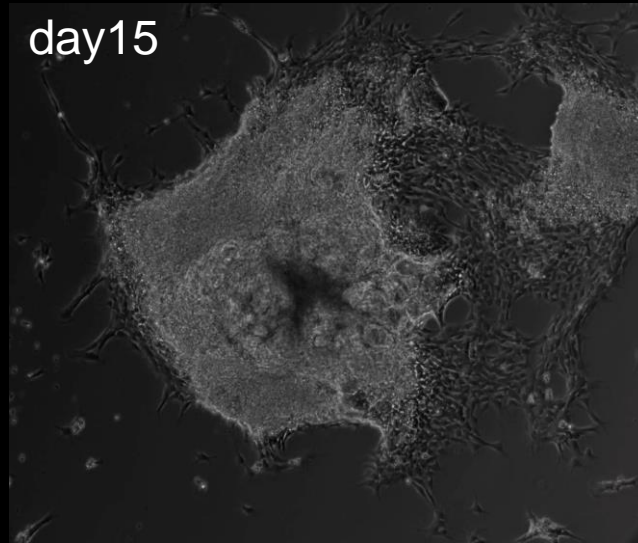
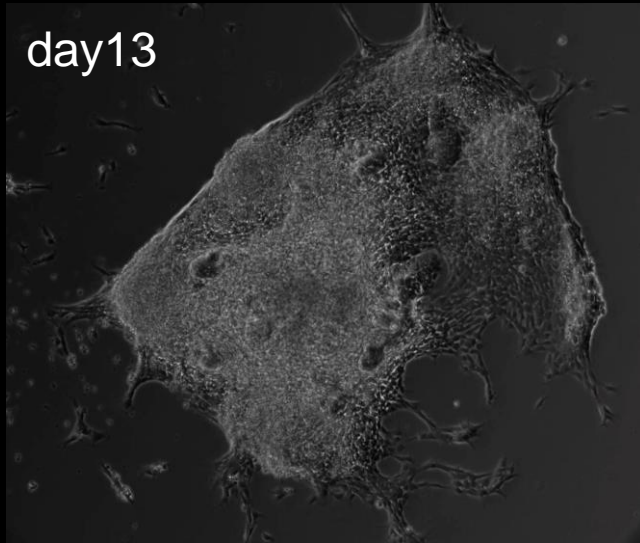
11 days



18 days

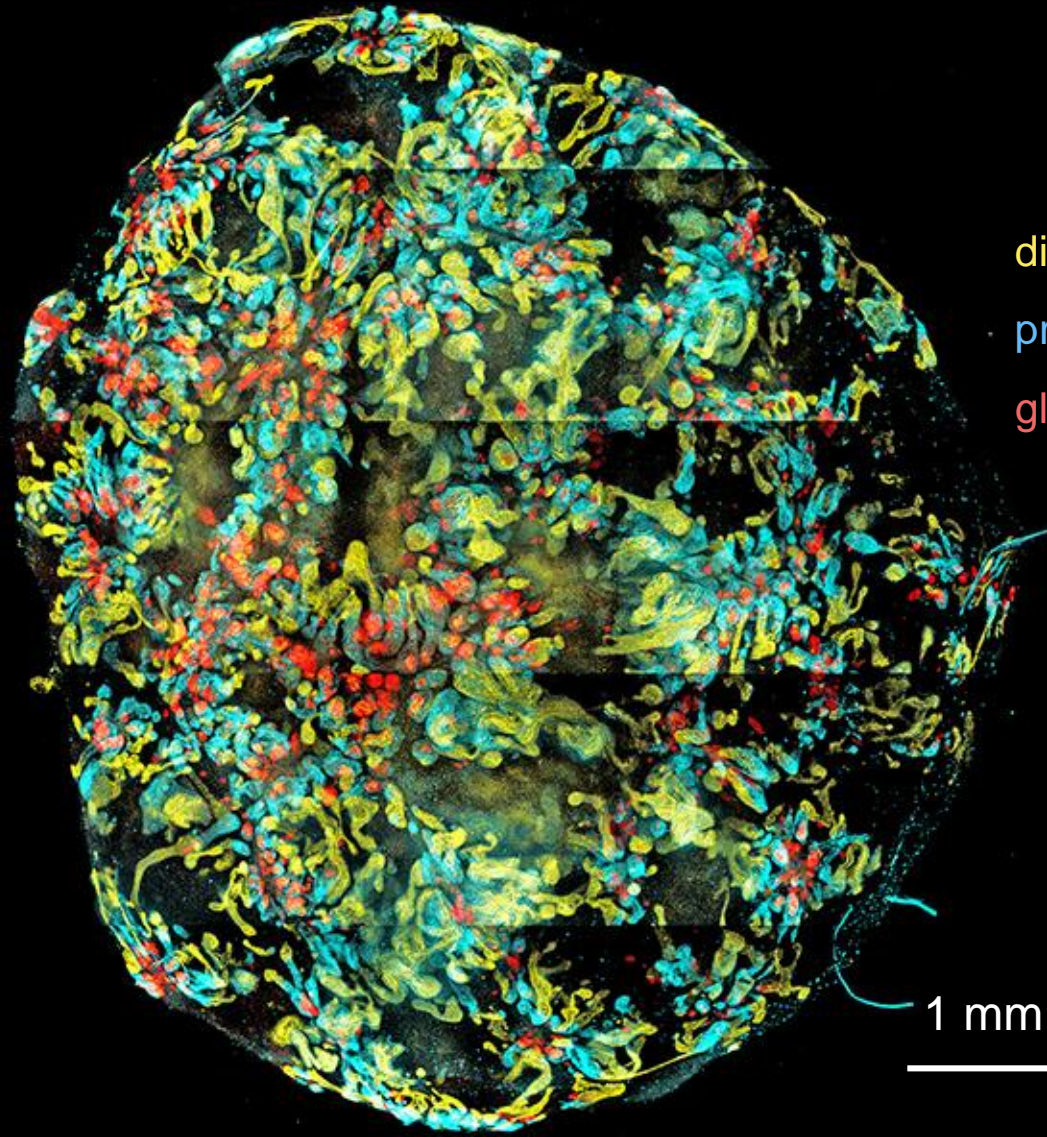


*Time-lapse of a developing kidney organoid*





*Kidney organoids comprised nephrons*



distal tubules (ECAD)

proximal tubules (LTL)

glomeruli (NPHS1)

1 mm

*Self-organizing nephrons consisted of 4 segments*

GATA3

ECAD

LTL

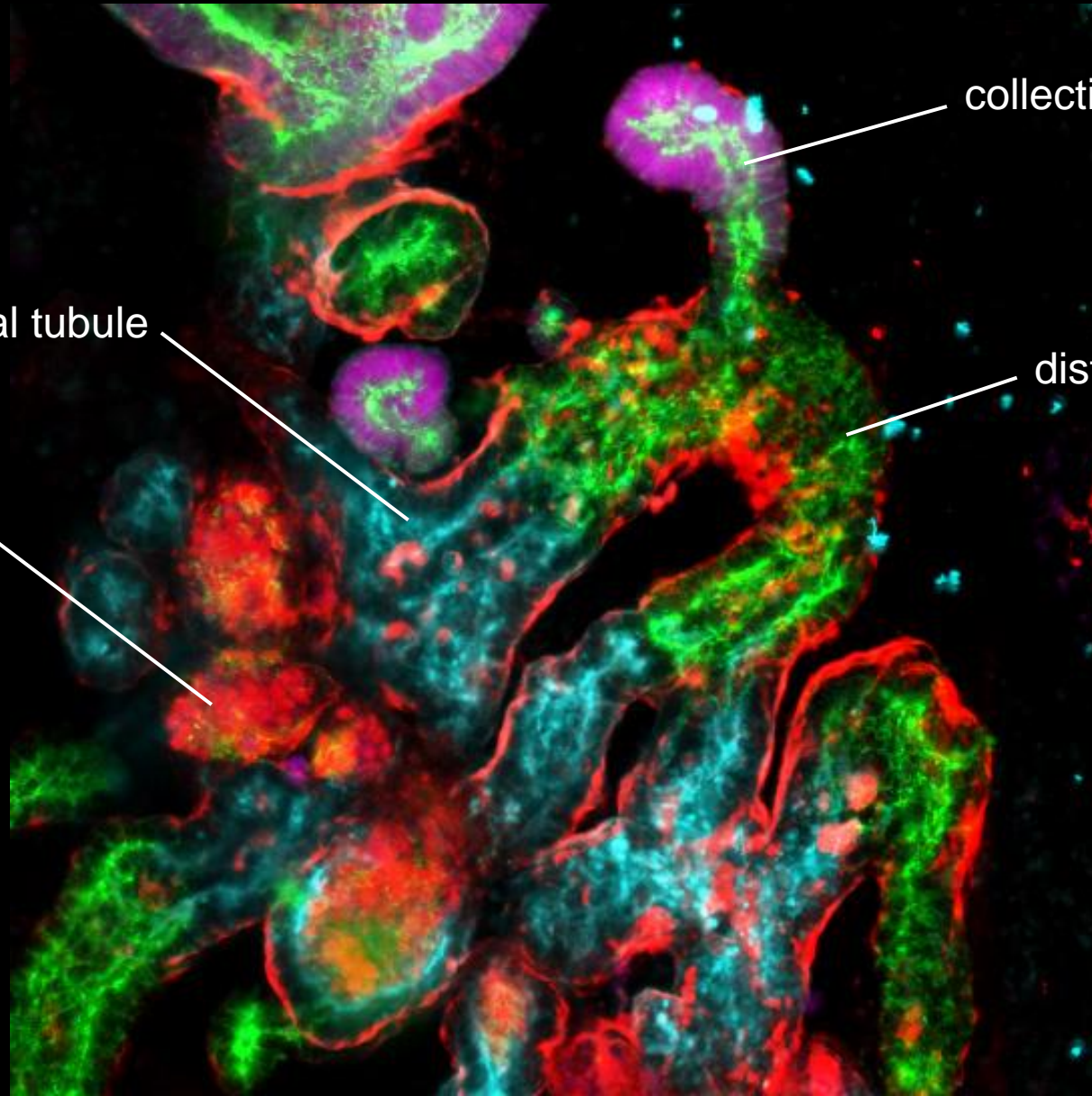
WT1

proximal tubule

collecting duct

distal tubule

glomerulus



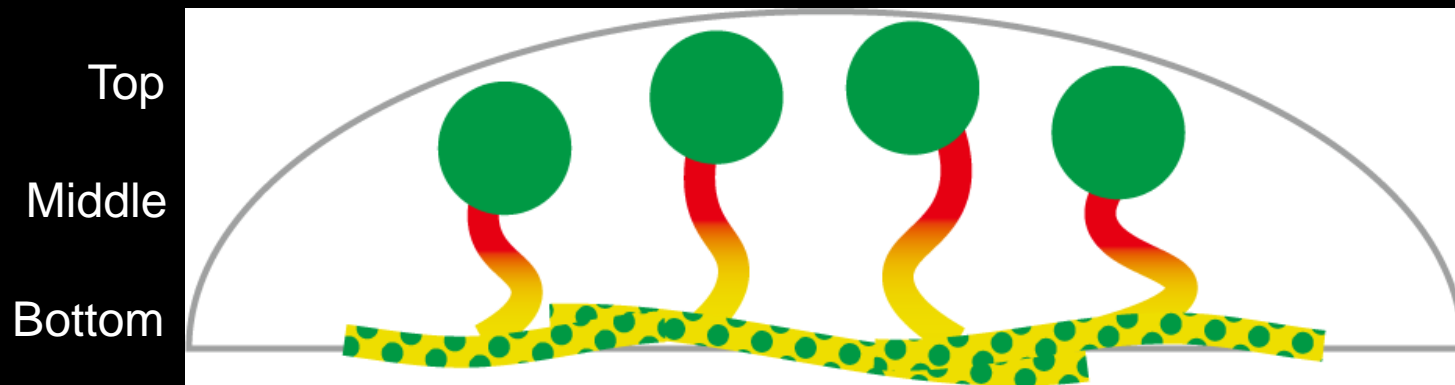
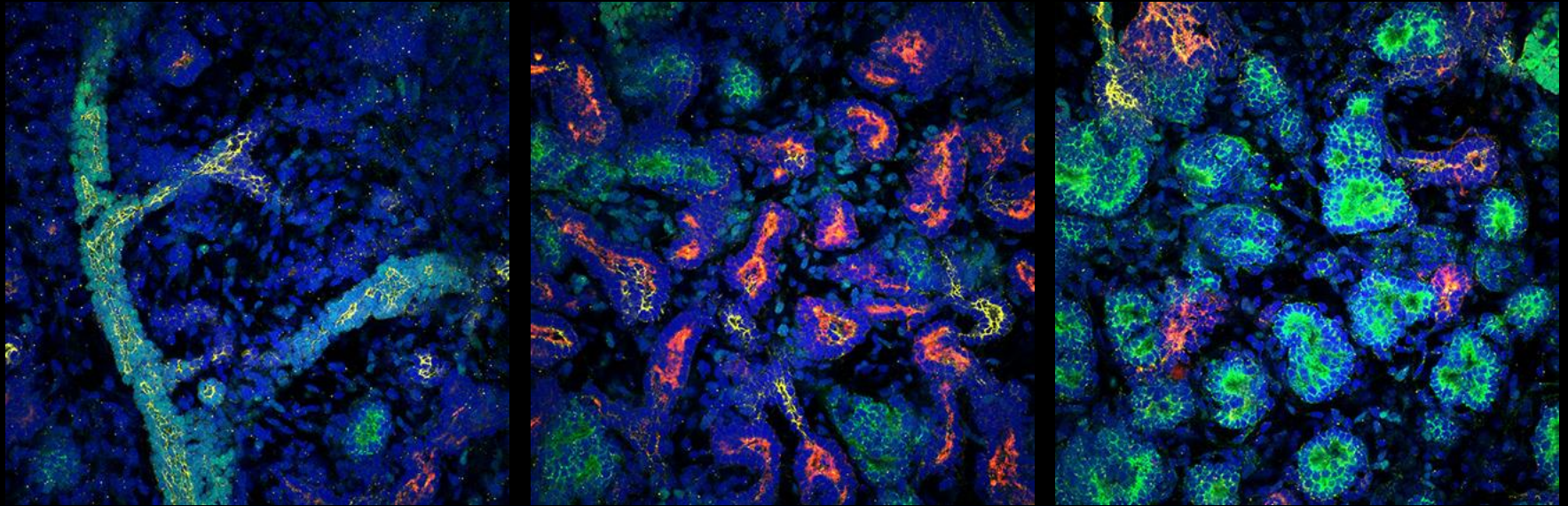


*Ducts in the bottom layer, Glomeruli in the outer layer*

Bottom

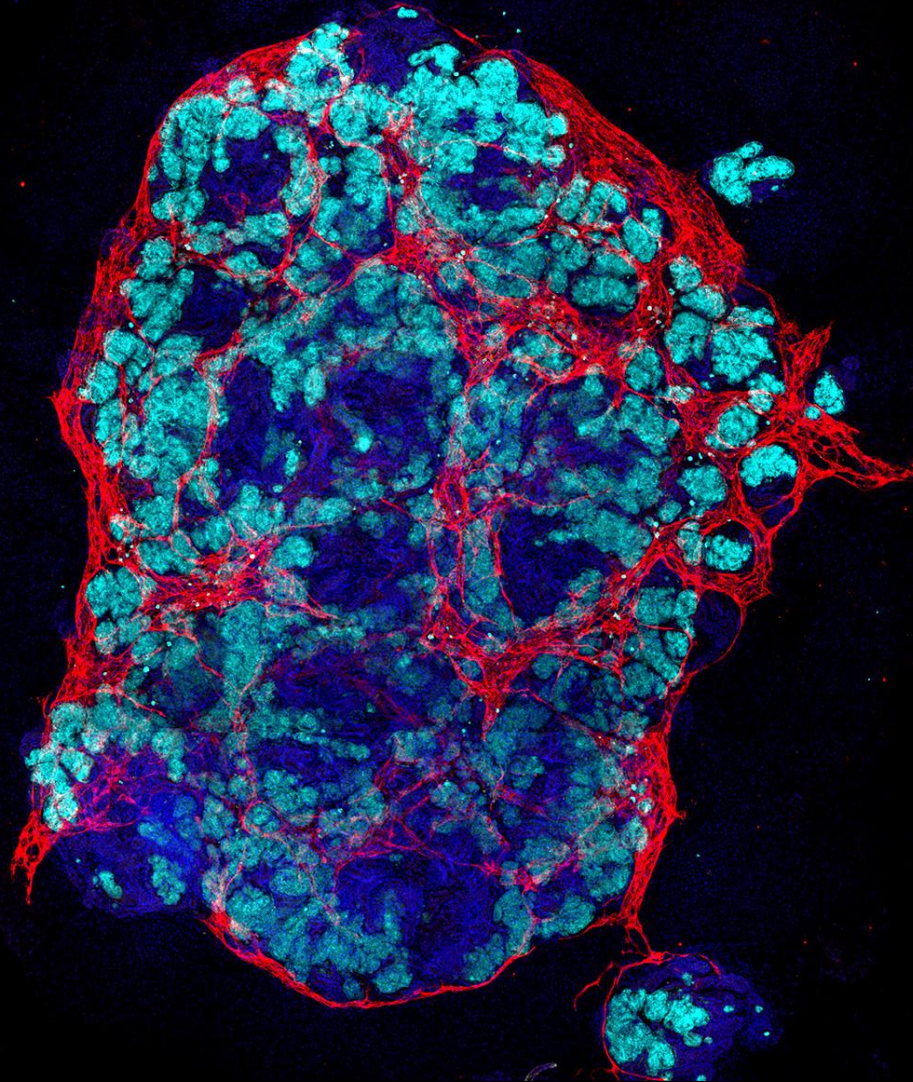
Middle

Top





*Kidney organoids contain vascular network*

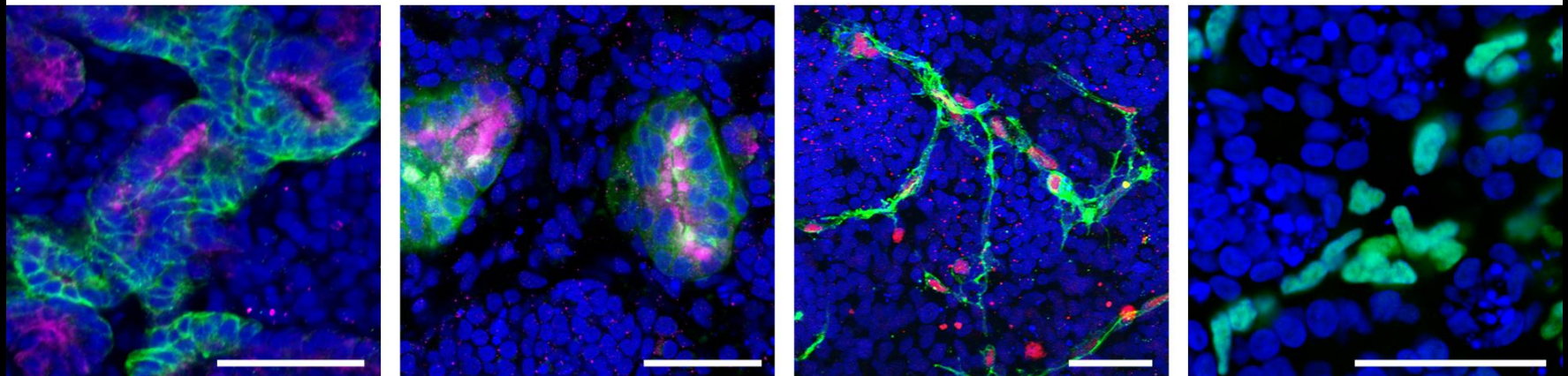
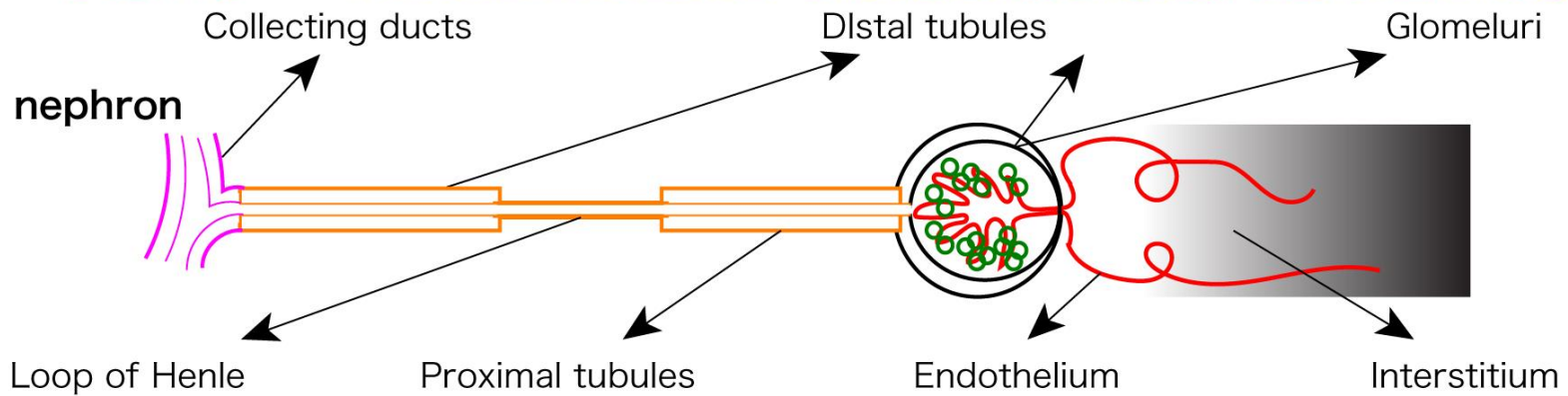
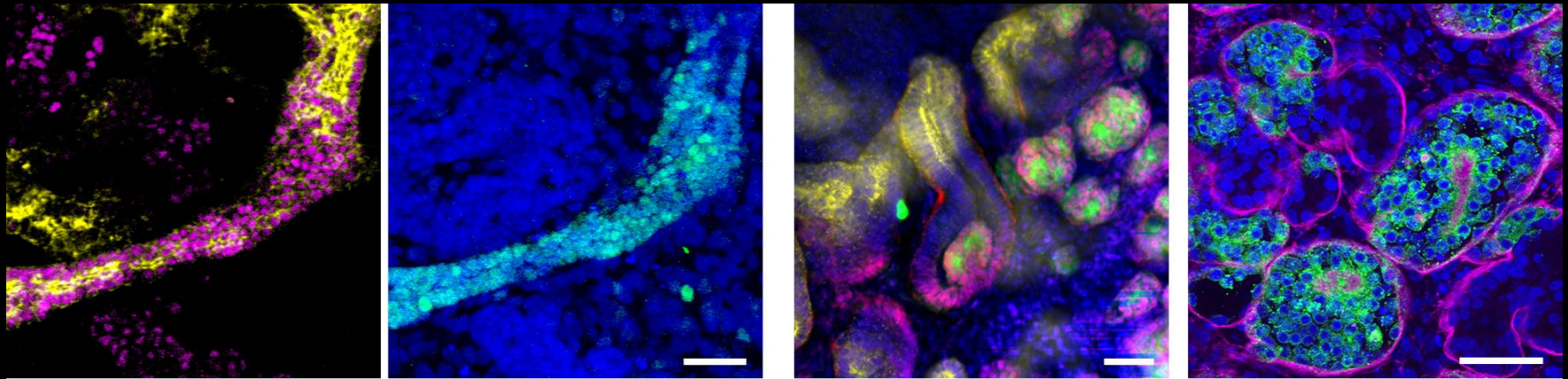


Glomerulus Vasculature

(Takasato et al., *Nature Protocols* 2016)

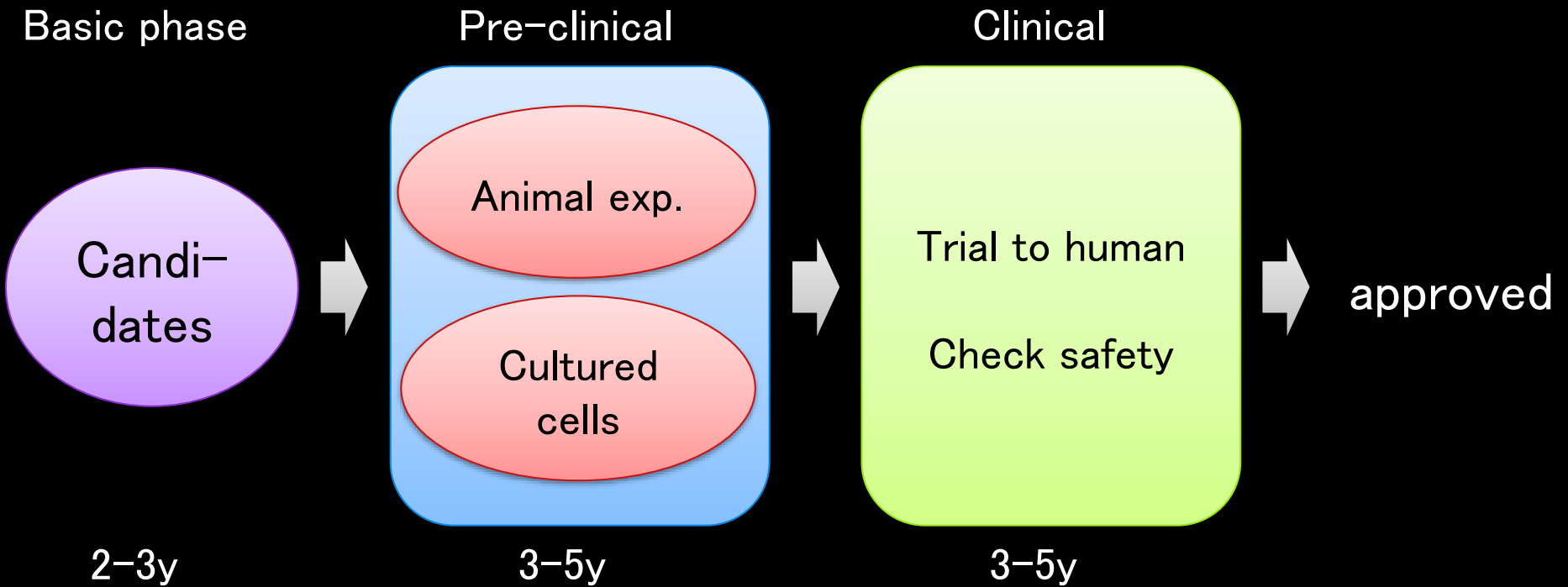


# All anticipated renal structures developed in kidney organoids



Scale = 100  $\mu$ m

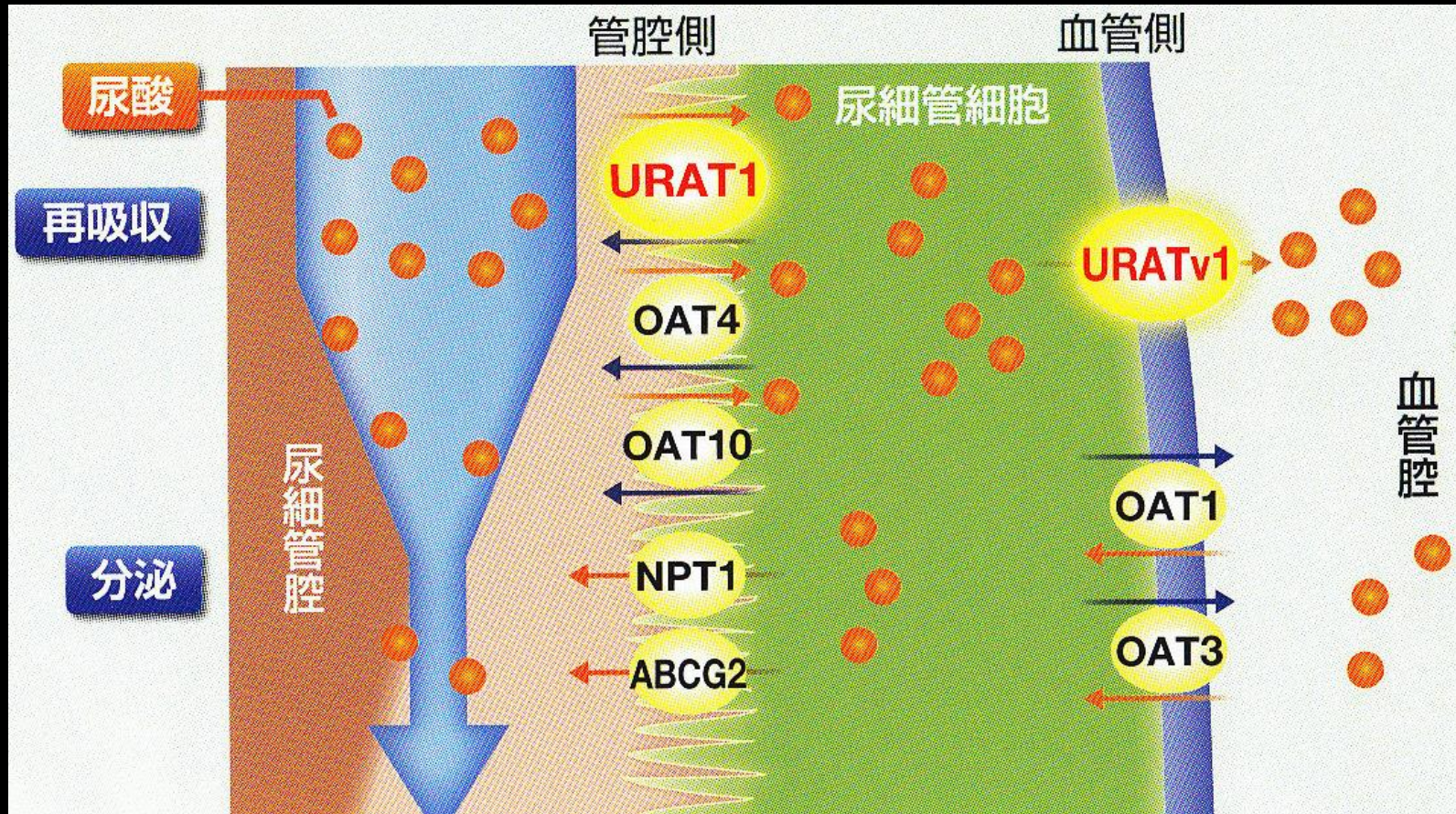
# *Nephrotoxicity assay for drug development*



Success rate: 1 / 30,000  
Cost: Billions of Yen

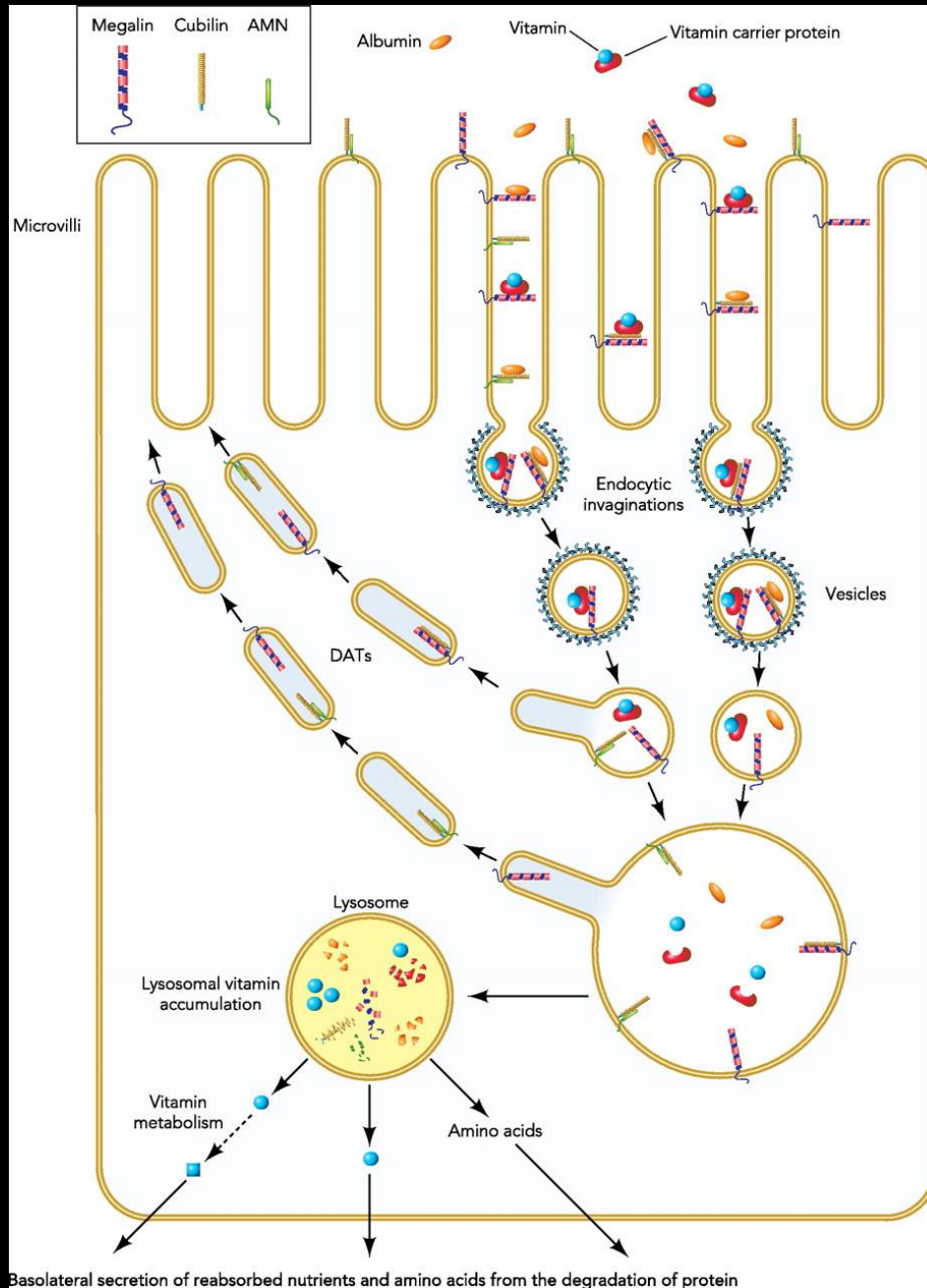


# Reabsorption function of renal proximal tubules



Physiological function drives pharmacokinetics

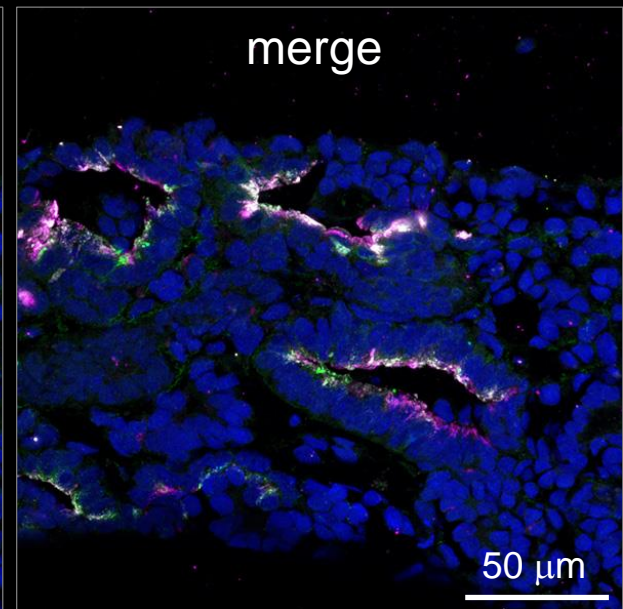
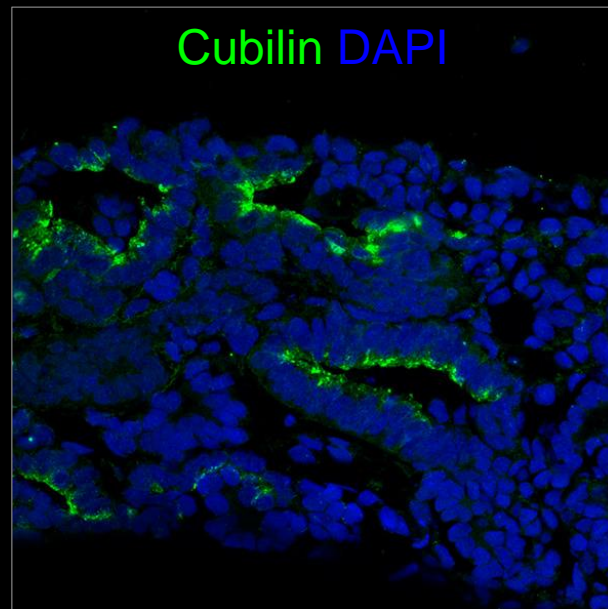
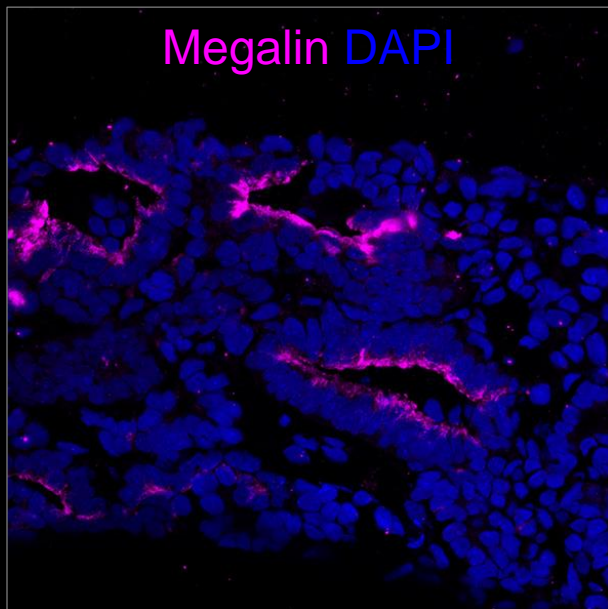
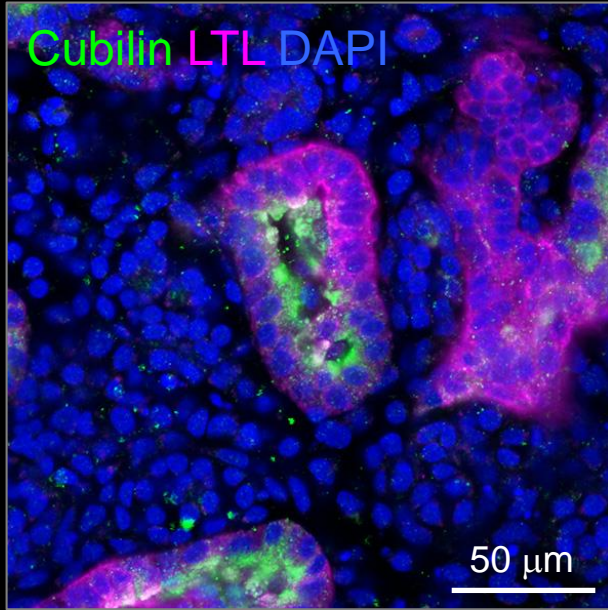
# Megalin-Cubilin mediated endocytosis in PT



(Christensen et al, *Physiology* 2012)



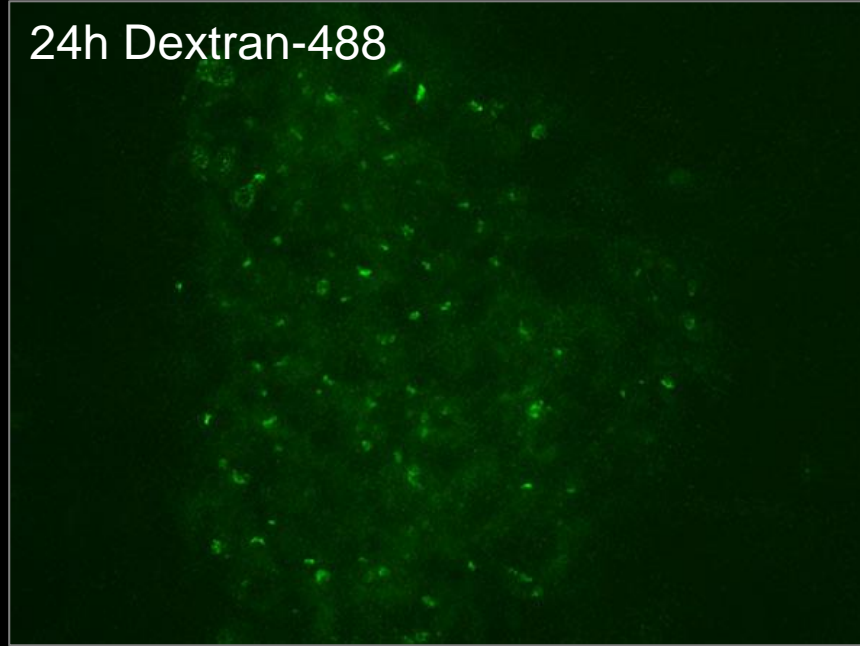
*PT expresses both Megalin and Cubilin*



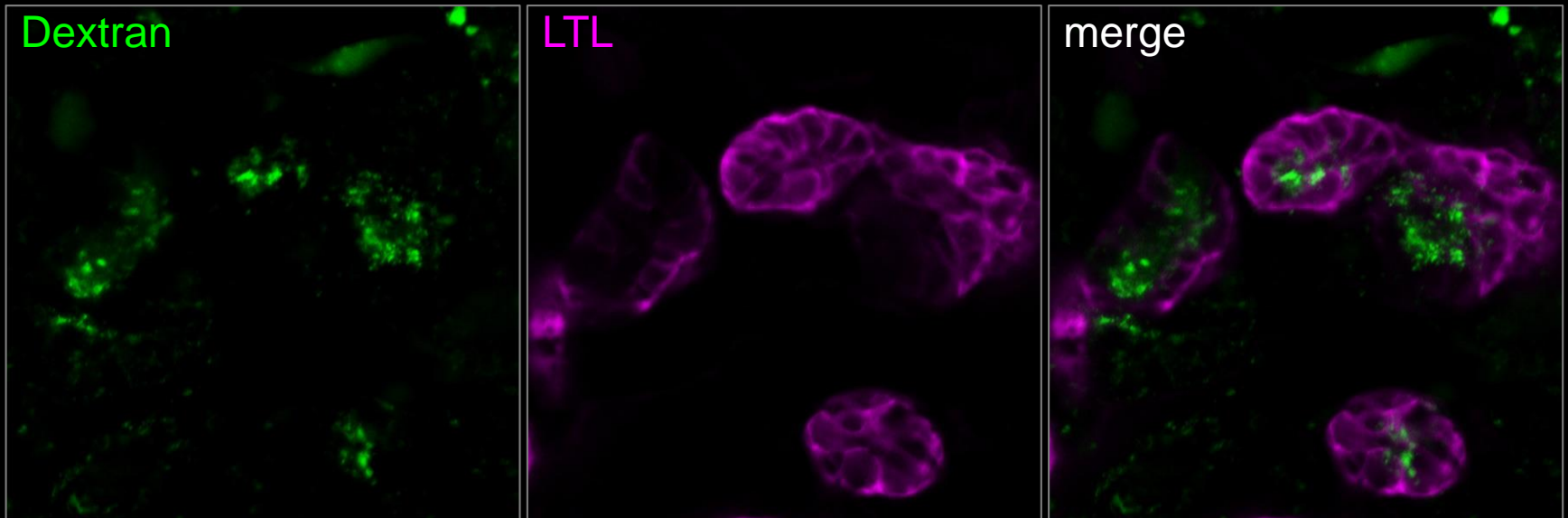


# *Dextran uptake in proximal tubules*

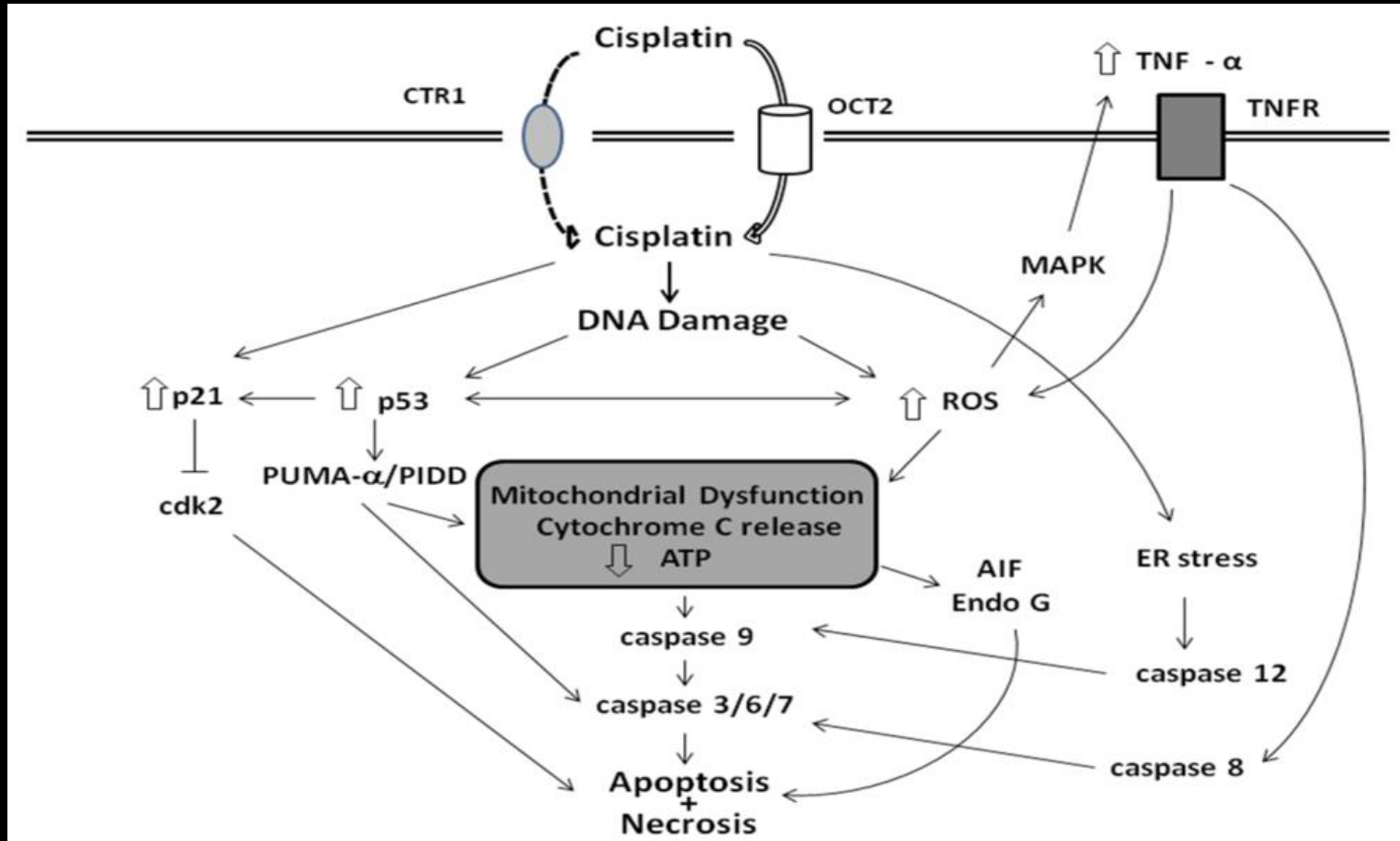
24h Dextran-488



Dextran: a polysaccharide endocytosed by the PT



# Cisplatin induces acute apoptosis in PT



(RP Miller et al, *Toxins* 2010)



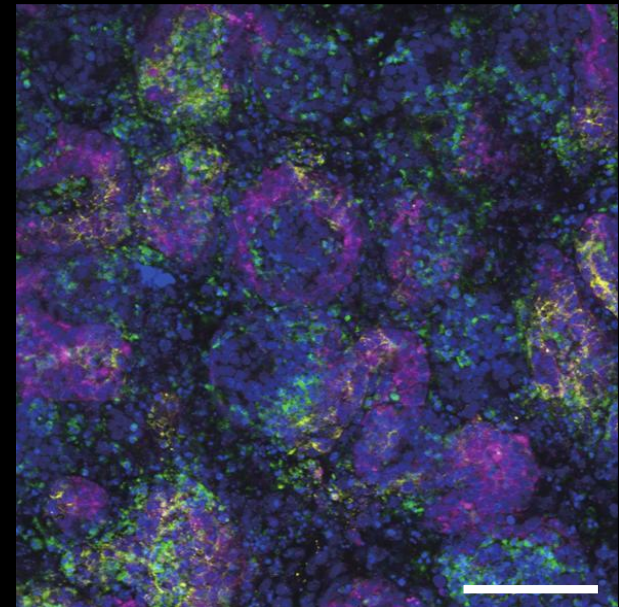
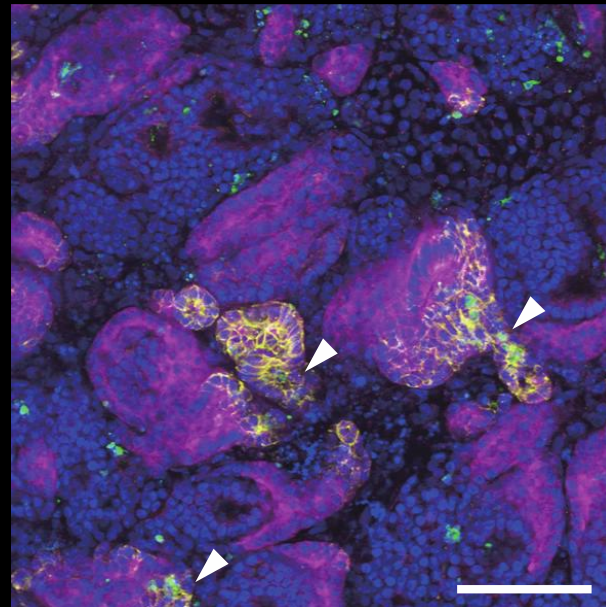
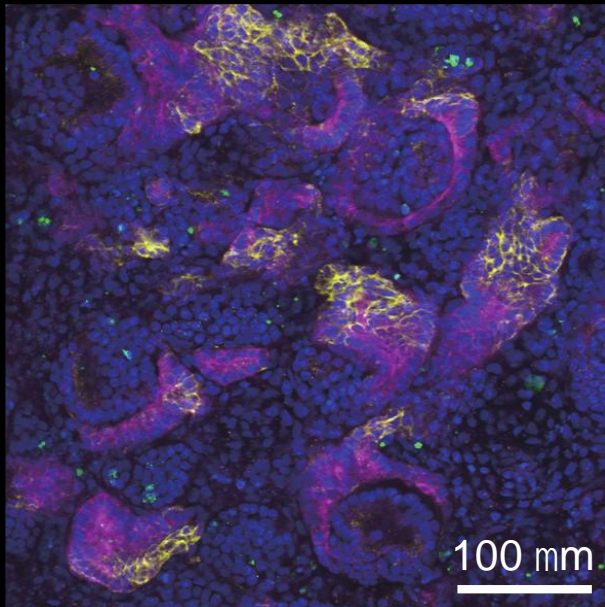
*Low dose Cisplatin induced tissue specific cell death*

24h Cisplatin [ $\mu\text{M}$ ]

0

20

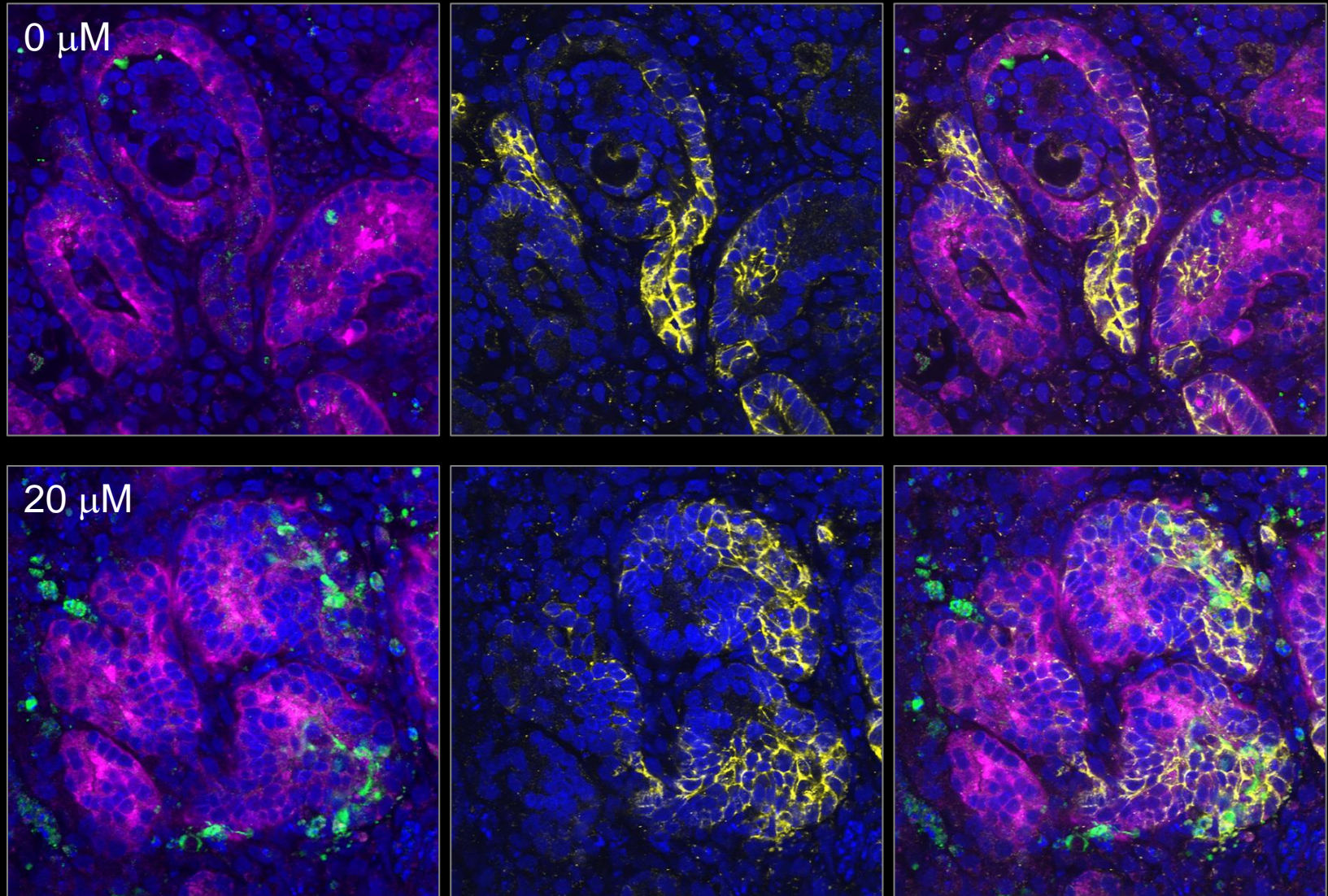
100



LTL ECAD cleaved-CASP3 DAPI



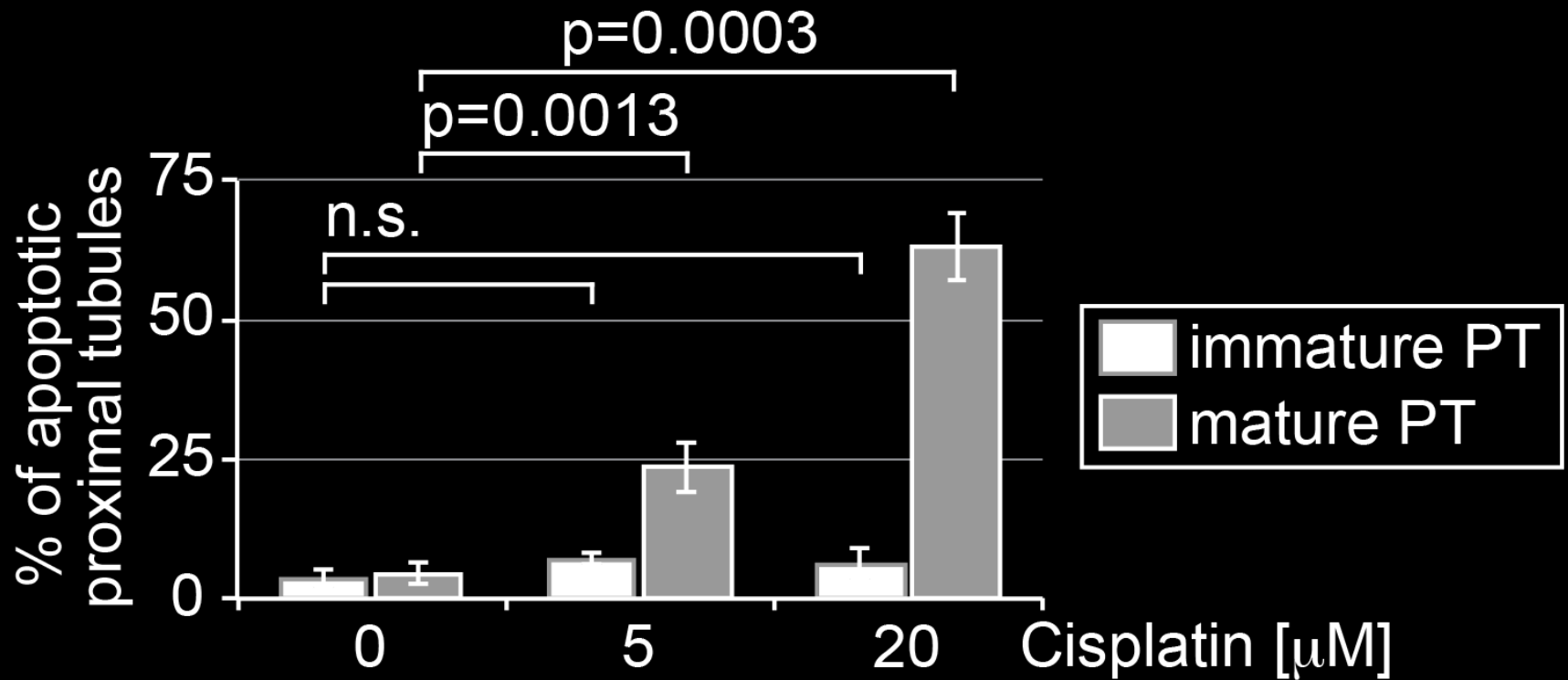
*Low Cisplatin induced matured PT-specific apoptosis*



LTL ECAD cleaved-CASP3 DAPI

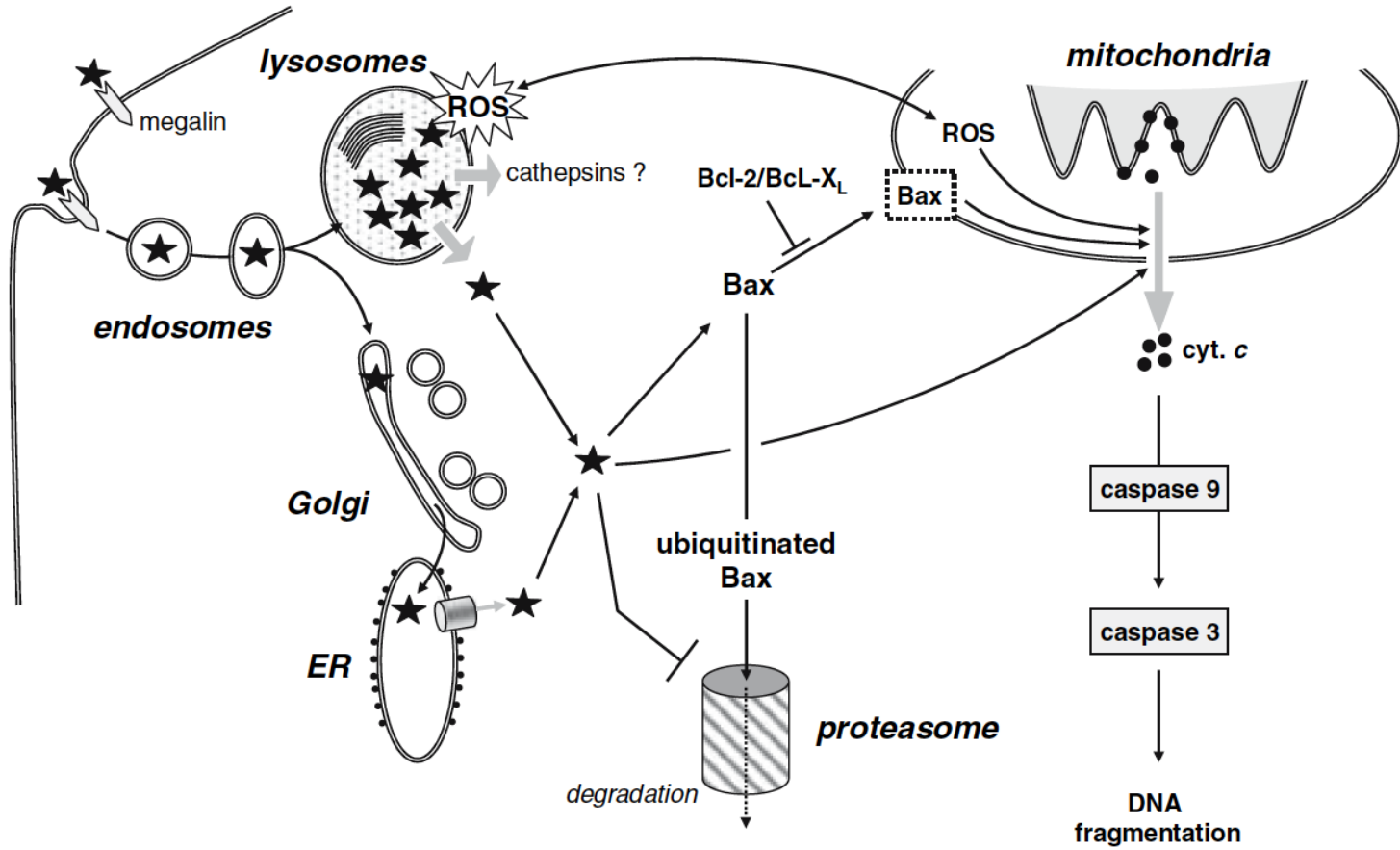


*Low Cisplatin induced matured PT-specific apoptosis*



# Renal tubules uptake gentamicin by endocytosis

## B gentamicin



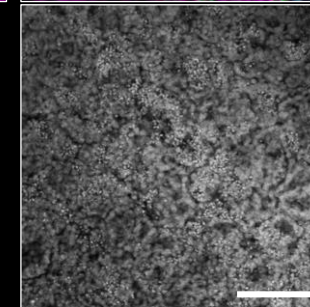
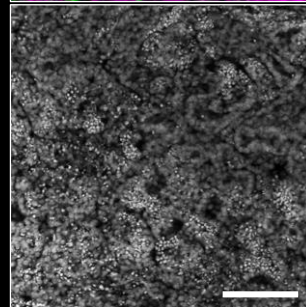
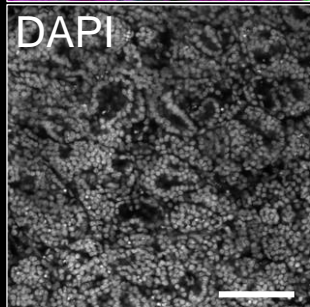
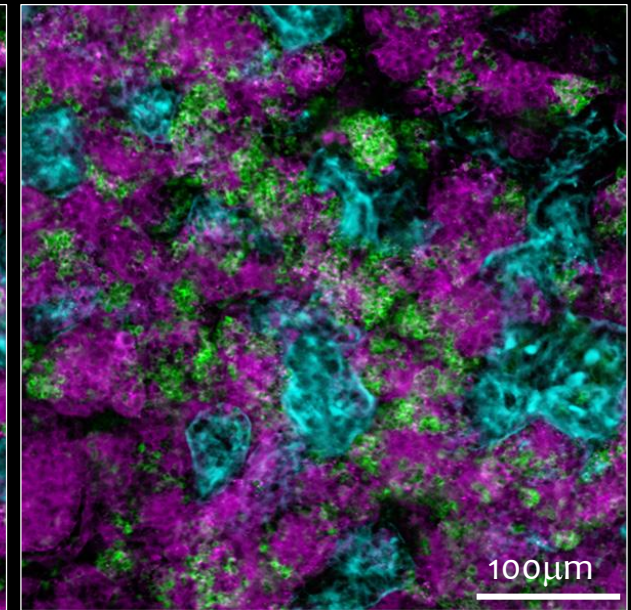
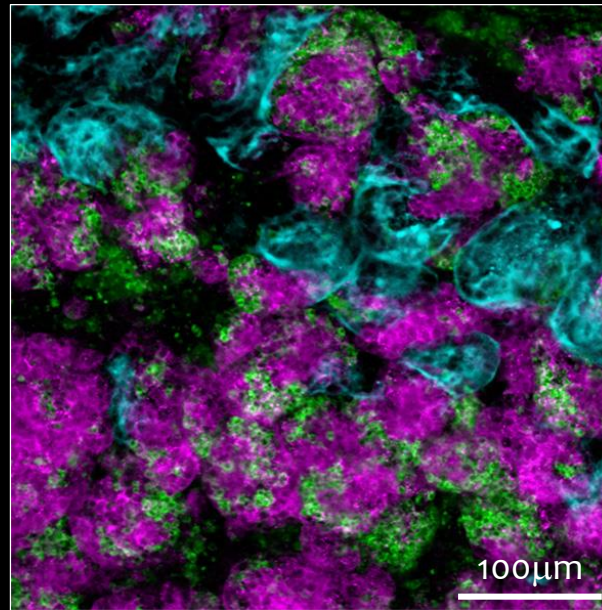
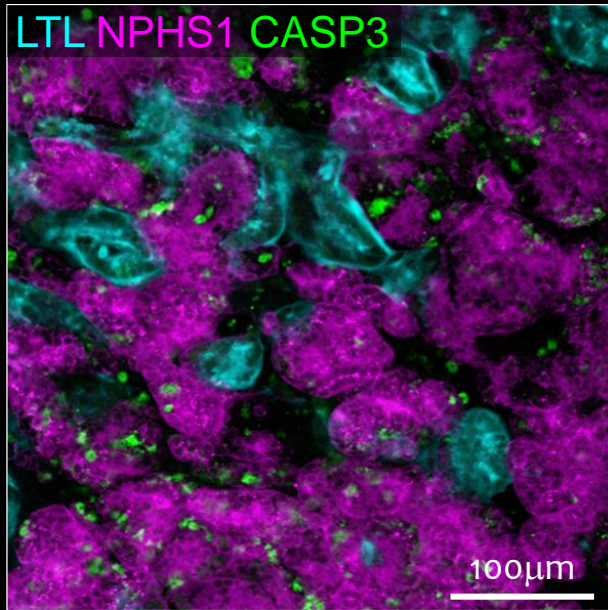
(Servais H. Apoptosis 2008)



# *Gentamicin induced podocyte specific cell death*

0mM Gentamicin 48h

10mM Gentamicin 48h

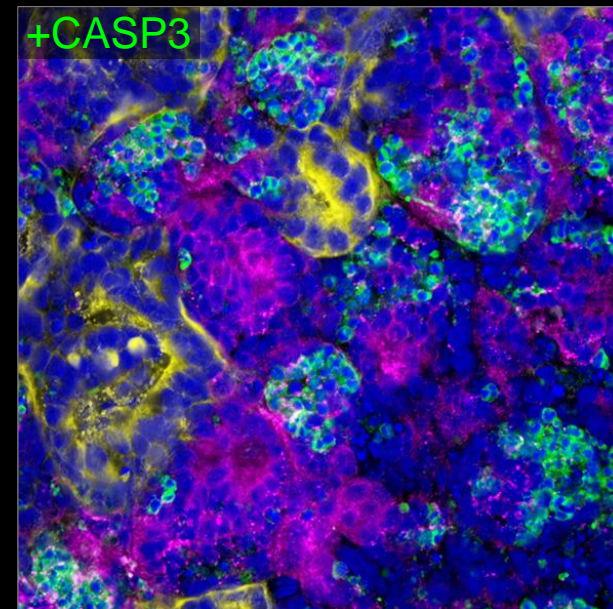
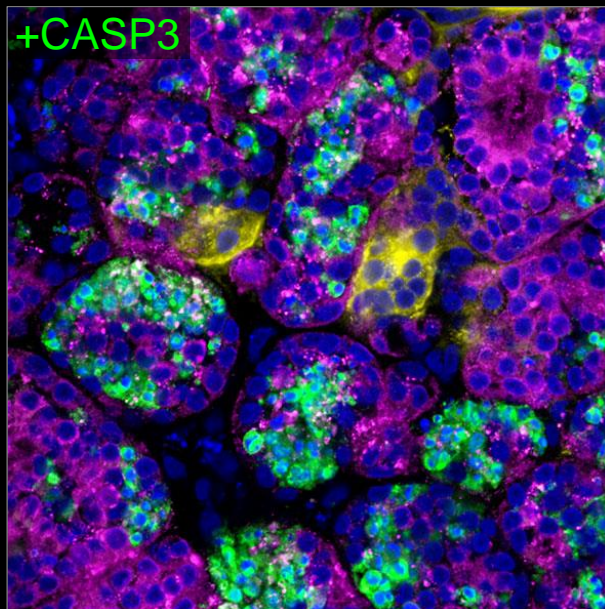
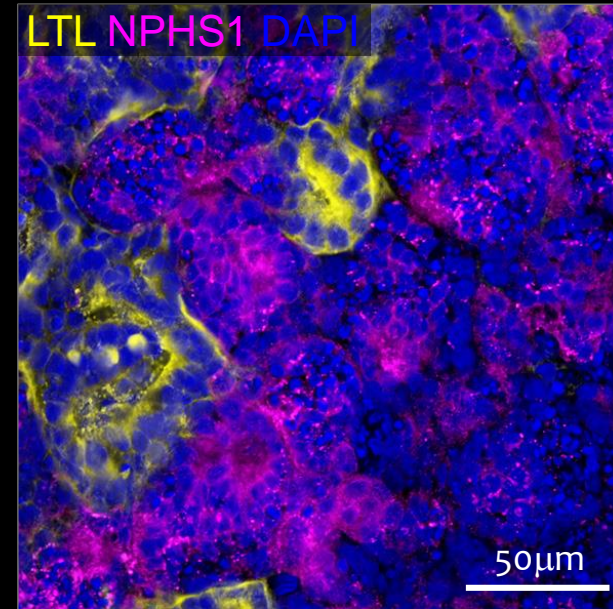
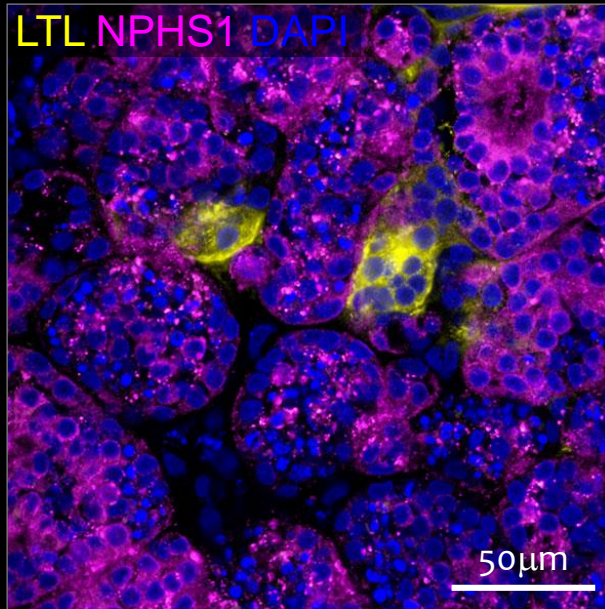




# *Gentamicin induced podocyte specific cell death*

#1

#2

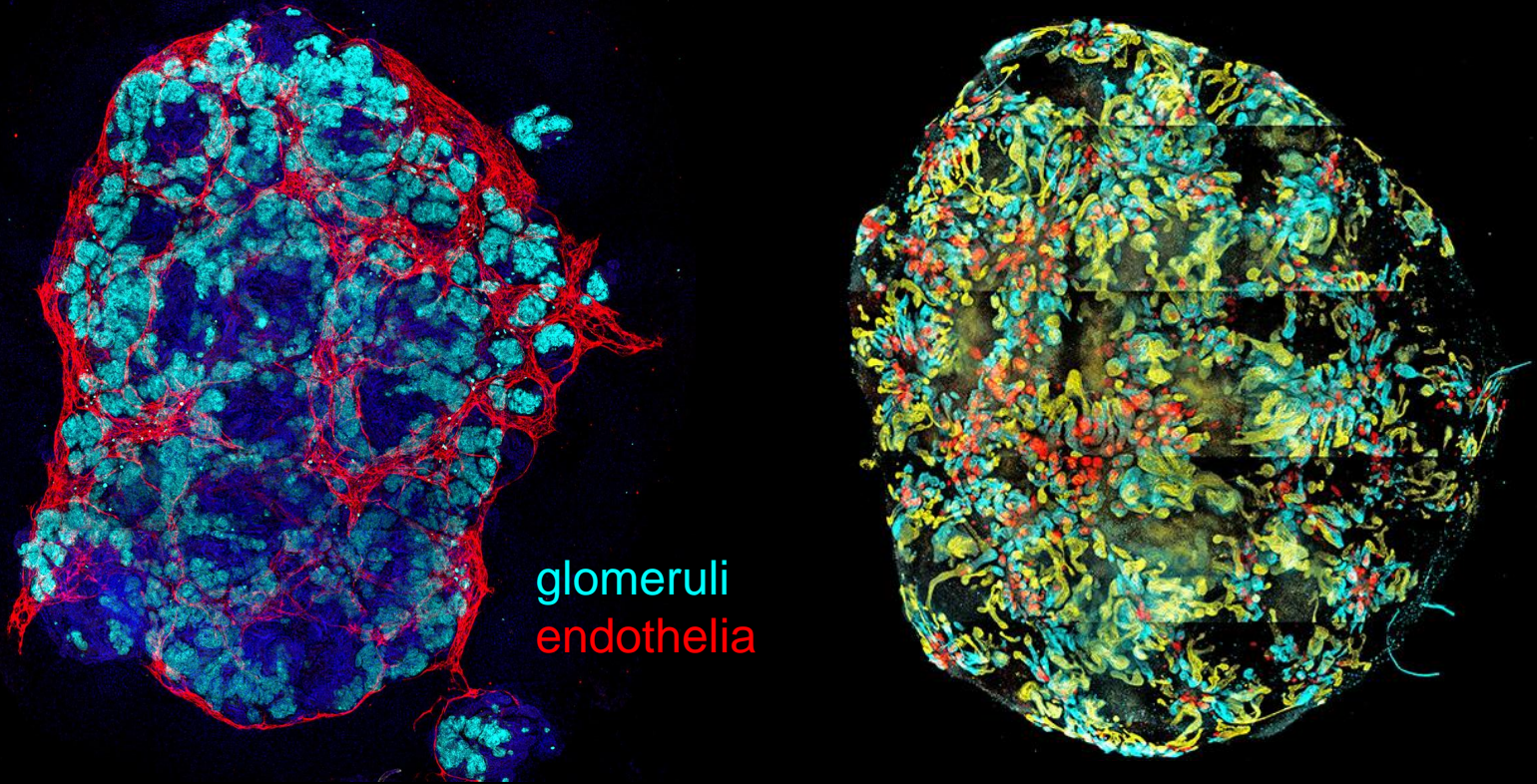


(Unpublished)



# *kidney organoids are not transplantable yet*

1. Glomeruli are not fully vascularized
2. Ureter does not develop in the organoid

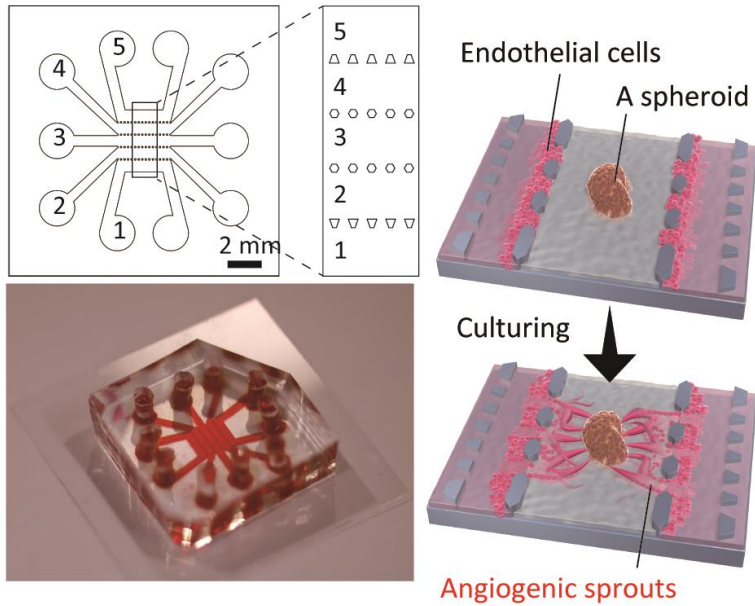


3. Cell heterogeneity is not perfectly controlled.
4. Cells are not well matured

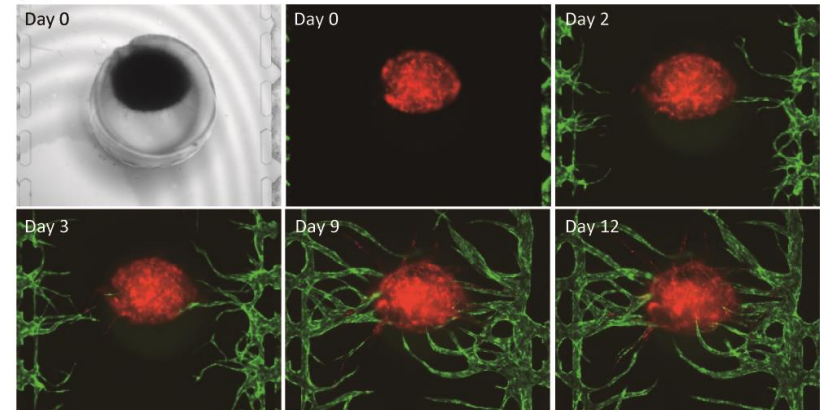
# Organo-on-a-chip for vascularisation of organoids

代表: 横川 隆司、分担: 荒岡 (CiRA)、山下 (CiRA)、高里 (理研)、榎木 (タカラバイオ)

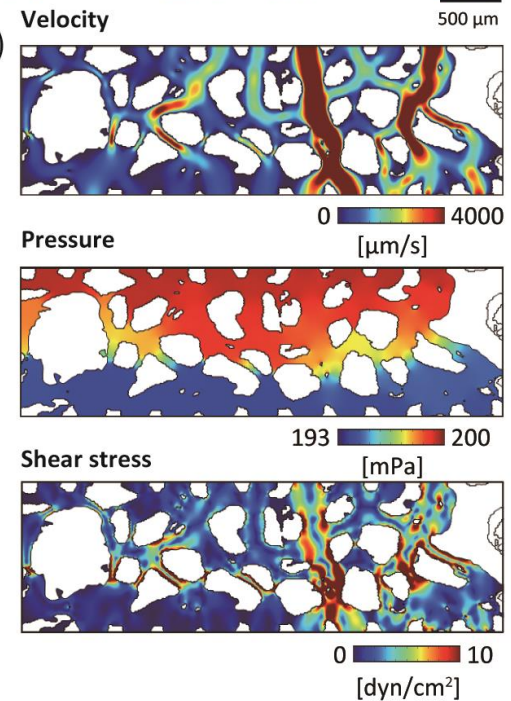
(a)



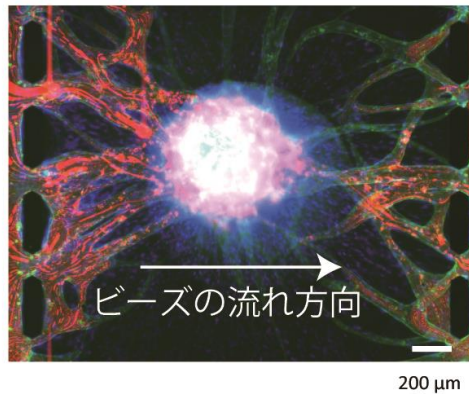
(b)



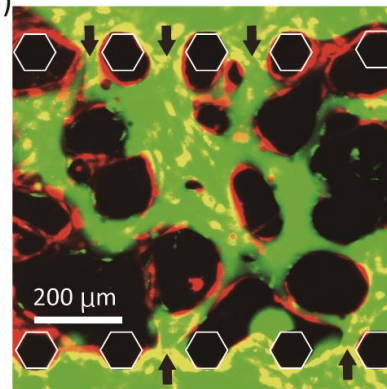
(e)



(c)



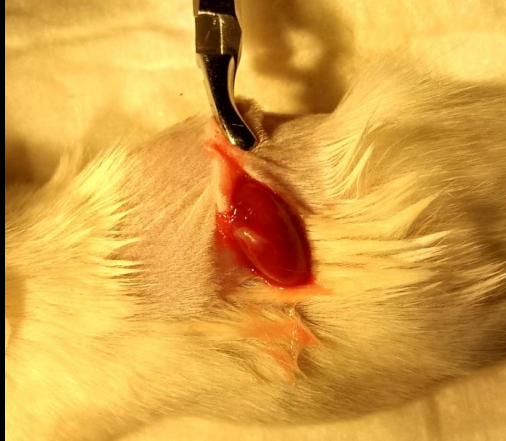
(d)





# Transplanted kidney organoids were vascularized

Transplanted under renal capsule

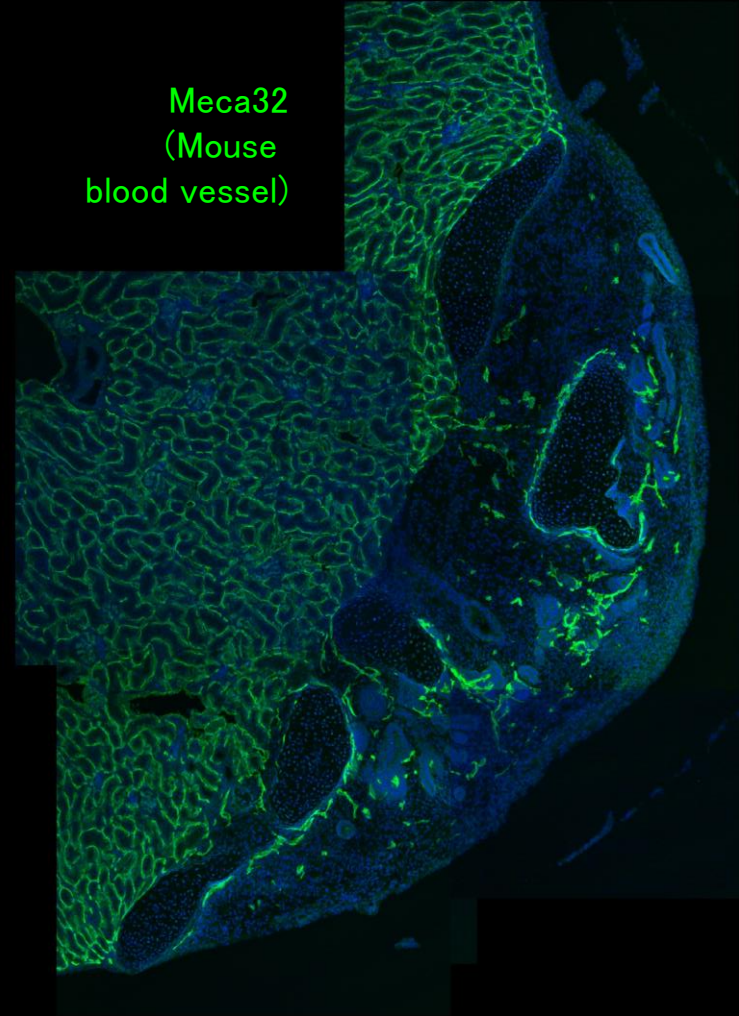


7 days after

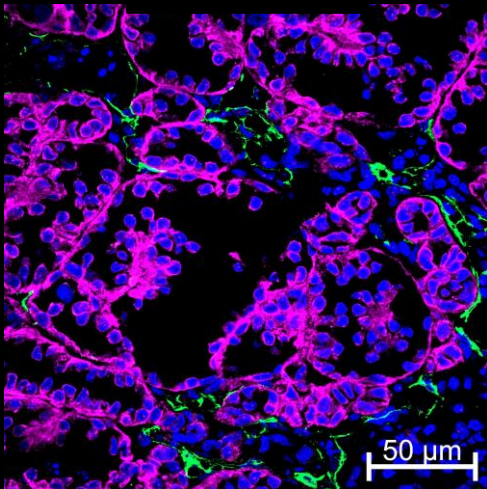


(Stem Cell Reports, 2018)

Meca32  
(Mouse  
blood vessel)

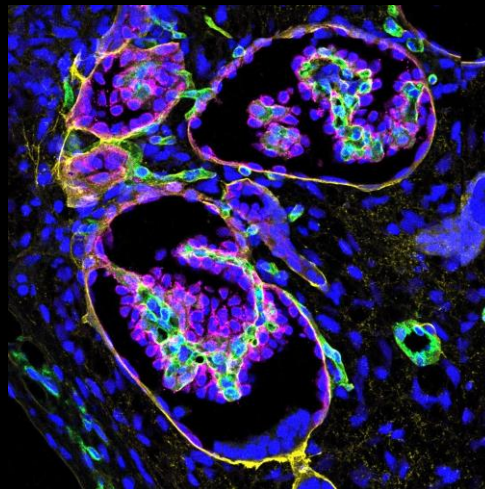


In vitro



Glomerulus Blood vessels

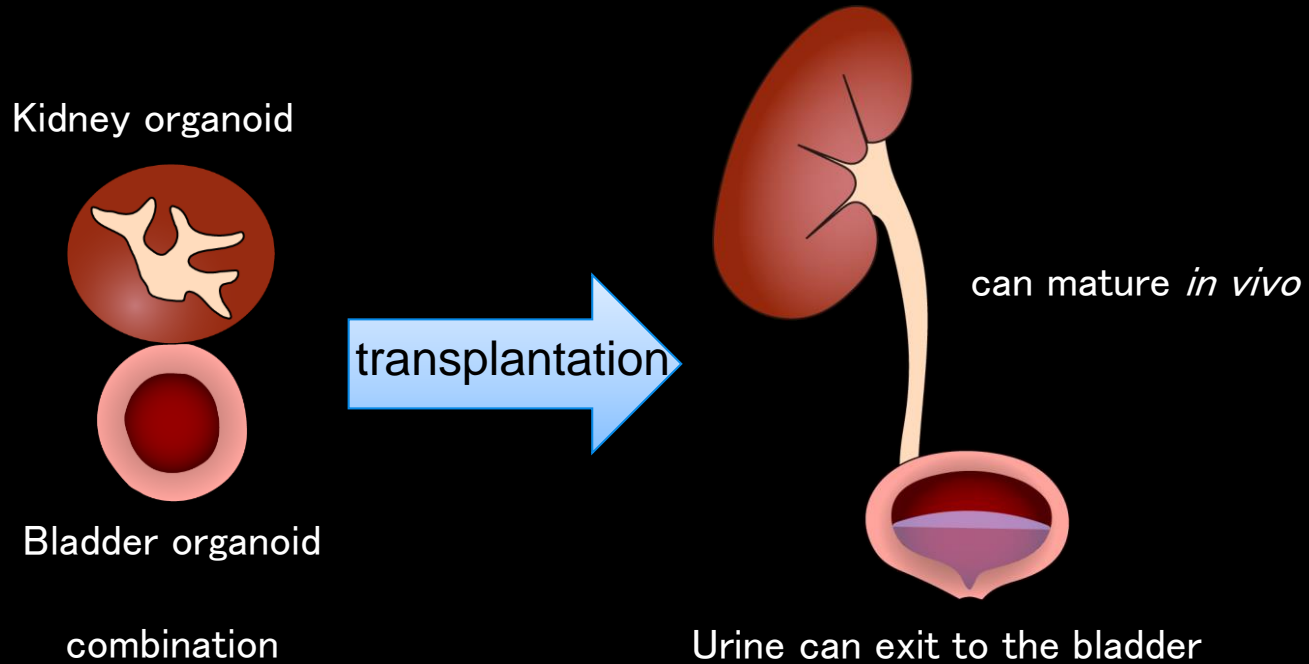
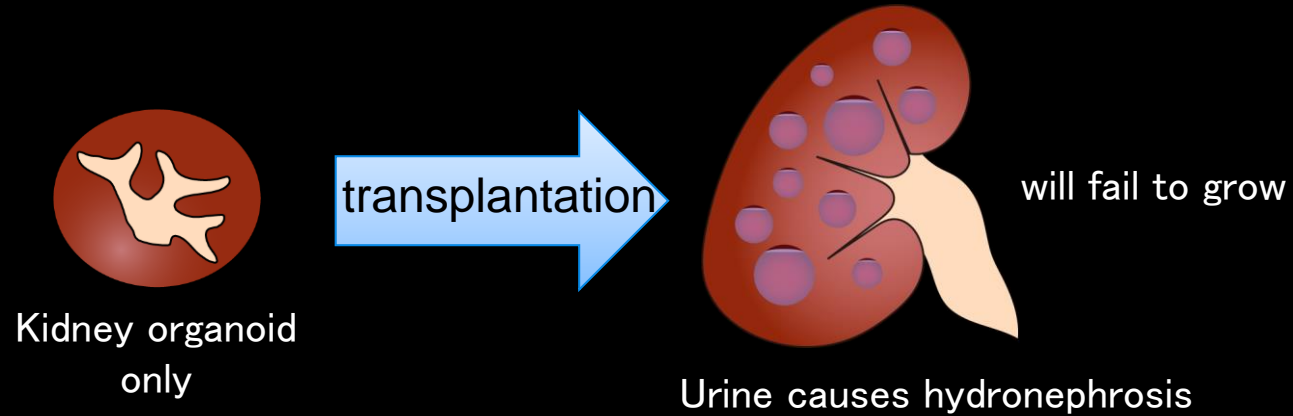
transplantation



(Yabuuchi, Unpublished data)



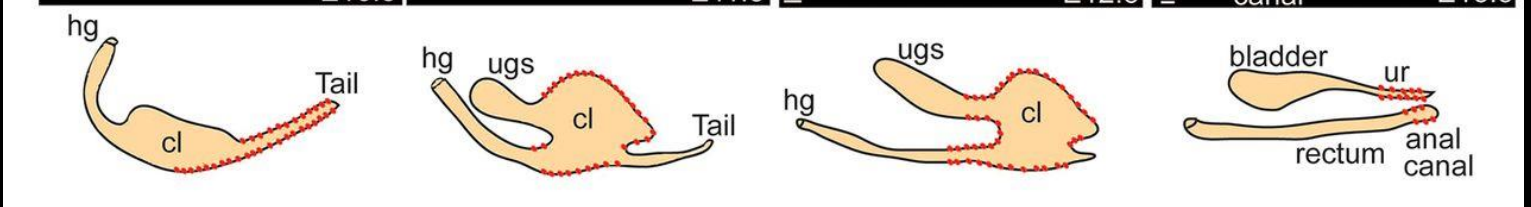
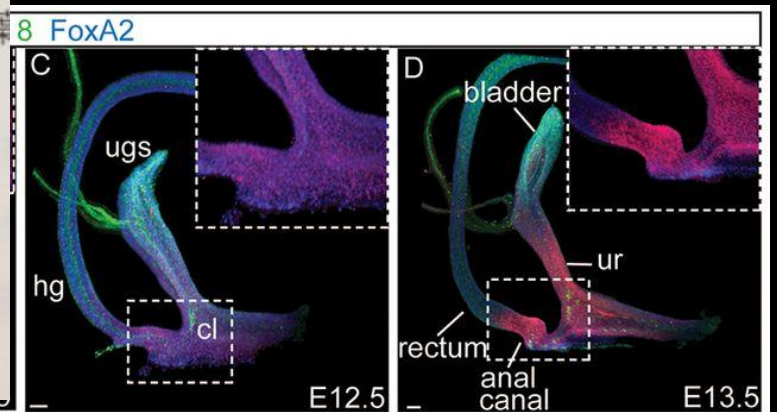
*Kidney organoids can survive with the bladder under transplantation*



# Bladder Development



iPS → endoderm → hind gut  
 → Cloaca → urothelium



# 謝辞

## *Takasato Lab RIKEN BDR*

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Yoshiki Sahara (D3)

Kazuhiro Ofuji (D3)

Rio Noto (D4)

Wataru Uno (D3)

Masaya Goto (M2)

Hiroki Matsunaga (OB)

Thomas Kluiver (visiting stu.)



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*Leiden University Medical Center*

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