

# Glyco-engineering for Glycoprotein Production and Development of a Visualization Tool for Glycan Metabolic Pathways

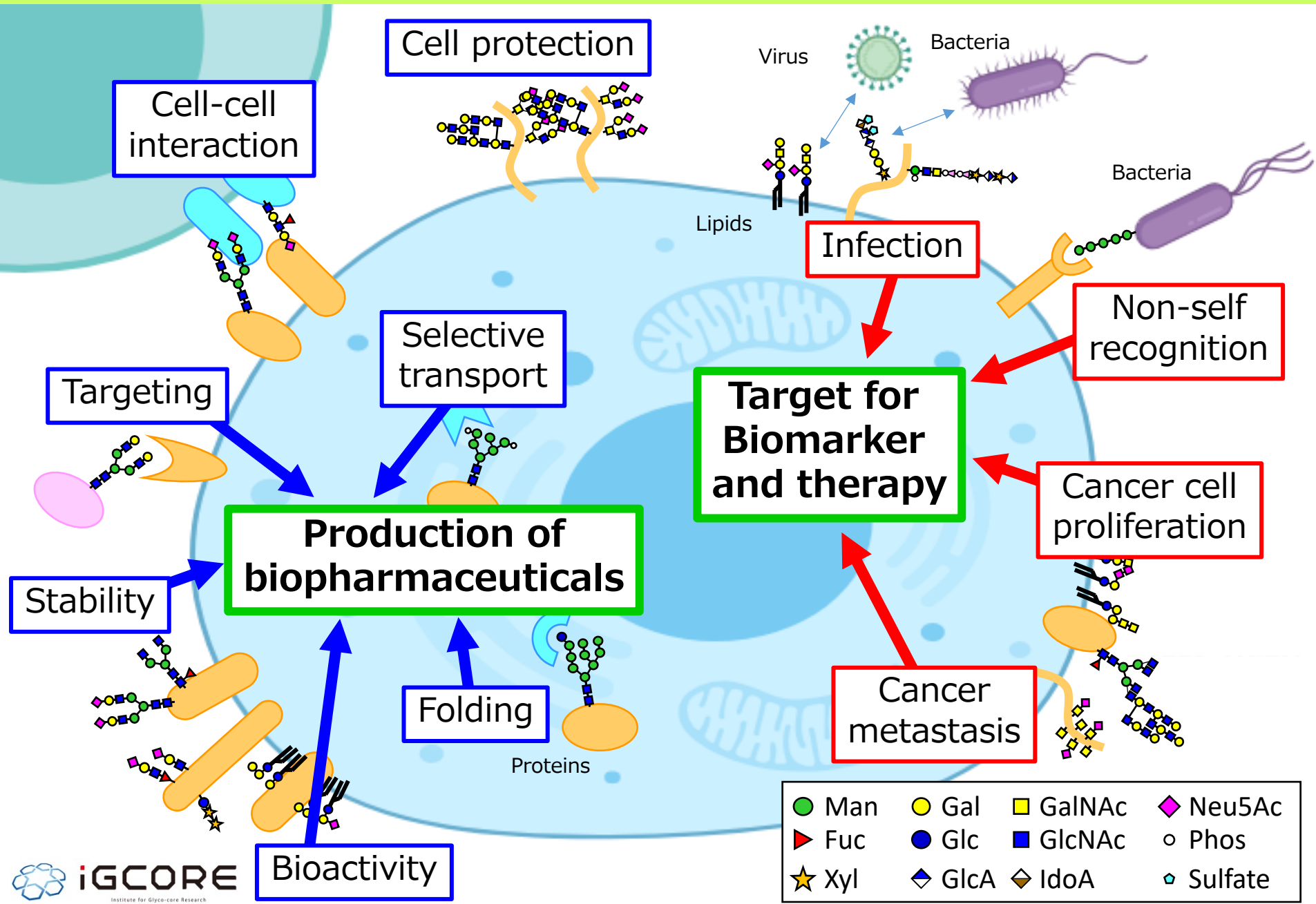
Glycoforum

Morihisa Fujita

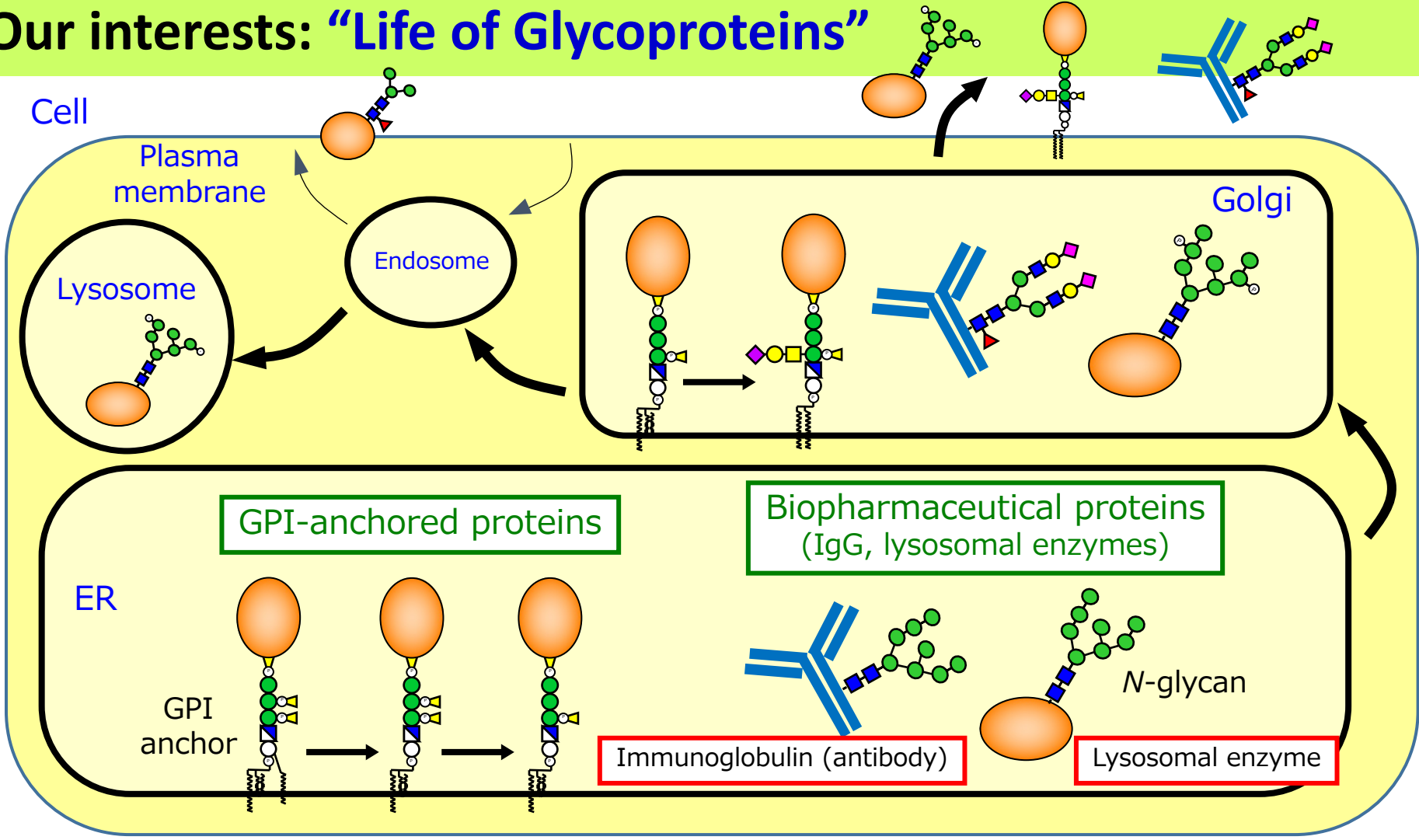
Institute for Glyco-core Research (iGCORE)  
Gifu University



# Glycan functions and importance



# Our interests: "Life of Glycoproteins"



Regulation of biosynthesis and transport of glycoproteins

Engineering of mammalian cells to produce biopharmaceuticals

# Contents

## **Simplification of N-glycan structures in HEK293:**

Production of recombinant proteins

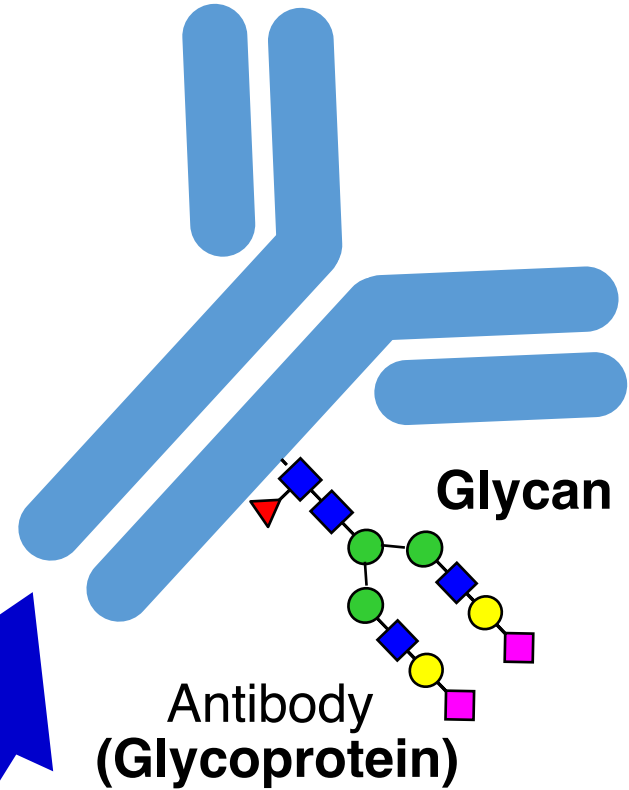
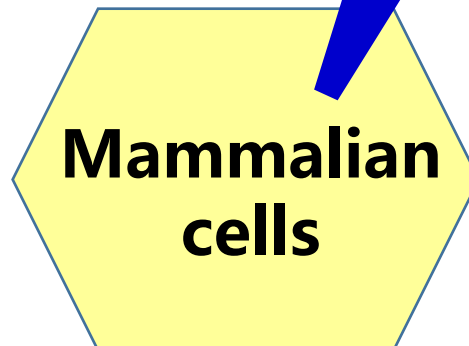
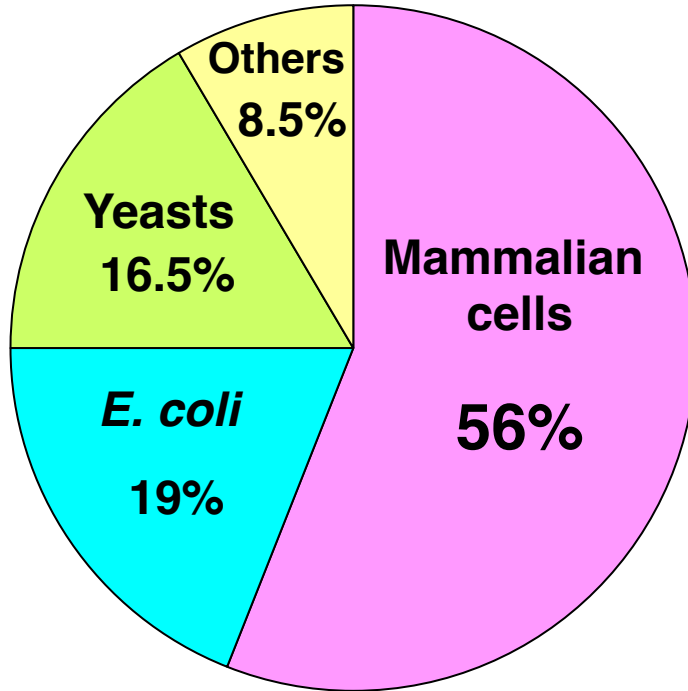
only having high-mannose type N-glycans

## **Development of a visualization tool for glycan metabolic pathways based on gene expression profiles:**

Glyco-engineering and Glyco-comparison using GlycoMaple

# Biopharmaceutical proteins

## Biopharmaceutical proteins



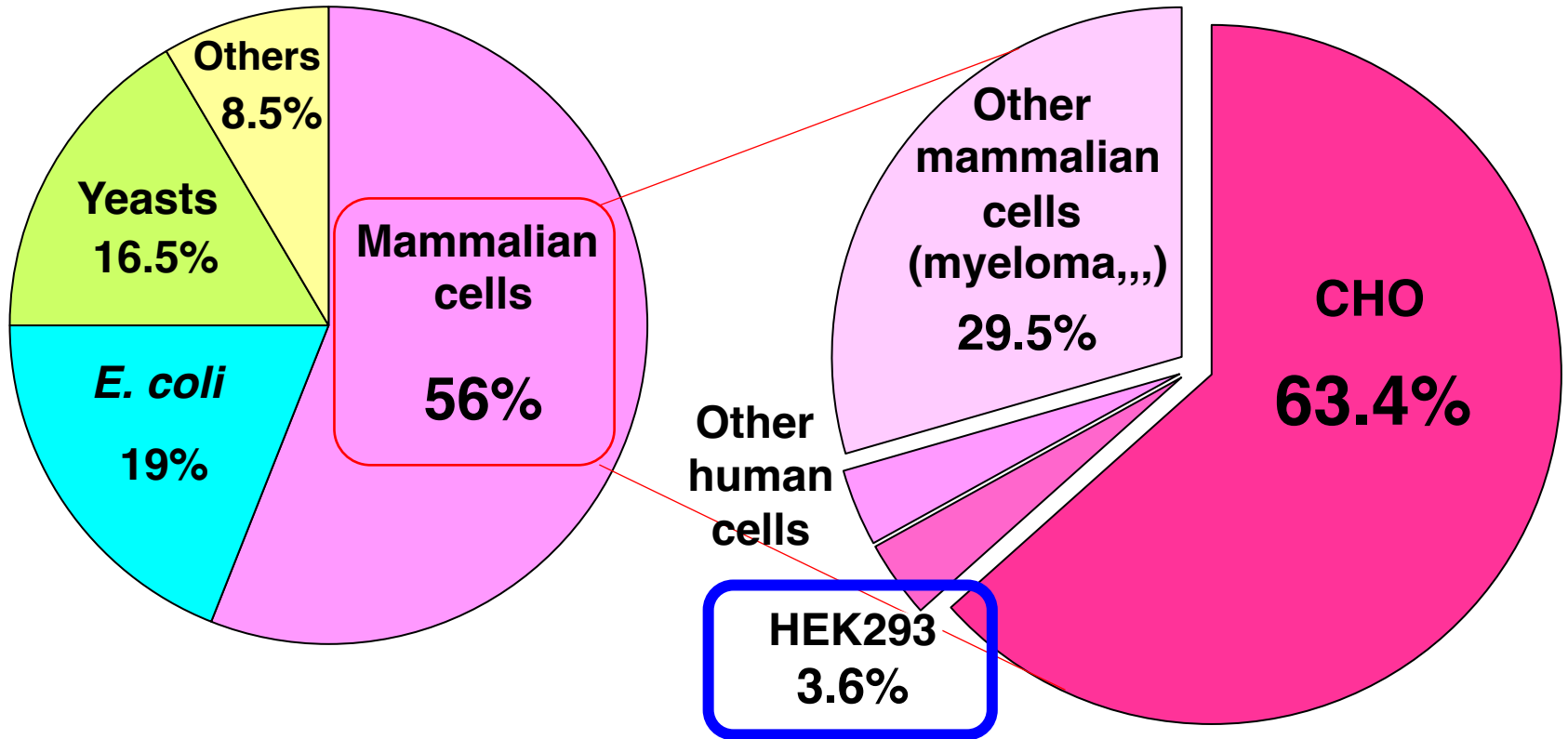
Protein folding and assembly  
Post-translational modifications

**Protein glycosylation**

**Stability**  
**Bioactivity**  
**Immunogenicity**

Modified from  
Walsh G (2014)  
*Nat. Biotech.* 12: 992

# Biopharmaceutical proteins produced by mammalian cells



# HEK293

## HEK293 (human embryonic kidney 293)

Established in 1977 by Frank Graham (McMaster Univ., Canada)

**Easy to handle** and **widely used** in basic and applied researches

Adaptable to **serum-free suspension** cultures

Used for production of some pharmaceuticals (ex. factor VIII, factor IX)

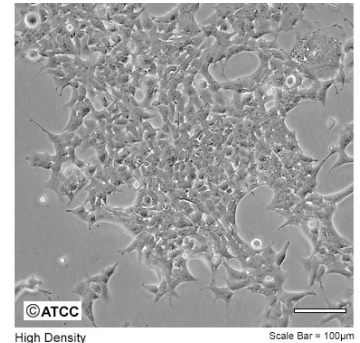
Used for viruses or virus-like particles (**VLPs**) production

Less competitive for host development, compared to CHO cells

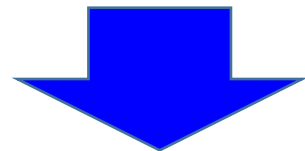
**Accumulated genetic information** in human databases

### Glycosylation in HEK293

**Human glycoform**



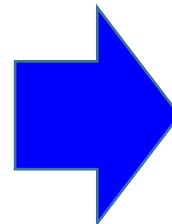
HEK293



Drawbacks of protein production  
in mammalian cells

**Heterogeneity of glycans**

**Lack of  
functional / suitable glycans**



### Glyco-engineering

- 1) Customization
- 2) Simplification

# Engineering of mammalian cells to produce proteins with homogenous glycans

One issue for recombinant protein production in mammalian cells

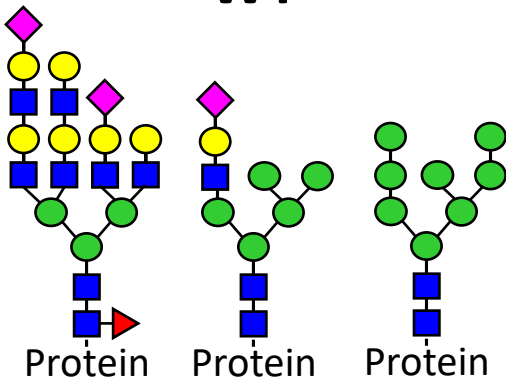
Heterogeneity of glycans on proteins

Heterogeneous glycan

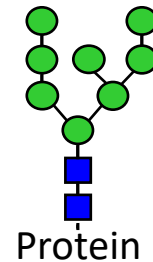
Genome-editing using  
CRISPR/Cas9 system

Uniform glycan

WT



Only **high Man-type**  
No Complex-type

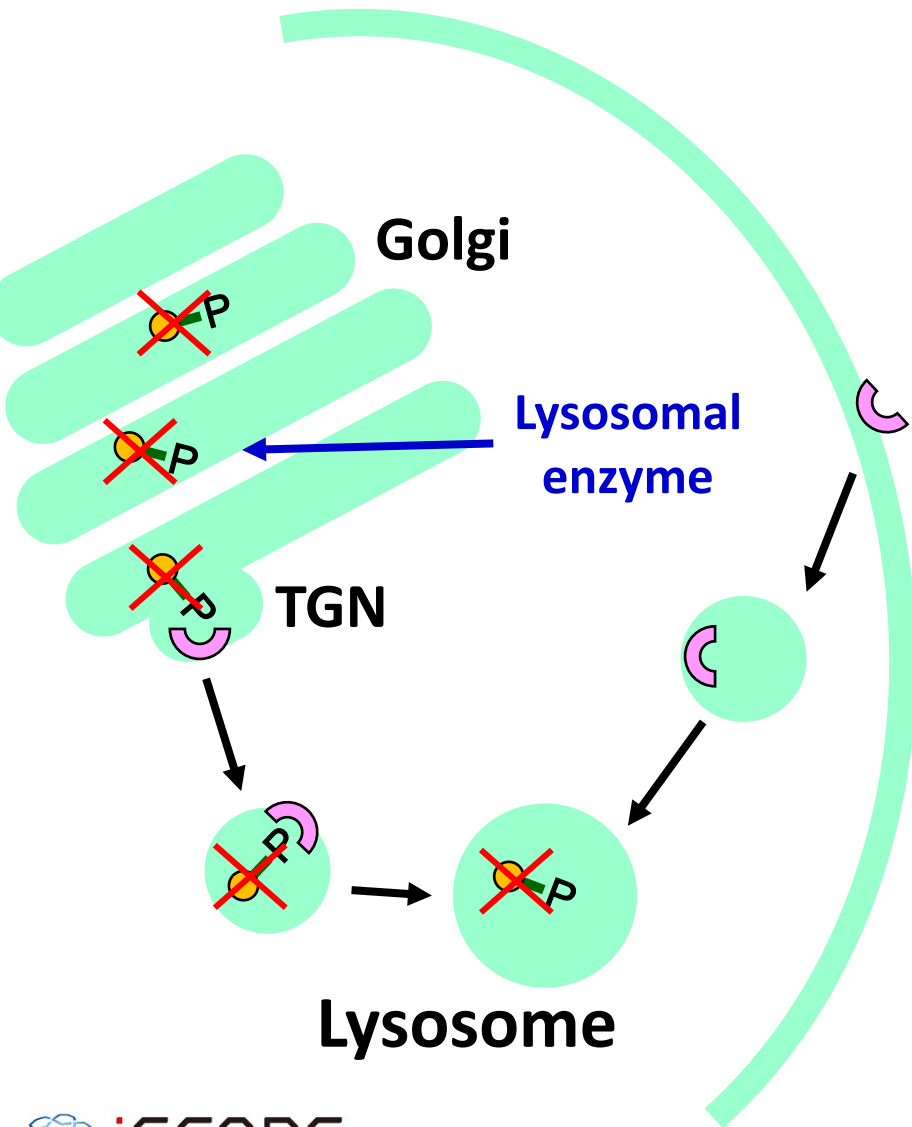


**High Man-type**  
N-glycan with **Man8** or **Man9**

Drug for Lysosomal storage diseases  
(Recombinant lysosomal enzymes)  
Vaccine candidate for HIV-1 (gp120)



# Enzyme replacement therapy for lysosomal diseases



## Lysosomes

Garbage disposals in the cell

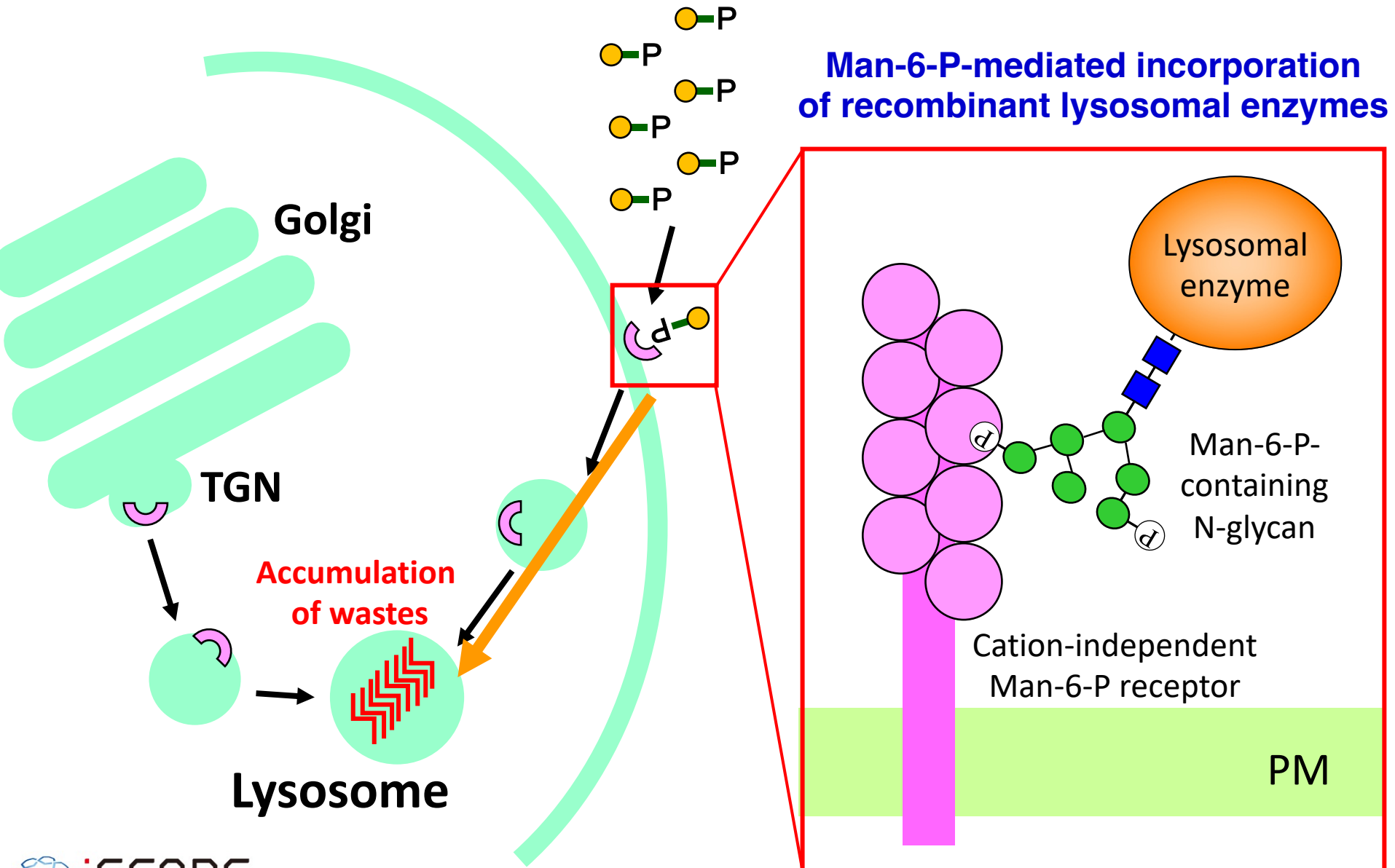
> 50 Lysosomal enzymes to hydrolyze substrates (nucleic acids, proteins, lipids, glycans)

Defects in lysosomal enzyme genes cause lysosomal storage diseases

## Enzyme replacement therapy

Recombinant lysosomal enzymes are intravenously injected

# Enzyme replacement therapy for lysosomal diseases



**Man-6-P-mediated incorporation of recombinant lysosomal enzymes**

Lysosomal enzyme

Man-6-P-containing N-glycan

Cation-independent Man-6-P receptor

PM

# Broadly neutralizing antibodies for HIV-1

broadly neutralizing antibodies (bnAbs) isolated from HIV-1 patients

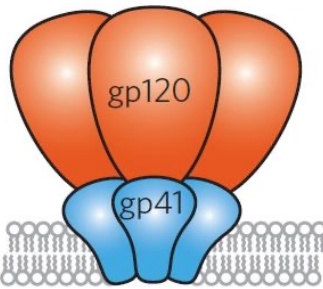
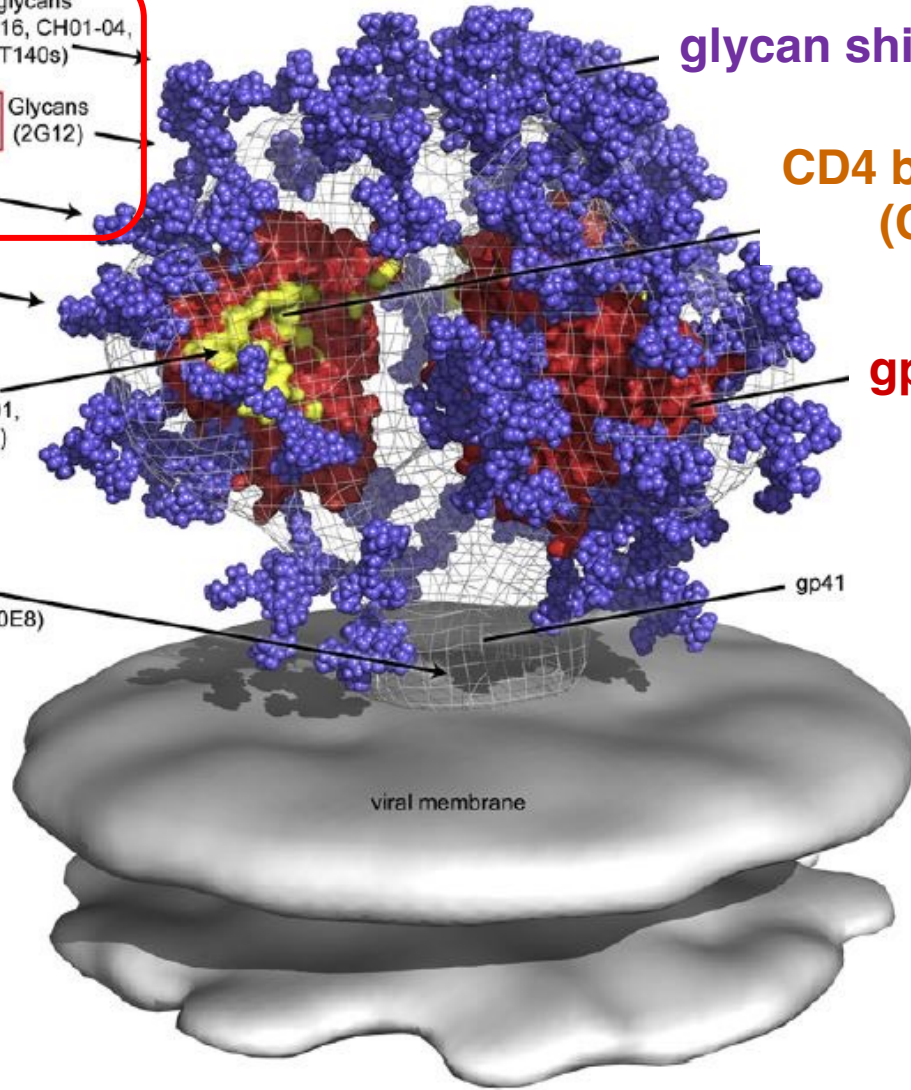
## Binding to N-glycans

- >12 MAbs V2/glycans (PG9, PG16, CH01-04, PGT140s)
- >25 V3/V4/glycans (cluster of targets: PGT120s, PGT130s)
- 1 Glycans (2G12)

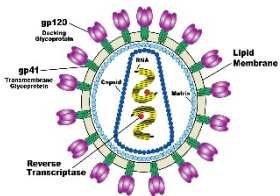
2 V3/CD4i (3BC176)

>25 CD4bs (b12, VRC01, NIH 45-46)

>5 MPER (2F5, 4E10, 10E8)

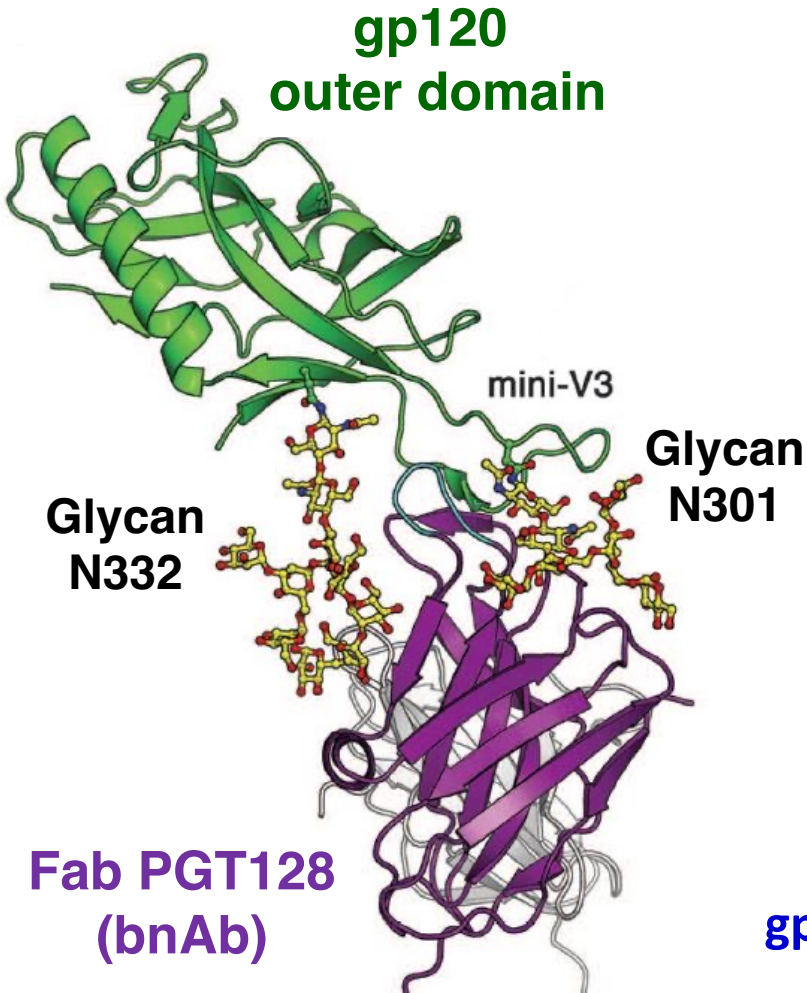


Functional mature Env

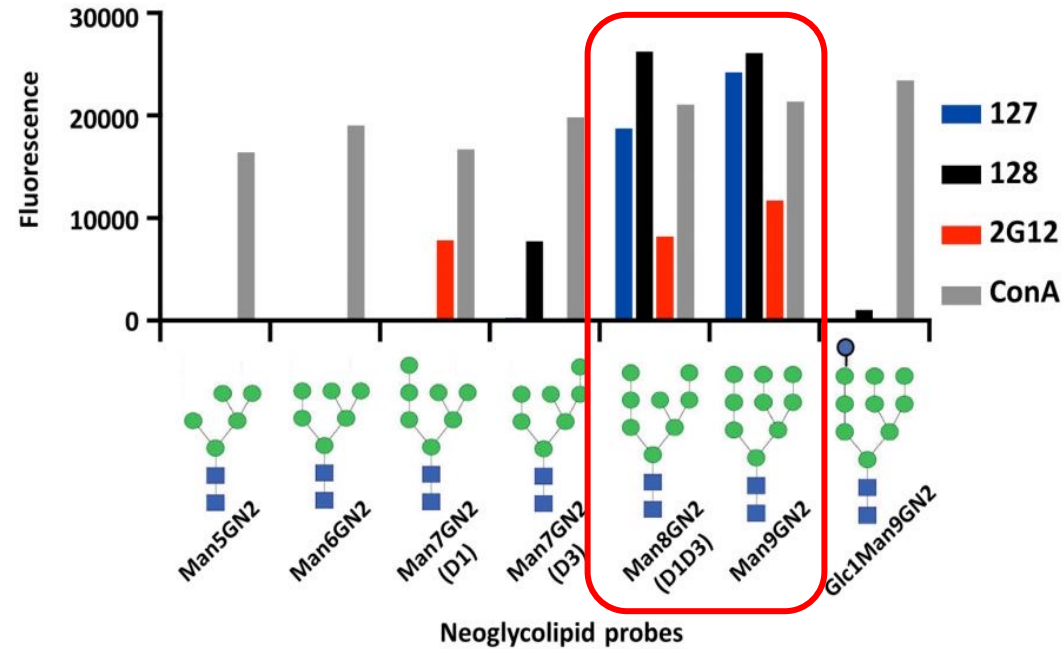


HIV-1

# Many bnAbs recognize high-Man type N-glycans on gp120

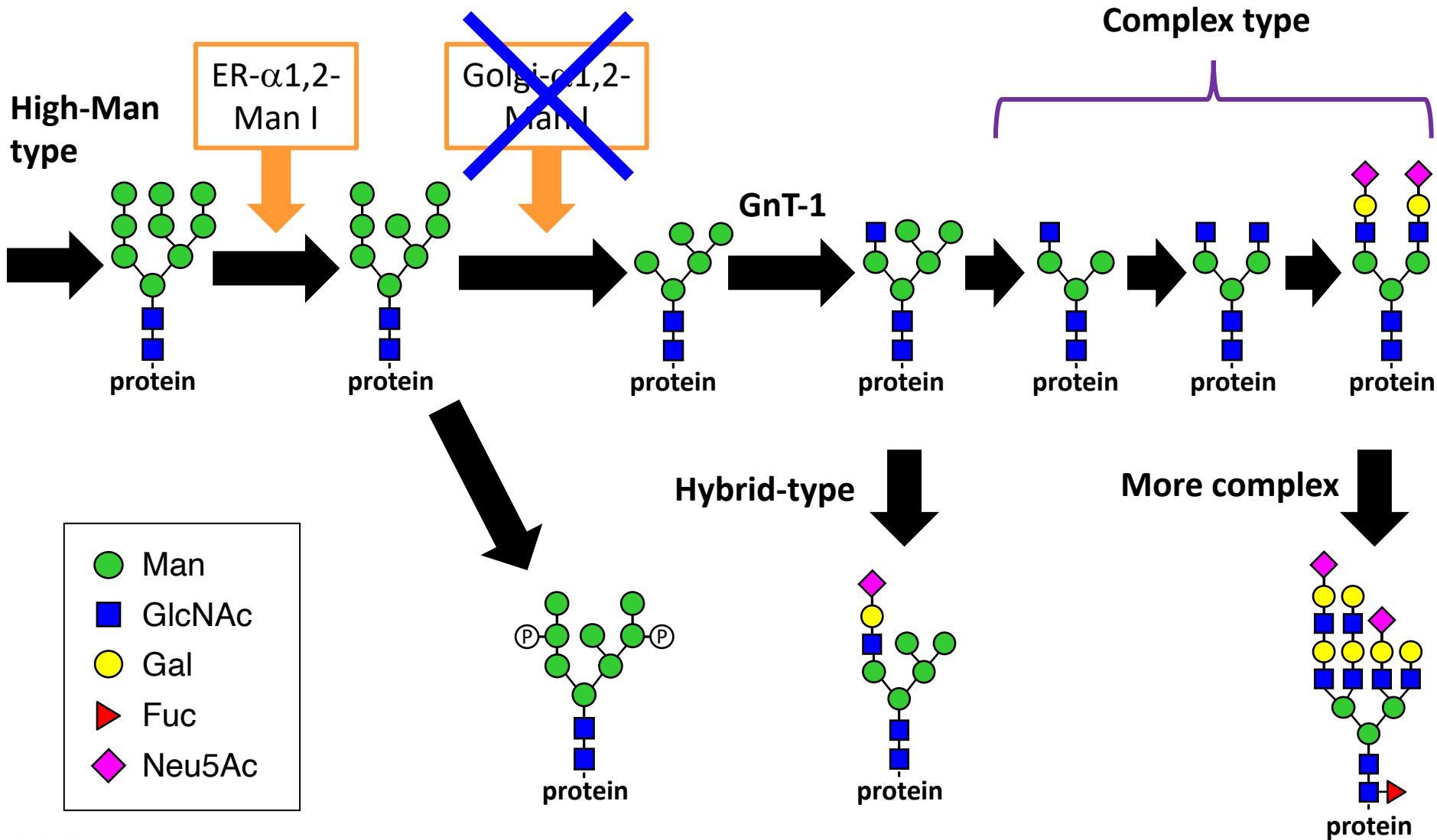


## Binding of bnAbs to N-glycan structures



**gp120 with M8 or M9 high-Man-type N-glycans is a candidate for the vaccine against HIV-1.**

# Disruption of Golgi-Man-I to express proteins containing high Man-type N-glycans



# Human GH47 ( $\alpha$ 1,2-mannosidases) in CAZy database



Golgi Man-I A  
(MAN1A1)

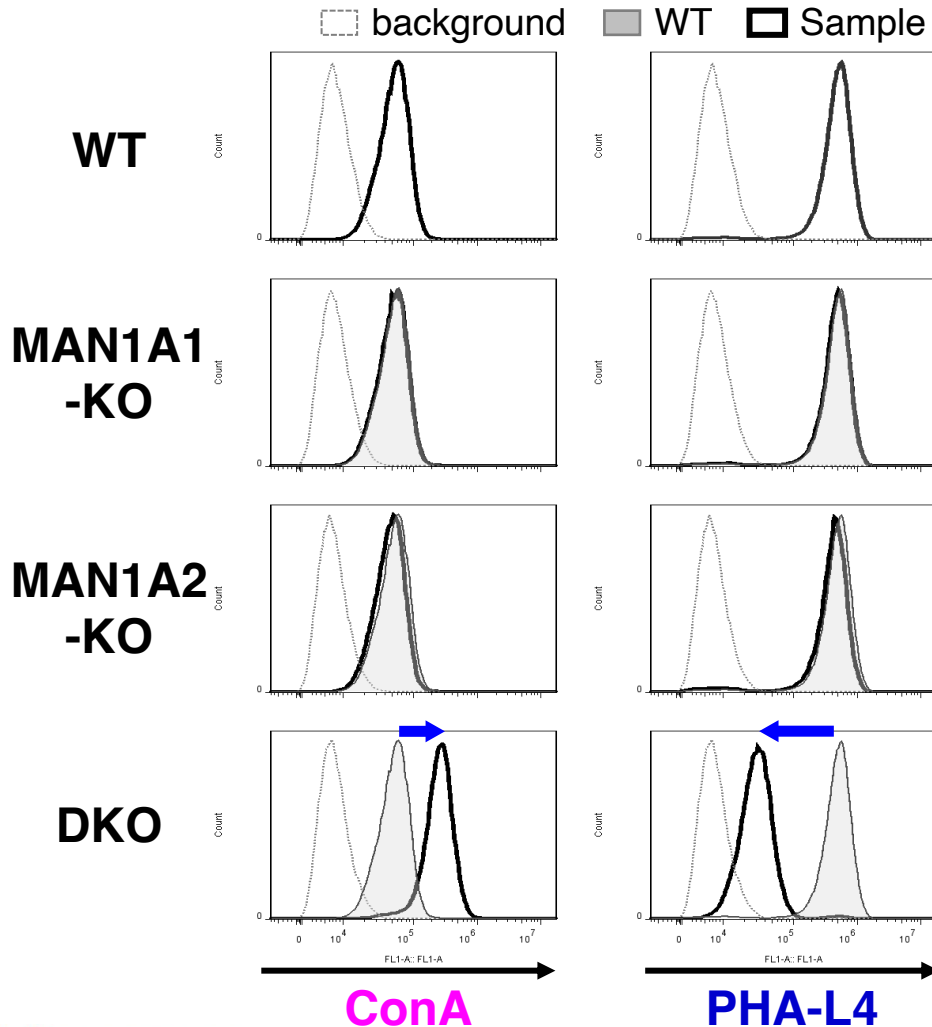
## Human GH47 ( $\alpha$ 1,2-mannosidases)

Protein name	Gene name	TPM	Function
Golgi Man-I A	<i>MAN1A1</i>	34.82	Glycan maturation
B	<i>MAN1A2</i>	25.65	
C	<i>MAN1C1</i>	5.24	
ER Man-I	<i>MAN1B1</i>	64.97	Protein quality control ?
EDEMs	<i>EDEM1</i>	28.91	ER-associated degradation
	<i>EDEM2</i>	24.59	
	<i>EDEM3</i>	24.01	

Tempel et al (2004) *J. Biol. Chem.*

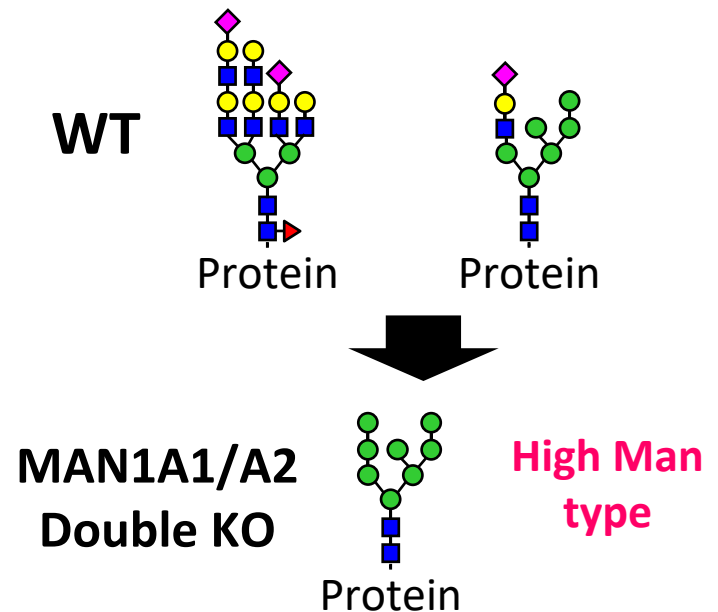
# MAN1A1 & MAN1A2 double KO cells decreased complex-type and increased high-Man-type glycans

Flow cytometric analysis of glycans on the cell surface using two lectins



**PHA-L4**: lectin binding to complex type of glycans

**ConA**: lectin binding to high Man type of glycans



# KO of other $\alpha$ 1,2-mannosidase genes in Double-KO

Human GH47 ( $\alpha$ 1,2-mannosidases)

Protein name      Gene name

Golgi Man-I A      ~~MAN1A1~~

B      ~~MAN1A2~~

C      MAN1C1

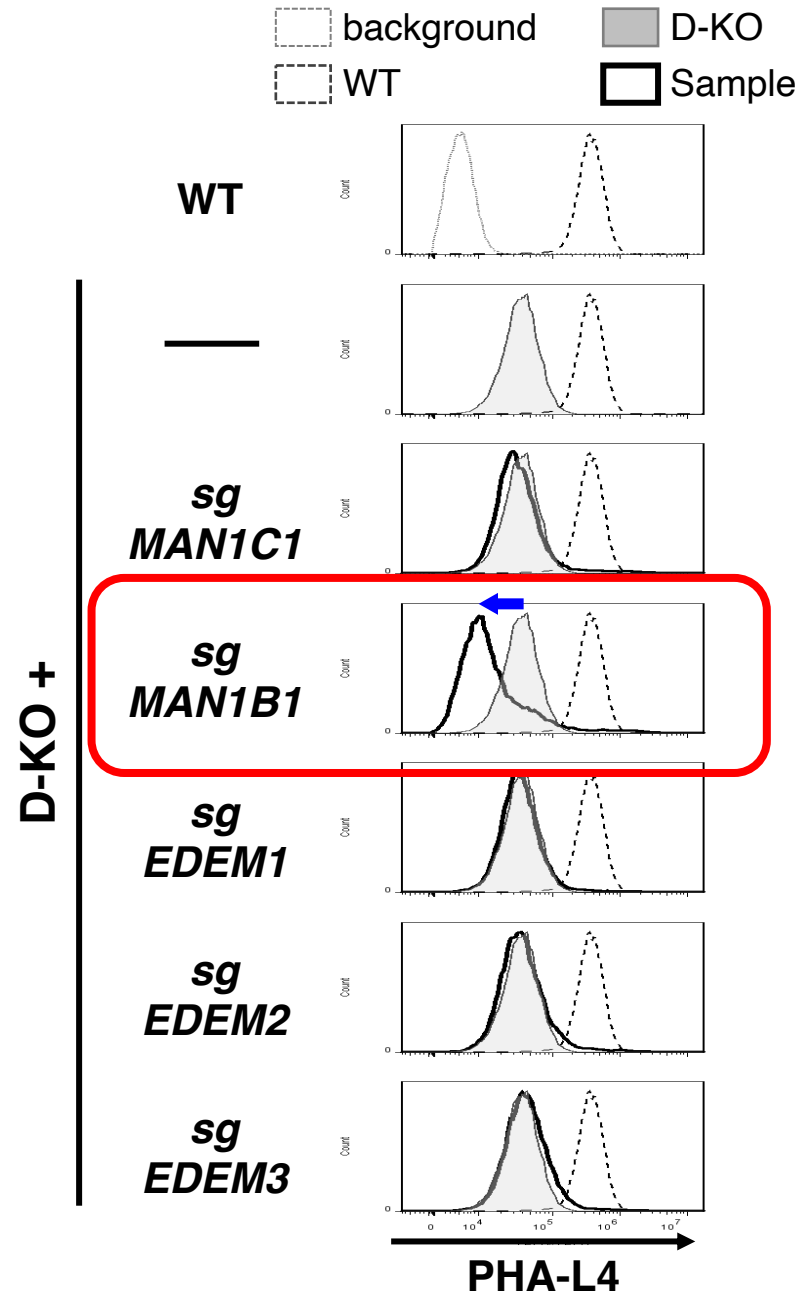
ER Man-I      MAN1B1

EDEMs      EDEM1

EDEM2

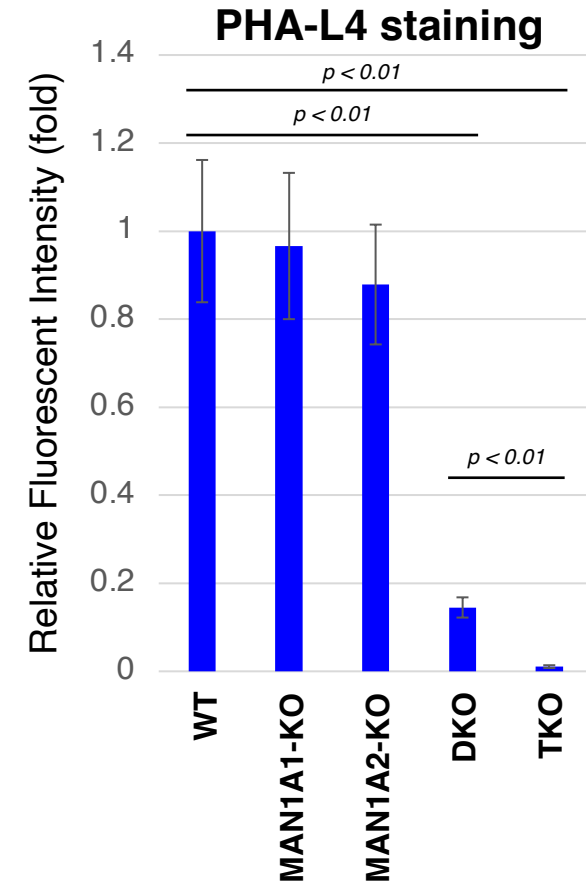
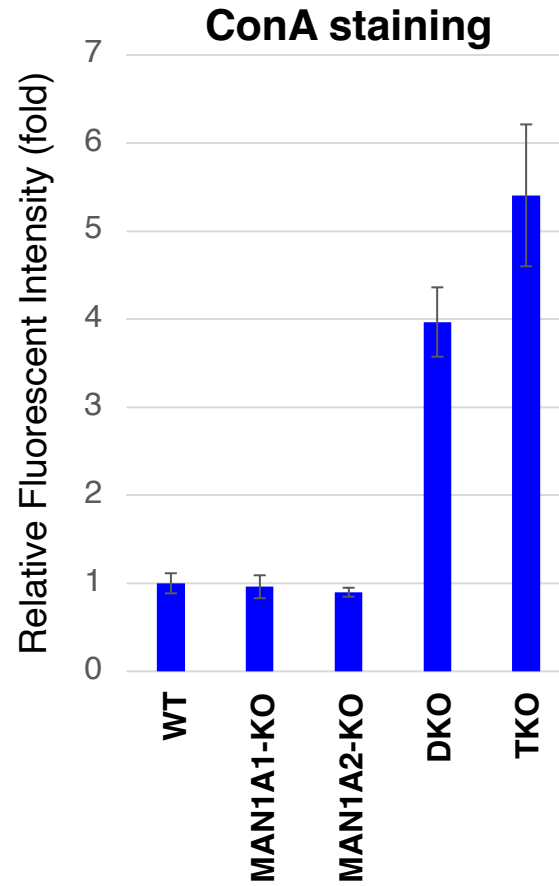
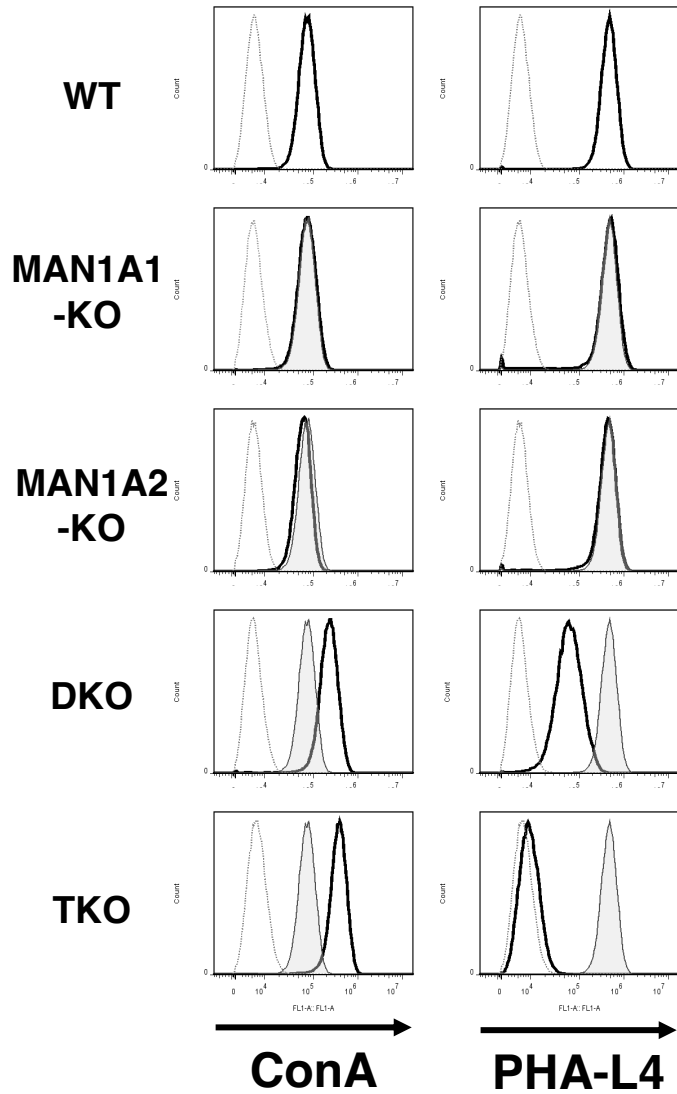
EDEM3

Knockout of MAN1B1 in double-KO cells almost completely diminished the PHA-L4 staining.





# MAN1A1/A2/B1-triple KO cells



Knockout of MAN1B1 in double-KO cells almost completely diminished the PHA-L4 staining.

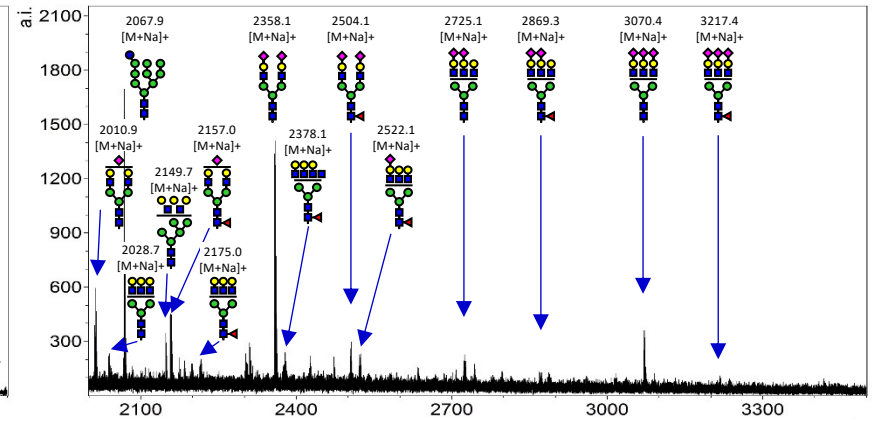
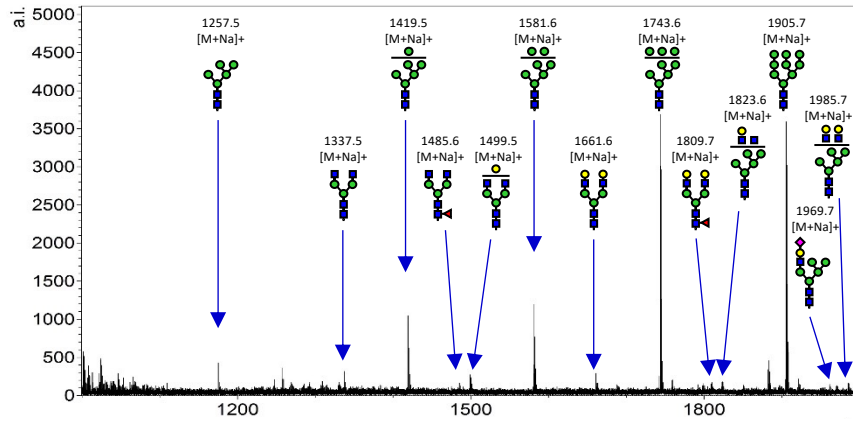
# N-glycan analysis by MALDI-MS (whole cell lysates)

$m/z = 1000 - 2000$

$m/z = 2000 - 3500$

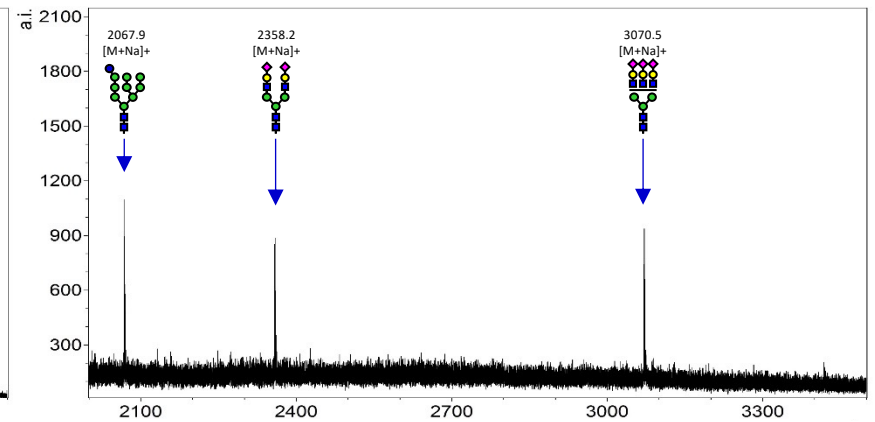
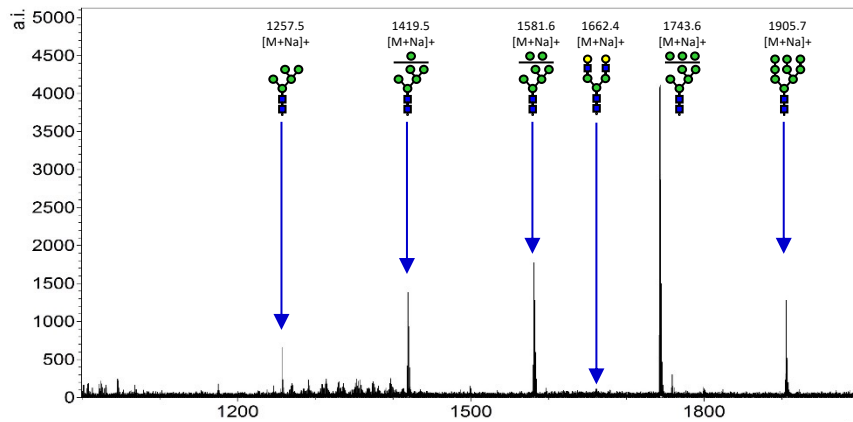
**A**

**WT**



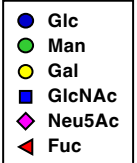
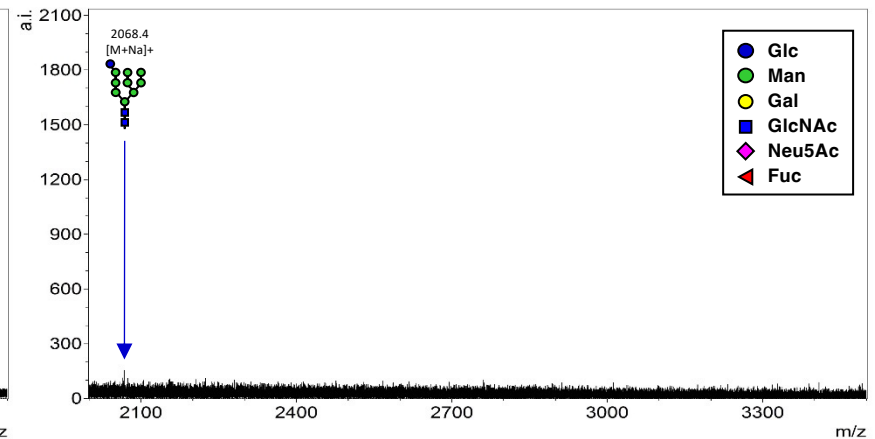
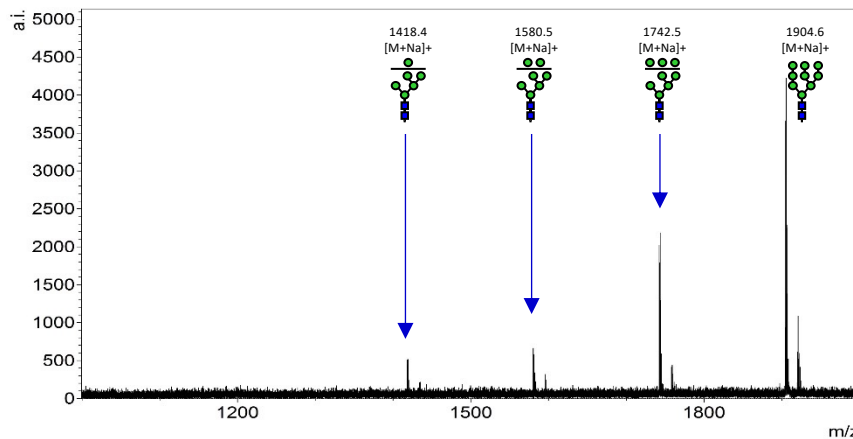
**B**

**D-KO**



**C**

**T-KO**



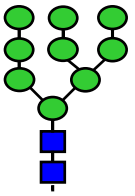
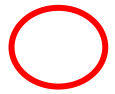
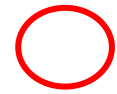
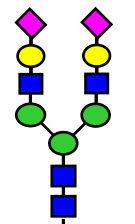
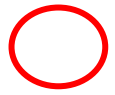

# Expression of recombinant lysosomal enzymes

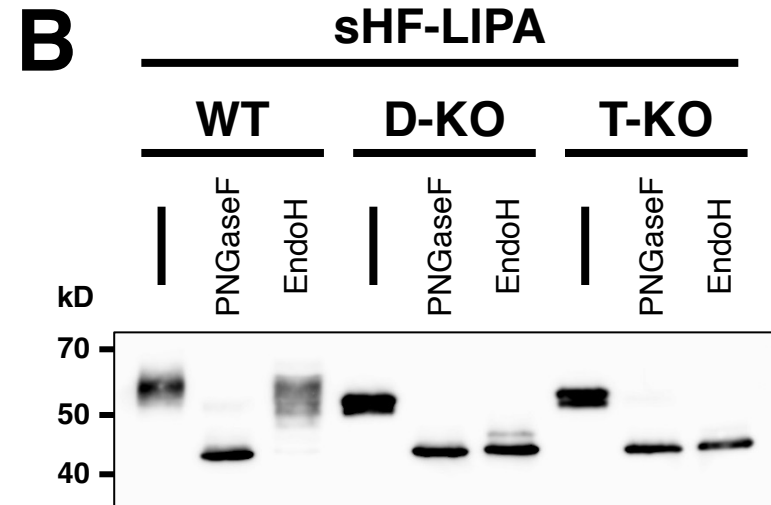
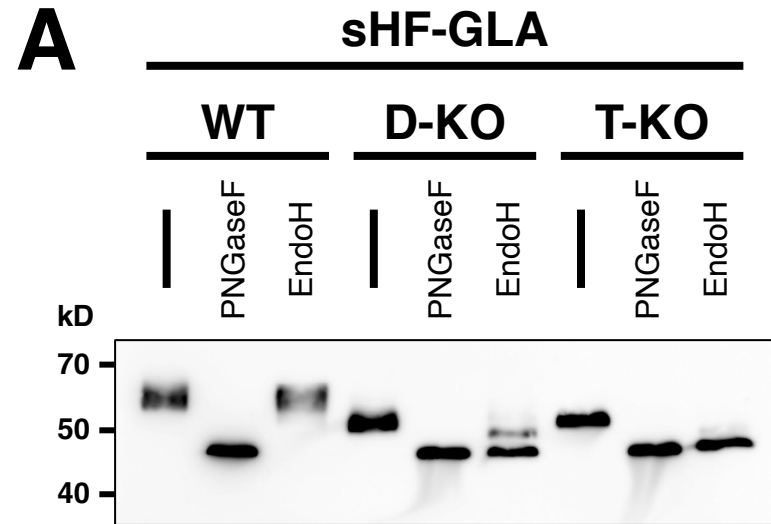
## $\alpha$ -Galactosidase A (GLA):

hydrolyzes a glycosphingolipid Gb3 in the lysosomes and the **mutations in GALA cause Fabry disease**

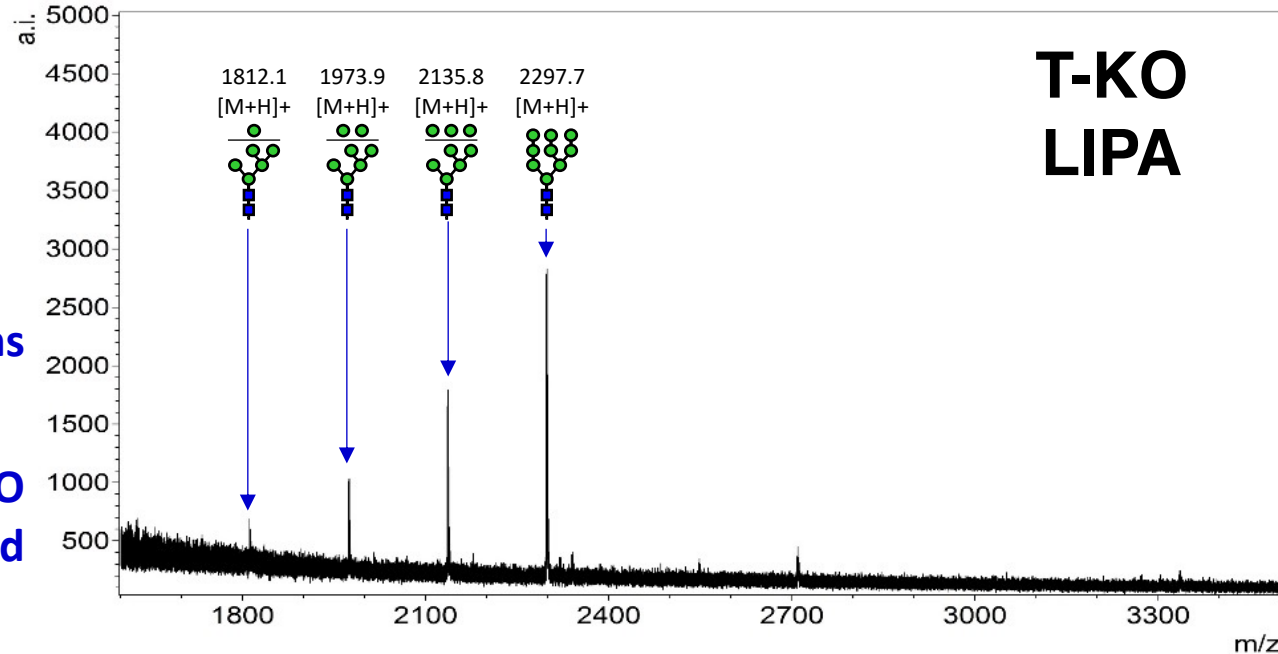
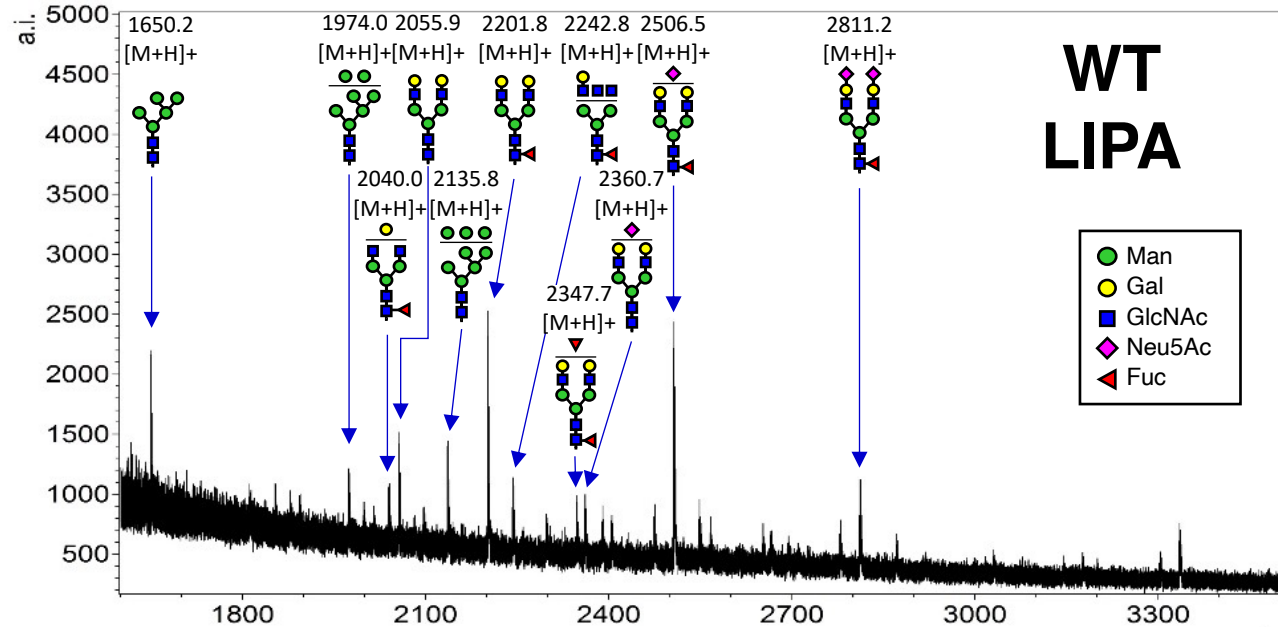
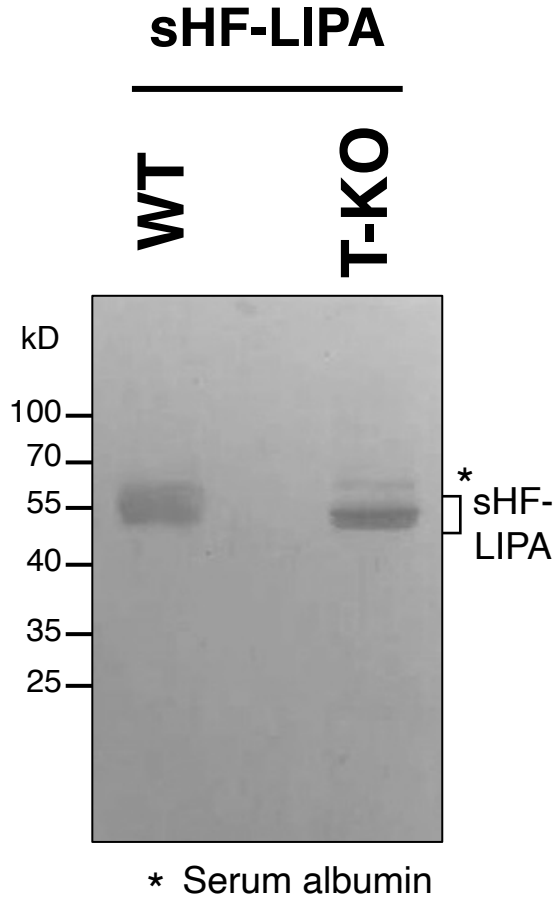
## Lysosomal acid lipase (LIPA):

breakdown of lipids such as cholesterol esters and triacylglycerols in lysosomes, **its deficiency leads Wolman disease and cholesteryl ester storage disease**

	PNGaseF	EndoH
		
		



# N-glycan analysis of recombinant LIPA from T-KO

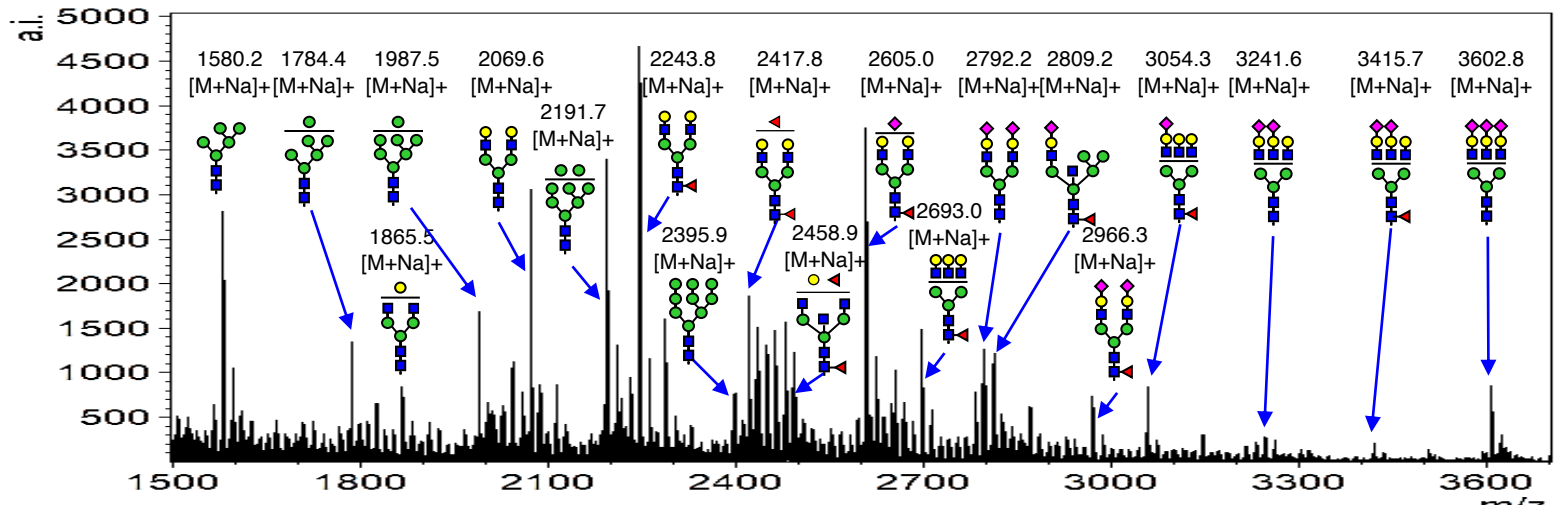
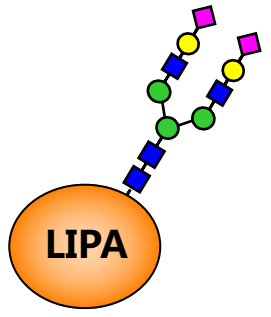


More than 30 different glycans were observed from WT-LIPA.

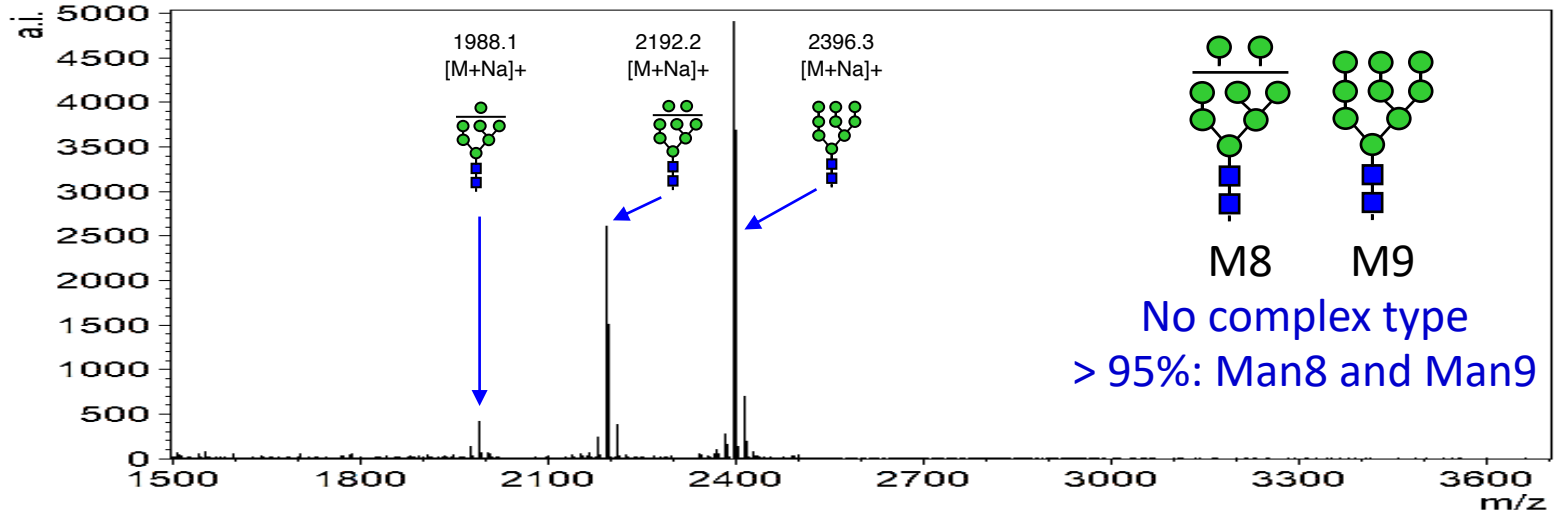
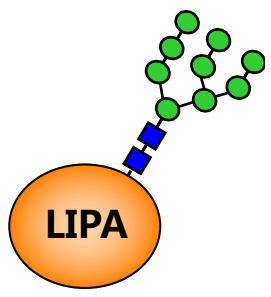
N-glycans on LIPA from T-KO were simplified and changed to high-Man-types.

# MAN1A1/A2/B1/C1-quadruple gene-KO HEK2993 cells

**WT**



**MAN1A1/A2/B1/C1-KO**

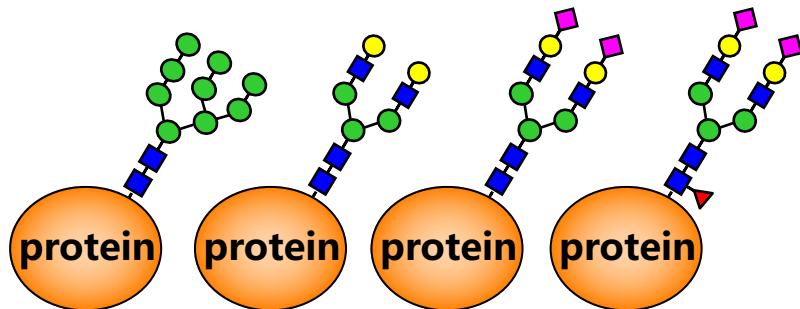


# Summary 1: Engineering of HEK293 cells producing recombinant proteins with high-mannose-type N-glycans

Heterogeneous glycans

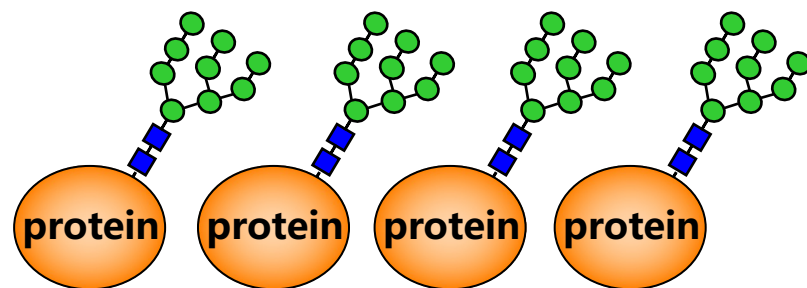
High-Man-type

Complex-type



Homogeneous glycans

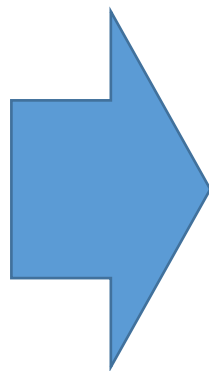
High-Man-type



Cell

Golgi Man-I

ER Man-I



Cell

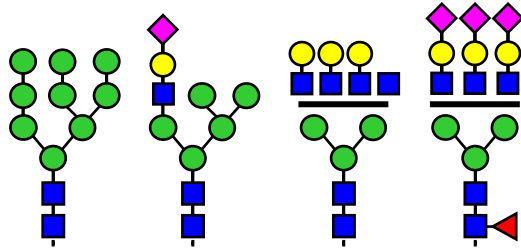
~~Golgi Man-I~~

~~ER Man-I~~

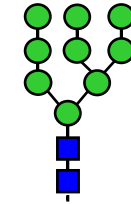
Established multiple mannosidase-I gene KO HEK293 cells, which would be suitable for production of proteins having high-Man-type N-glycans

# Simplification of glycans in HEK293 cells

## N-glycans

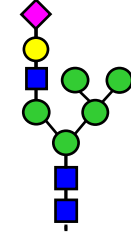


MAN1A1/A2/B1/C1-KO



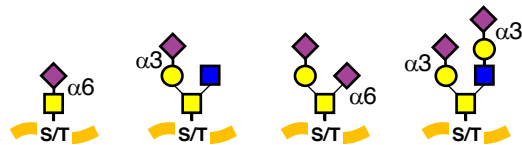
High-Man

MAN2A1/A2/FUT8-KO + MsdS-OE



Hybrid

## O-glycans

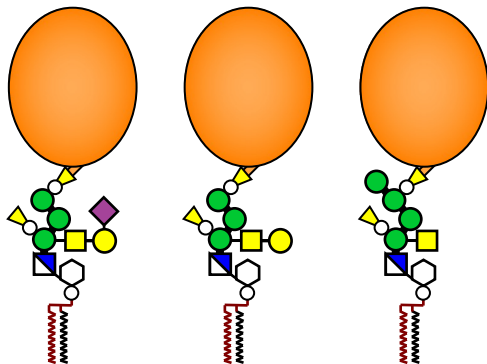


C1GALT1/SLC35A1-KO

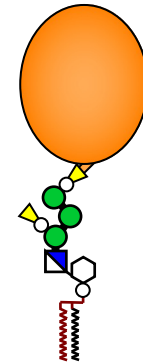


Tn

## GPI-APs



PGAP4/PIGZ-KO + PGAP1-OE



GPI core

# Contents

## **Simplification of N-glycan structures in HEK293:**

Production of recombinant proteins

only having high-mannose type N-glycans

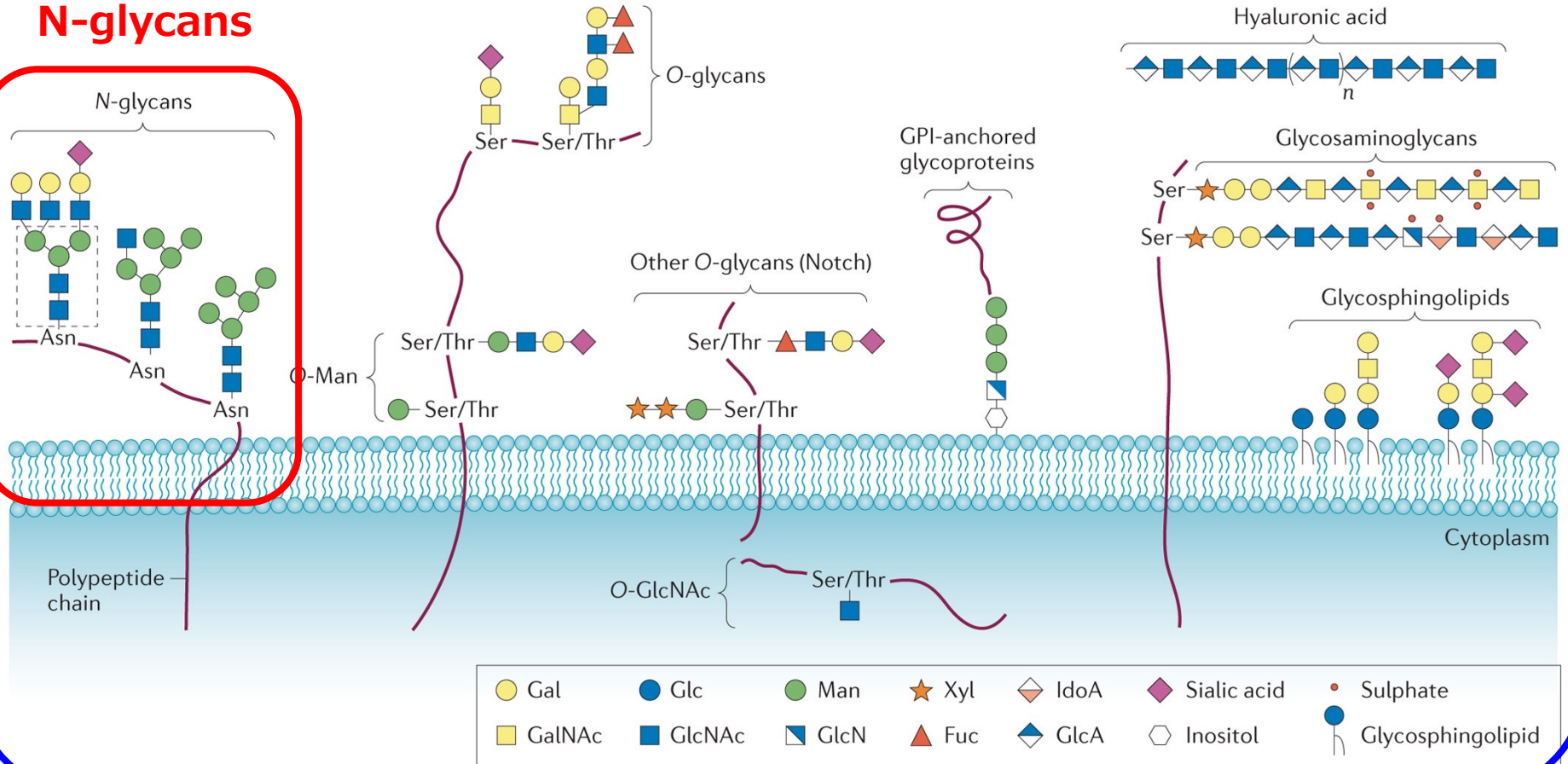
## **Development of a visualization tool for glycan metabolic pathways based on gene expression profiles:**

Glyco-engineering and Glyco-comparison using GlycoMaple



# Mammalian glycan structures

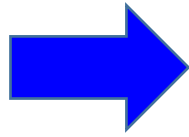
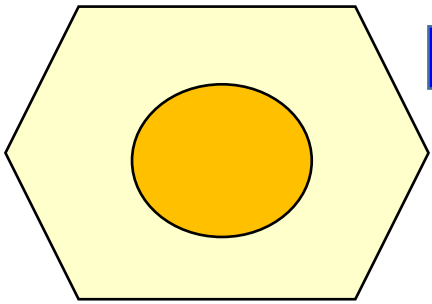
## N-glycans



Nature Reviews | Cancer

**Glycome: Whole glycan structures synthesized in cells/tissues**

# Prediction of Glycan structures



Glycan structures synthesized in cells



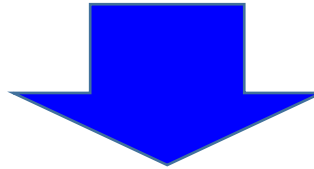
**Glycan analysis** (Mass Spec., HPLC)

Skillful technique is required. It is hard to analyze glycan structures with high molecular weight, charged or isomers.



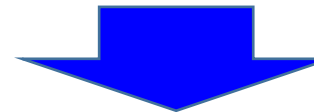
**Glycan research is still a big hurdle for many researchers. It's possible to miss important phenomena involving glycans.**

Need an easy way to access to glycan research



Need a method to analyze comprehensive glycan structures

**It is useful if Glycan structures are predicted from Gene expression profiles**

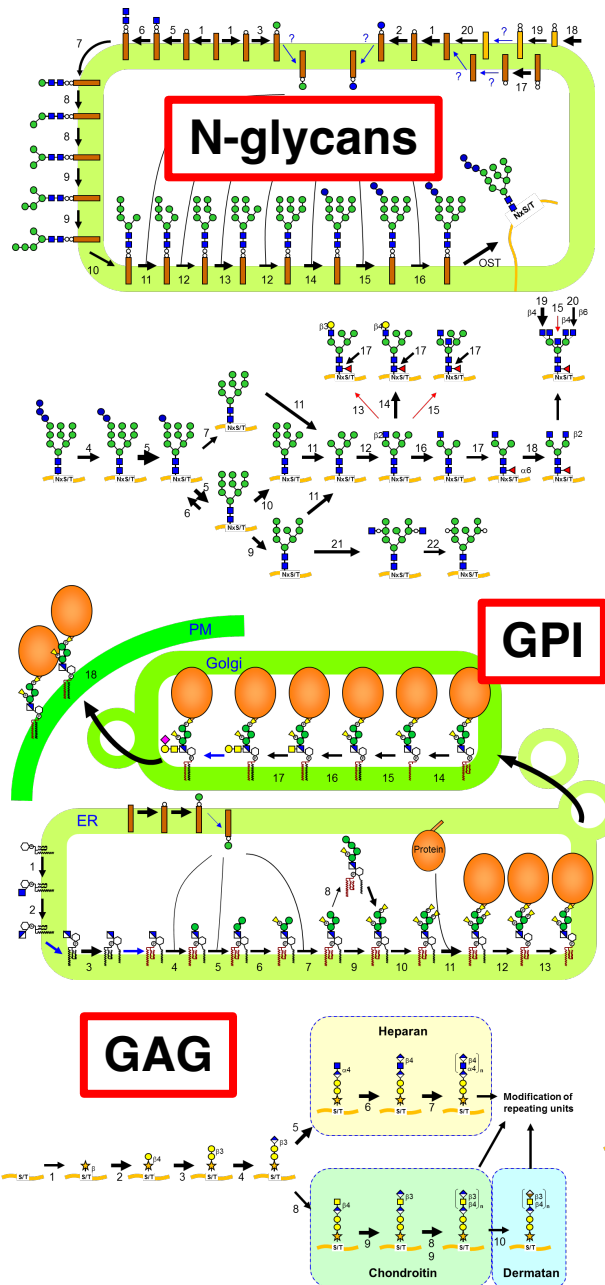


- (1) List of Glycan-related genes**
- (2) Maps of Glycan metabolisms**

# (1) List of glycan-related genes

Group	No. of genes
1. Lipid-linked oligosaccharide (LLO) biosynthesis	38
2. N-glycan processing and branching	41
3. Glycosaminoglycan (GAG) biosynthesis and proteins	64
4. O-glycan (mucin-type) biosynthesis	33
5. O-glycan (others) biosynthesis	36
6. Glycosphingolipid (GSL) biosynthesis	77
7. N-glycan / O-glycan / GSL modification	47
8. GPI biosynthesis and proteins	178
9. C-mannosylation	4
10. Sugar-nucleotide biosynthesis	56
11. Sugar transporters	47
12. Golgi homeostasis	19
13. Lectins	172
14. Glycogen synthesis/metabolism	14
15. Hyaluronan synthesis/metabolism	11
16. Sulfate related	18
17. Lysosomal degradation of glycans	16
18. Other Glycosyltransferase (CAZy)	33
19. Other Glycoside hydrolase (CAZy)	32
20. Carbohydrate binding module (CAZy)	15
<b>Total</b>	<b>951</b>

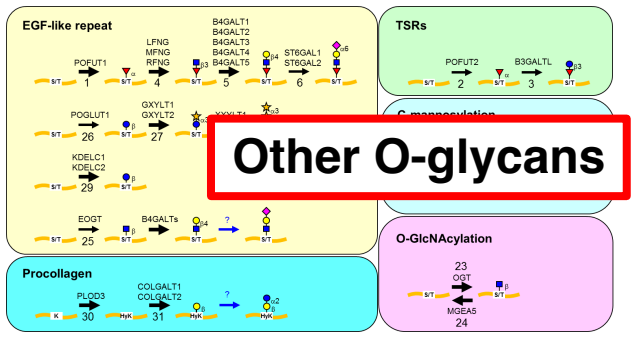
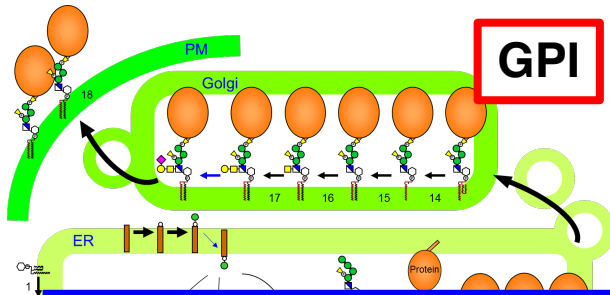
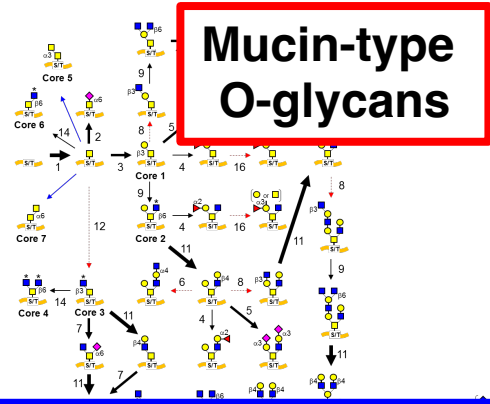
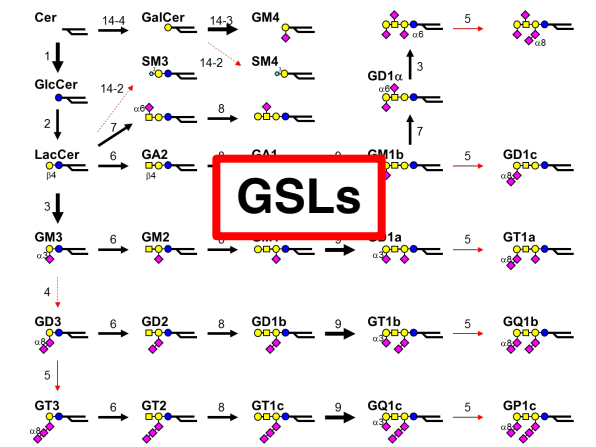
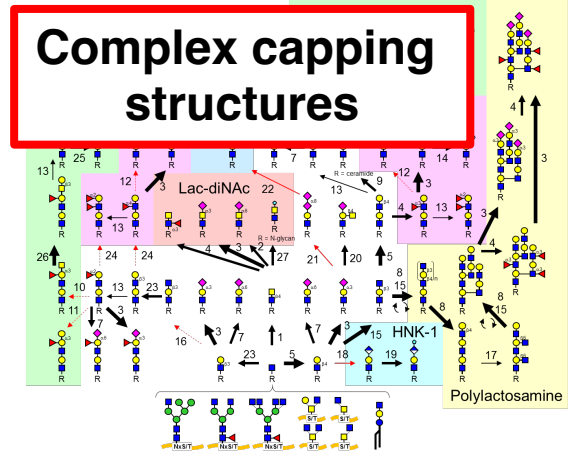
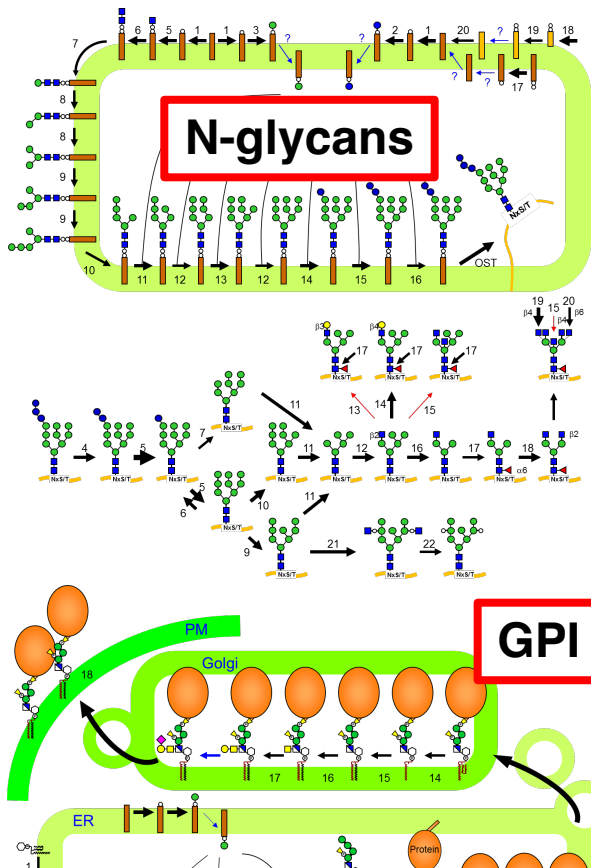
# (2) Drawing glycan metabolic maps



## Map Number

1. Lipid-linked oligosaccharide (LLO) biosynthesis
2. N-glycan processing and branching
3. Complex capping of N-glycan / O-glycan / GSLs
4. GPI biosynthesis
5. O-GalNAc (mucin-type) biosynthesis
6. O-Fuc / O-Glc / Col-Gal / O-GlcNAc / C-Man
7. O-Man biosynthesis
8. Glycosaminoglycan (GAG) biosynthesis
9. Heparan sulfate biosynthesis
10. Chondroitin sulfate and dermatan sulfate
11. Keratan sulfate
12. Glycosphingolipid (core) biosynthesis
13. Globoside biosynthesis
14. Ganglioside biosynthesis
15. Sugar nucleotide biosynthesis
16. Lysosomal degradation of N-glycans
17. Lysosomal degradation of GSLs
18. Lysosomal degradation of GAGs
19. Hyaluronic acid biosynthesis and catabolism
20. Human milk oligosaccharide

# Mapping to glycan metabolic pathways



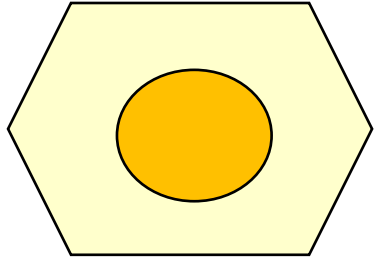
Development of "GlycoMaple", a tool to visualize and predict glycan metabolic pathways based on gene expression profiles

<https://glycosmos.org/glycomaple/index>



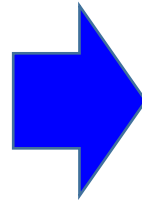
Huang et al. (2021) *Dev. Cell*

# Expression profiles of glycan-related genes in HEK293 cells



Model cell:  
**HEK293**

**RNA-seq**



Calculate each gene expression as  
TPM: Transcripts Per Million


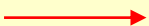
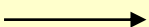




$$\text{TPM} = \frac{\text{Reads} / \text{length of transcript}}{\sum_{\text{All transcripts}} \left( \text{Reads} / \text{length of transcript} \right)} \times 10^6$$



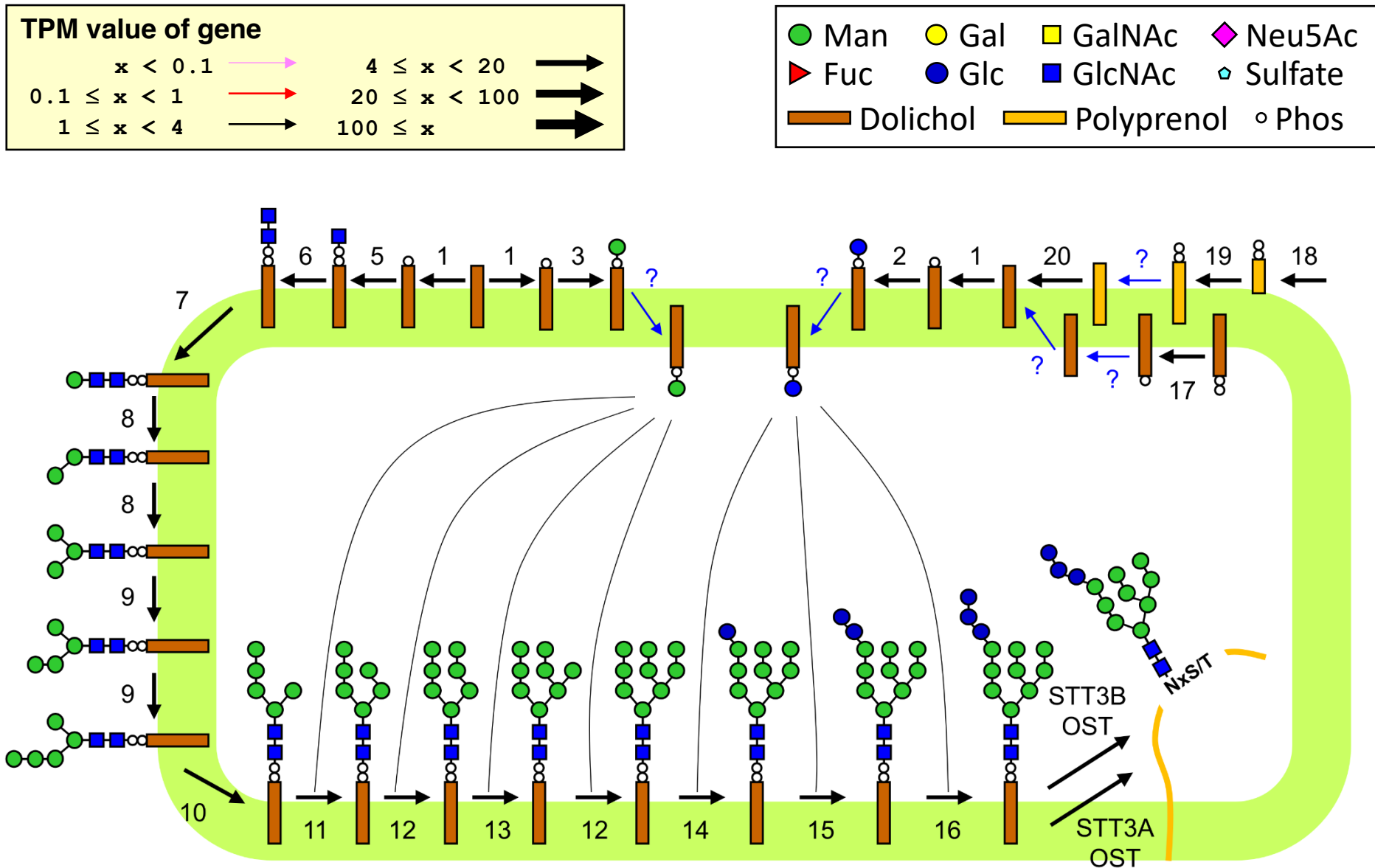
Analyze 951 glycan-related gene expression



Integrate the information to the glycan metabolic pathways

TPM value of gene	Biosynthetic pathway
$x < 0.1$	 } <b>Not expressed</b>
$0.1 \leq x < 1$	 } <b>Very weak</b>
$1 \leq x < 4$	 } <b>Expressed</b>
$4 \leq x < 20$	 }
$20 \leq x < 100$	 }
$100 \leq x$	 }
Unknown gene	

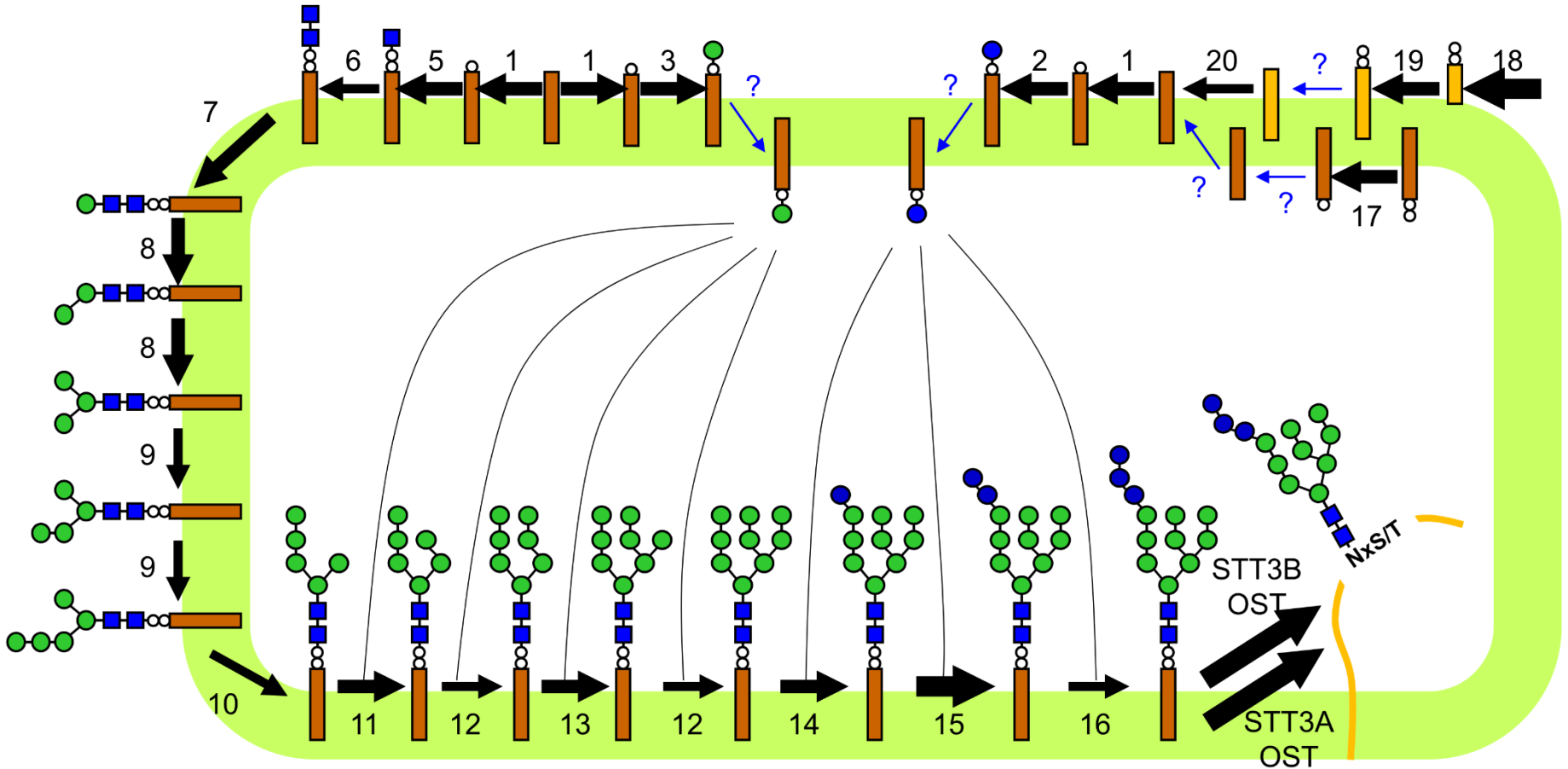
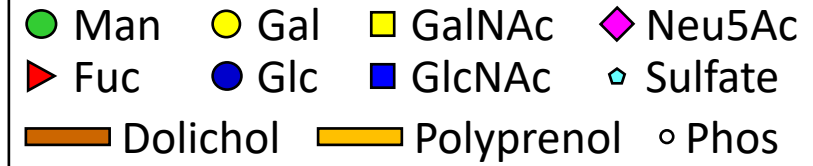
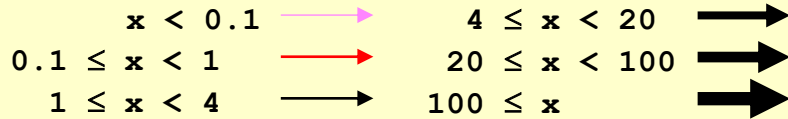
# LLO biosynthesis and OST



LLO: lipid-linked oligosaccharide; LLO is a precursor for N-glycosylation.

# LLO biosynthesis and OST in HEK293

TPM value of gene

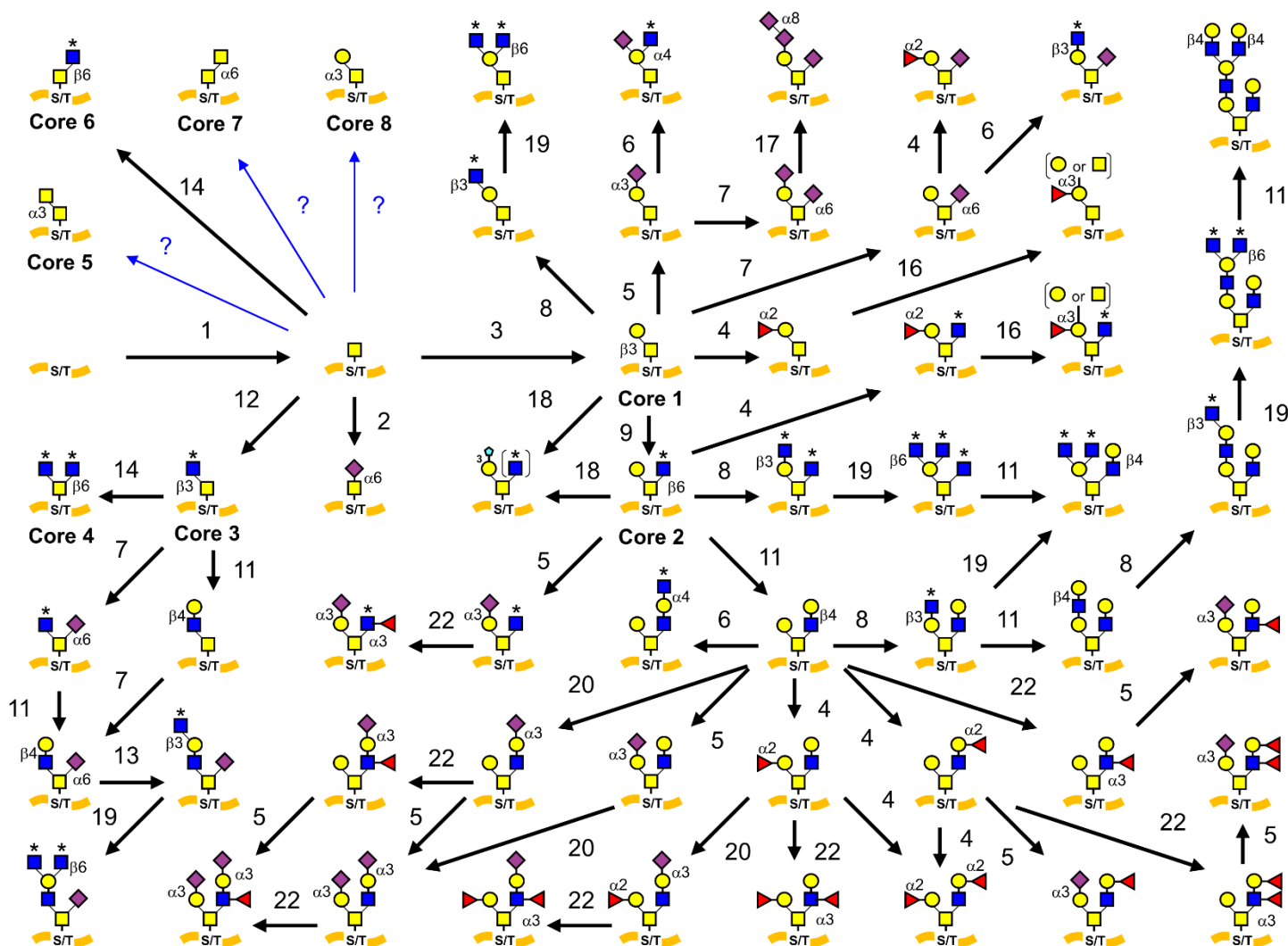
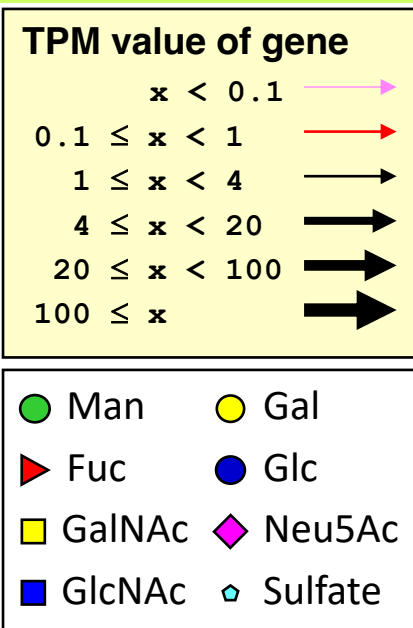


**The LLO pathway is very basic and essential for cells.**

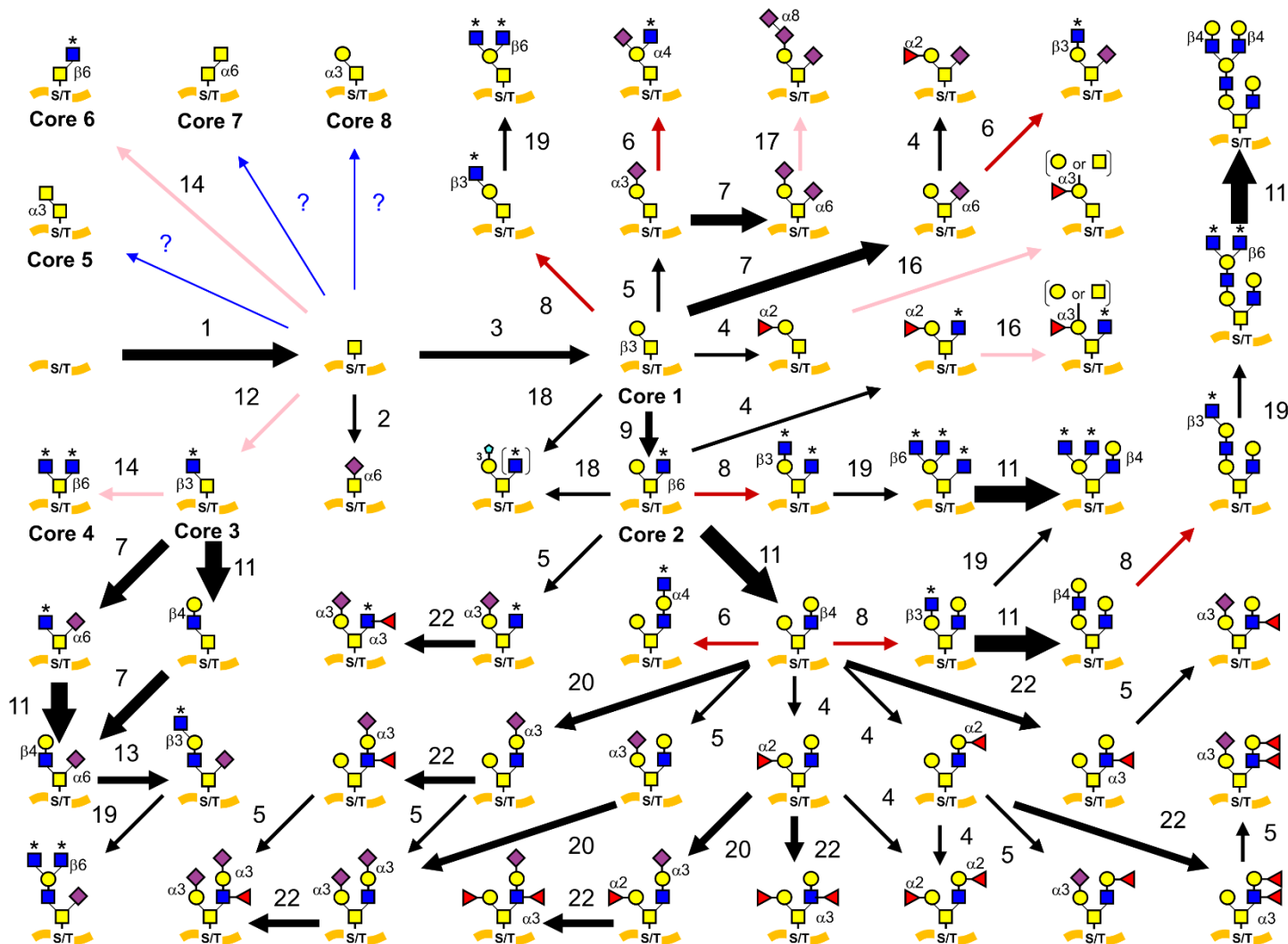
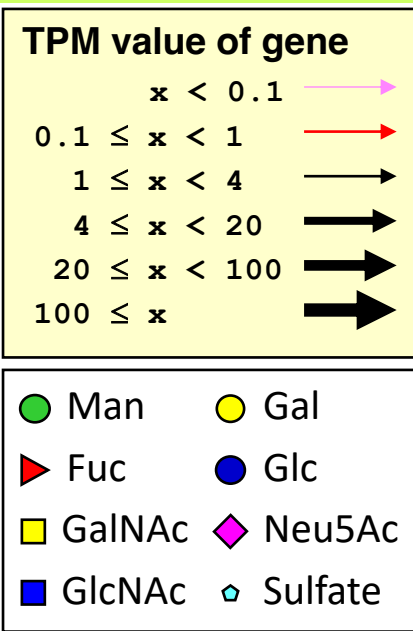
LLO: lipid-linked oligosaccharide; LLO is a precursor for N-glycosylation.



# Biosynthesis of mucin-type O-glycans

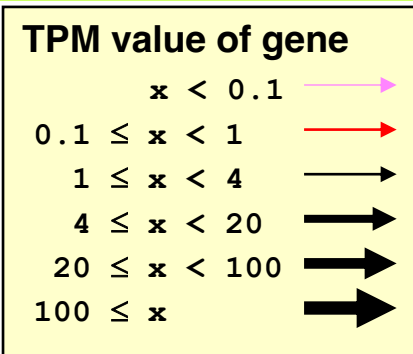


# Biosynthesis of mucin-type O-glycans in HEK293

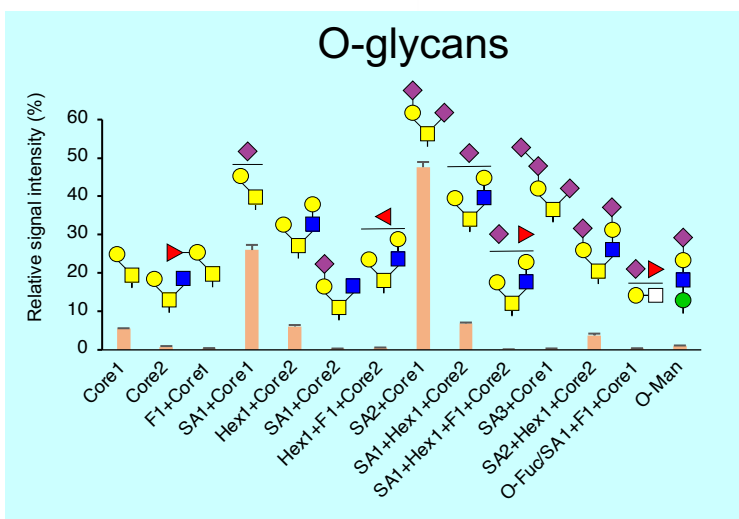


Several genes required for the pathways  
are not expressed in HEK293 cells

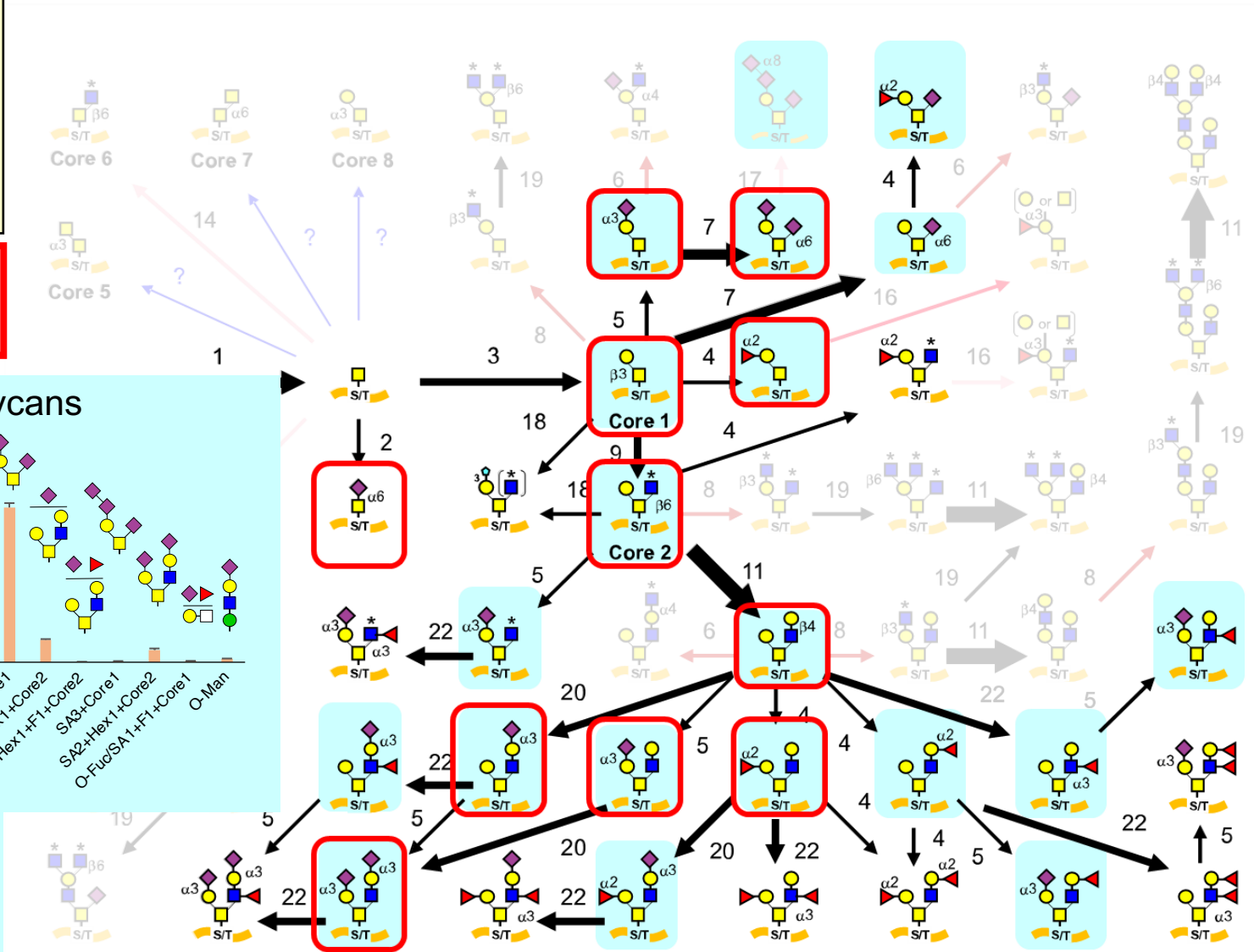
# Biosynthesis of mucin-type O-glycans in HEK293



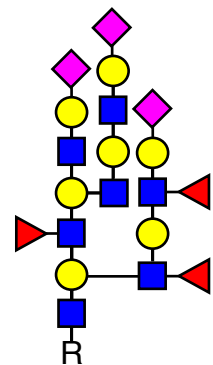
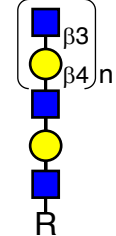
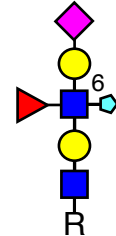
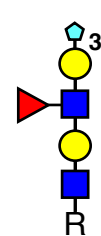
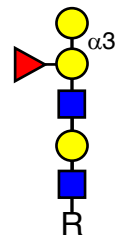
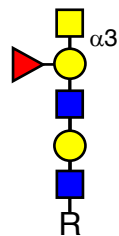
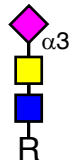
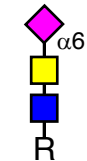
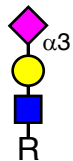
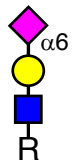
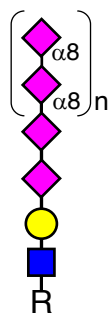
Razawi H et al. (2013)  
*Glycobiology* 23: 935-945



Our glycomic analysis  
 (ESI-MS)



# Complex capping of N-glycans / O-glycans / GSLs



Polysialic acid

LacNAc

Lac-diNAc

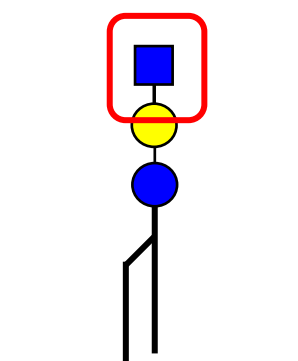
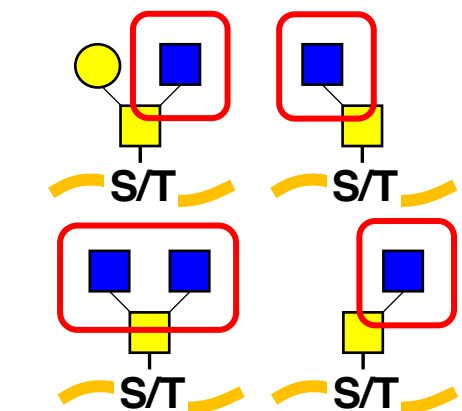
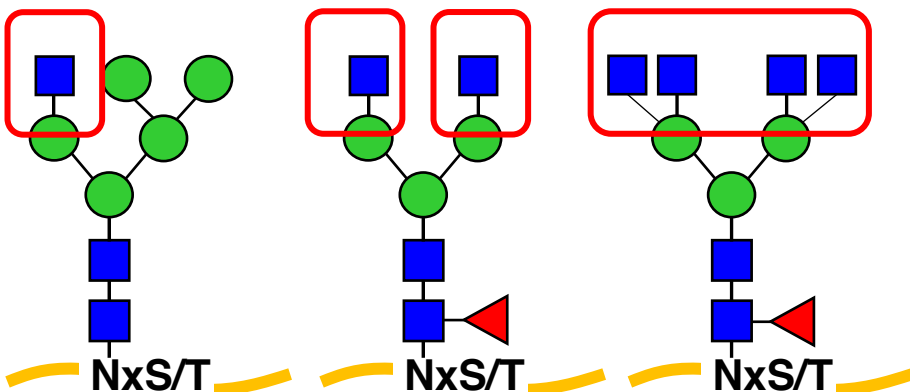
ABO

Lewis

Polylactosamine



Man	Gal	GalNAc	Neu5Ac
Fuc	Glc	GlcNAc	Sulfate

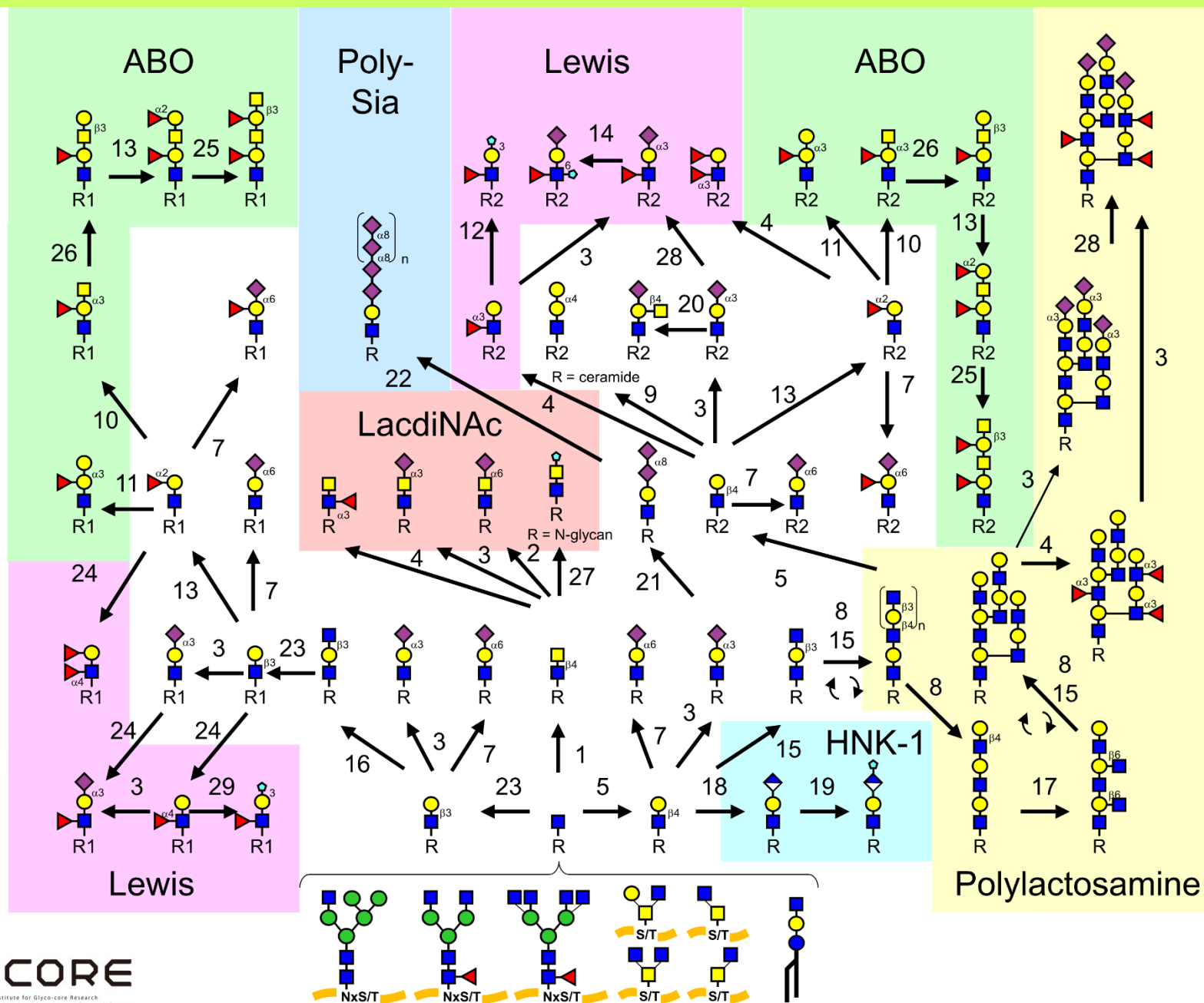


N-glycans

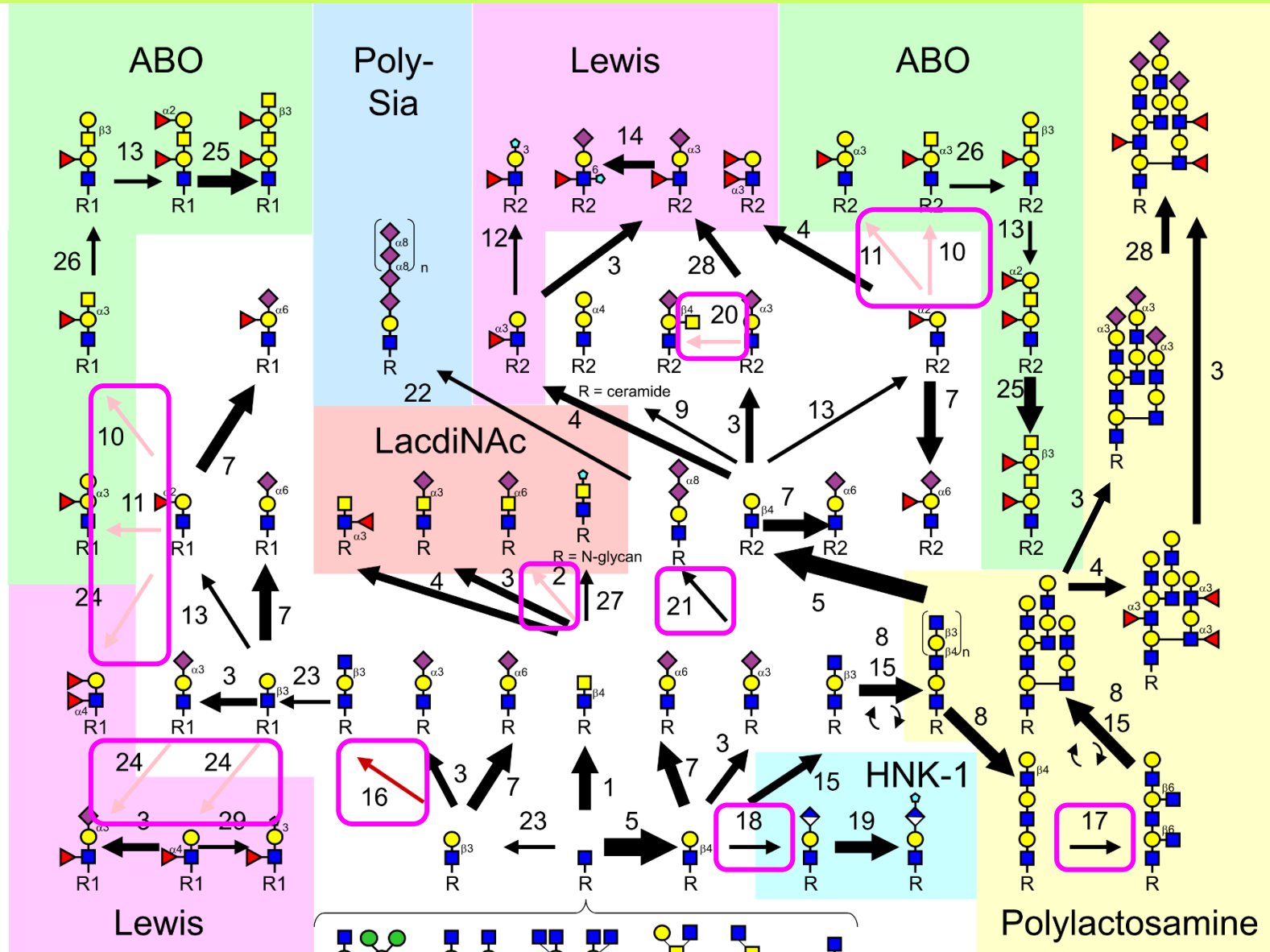
O-glycans

Glycolipids

# Complex capping of N-glycans / O-glycans / GSLs

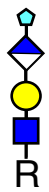


# Complex capping of N-glycans / O-glycans / GSLs in HEK293



**GlycoMaple could find rate-limiting steps**

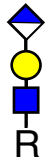
# Customization: Expression of HNK-1 epitope in HEK293



**HNK-1 epitope**

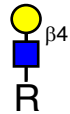


**CHST10: 25.40**



**B3GAT1: 0.61**

B3GAT2: 2.49



B4GALT1: 64.06

**B4GALT2: 92.05**

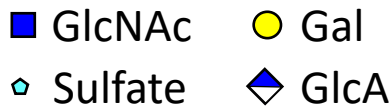


B4GALT3: 53.71

B4GALT4: 27.04

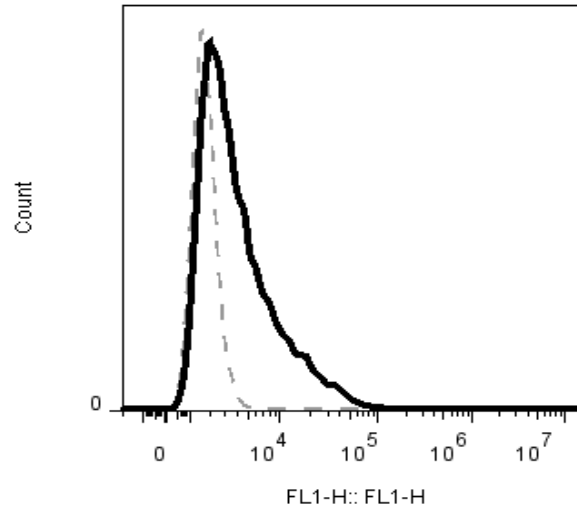


B4GALT5: 76.22

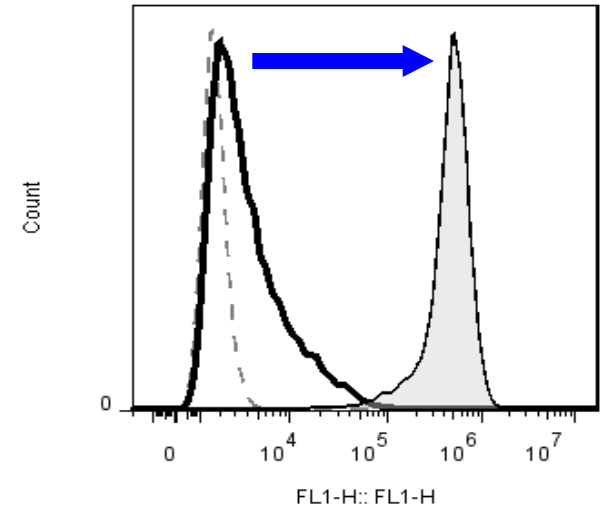


background  
 HEK293  
 + B3GAT1

**HEK293**



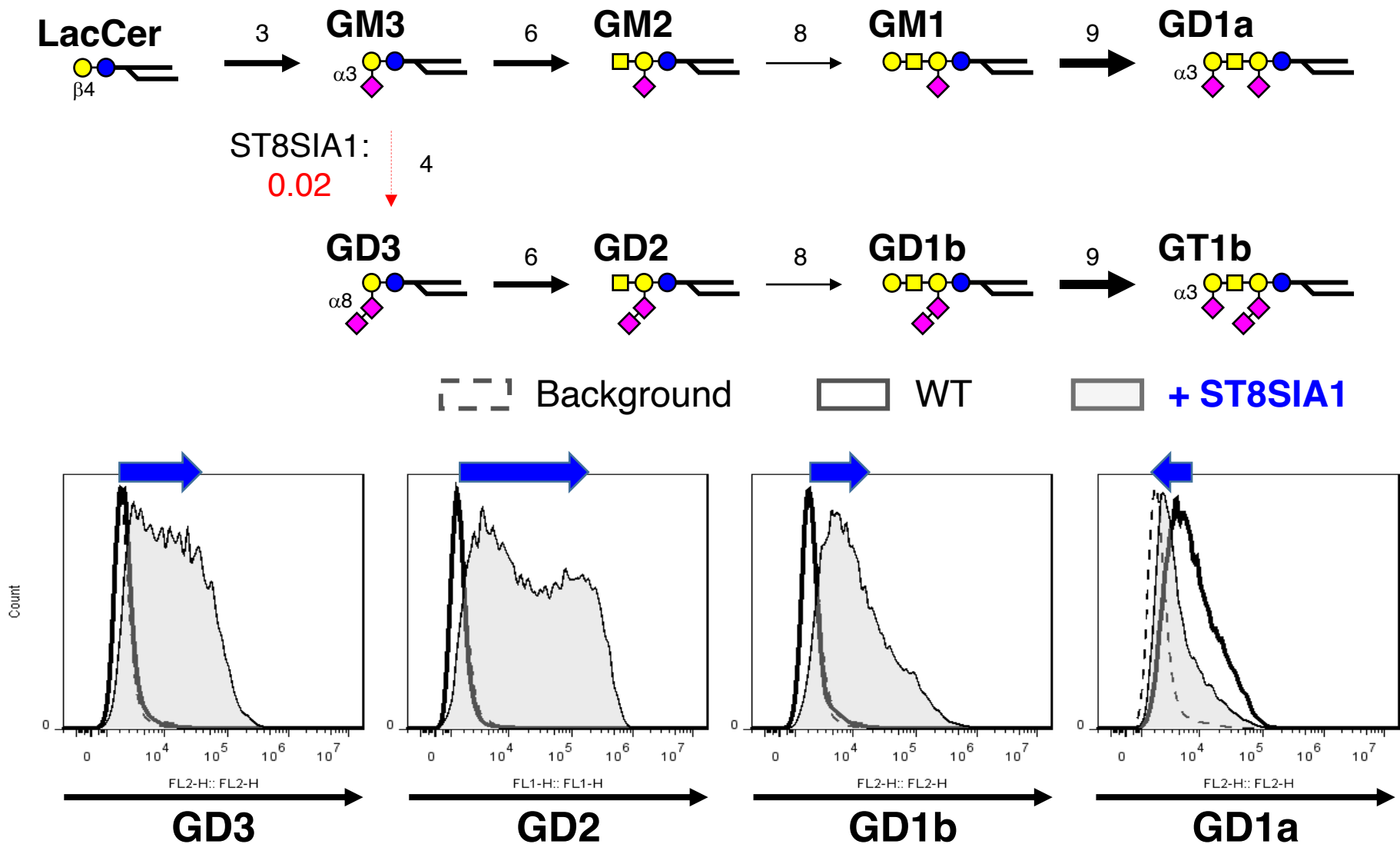
**+ B3GAT1**



**HNK-1 epitope (CD57)**

**GlycoMaple enables us to find rate-limiting steps and to customize glycosylation pathways.**

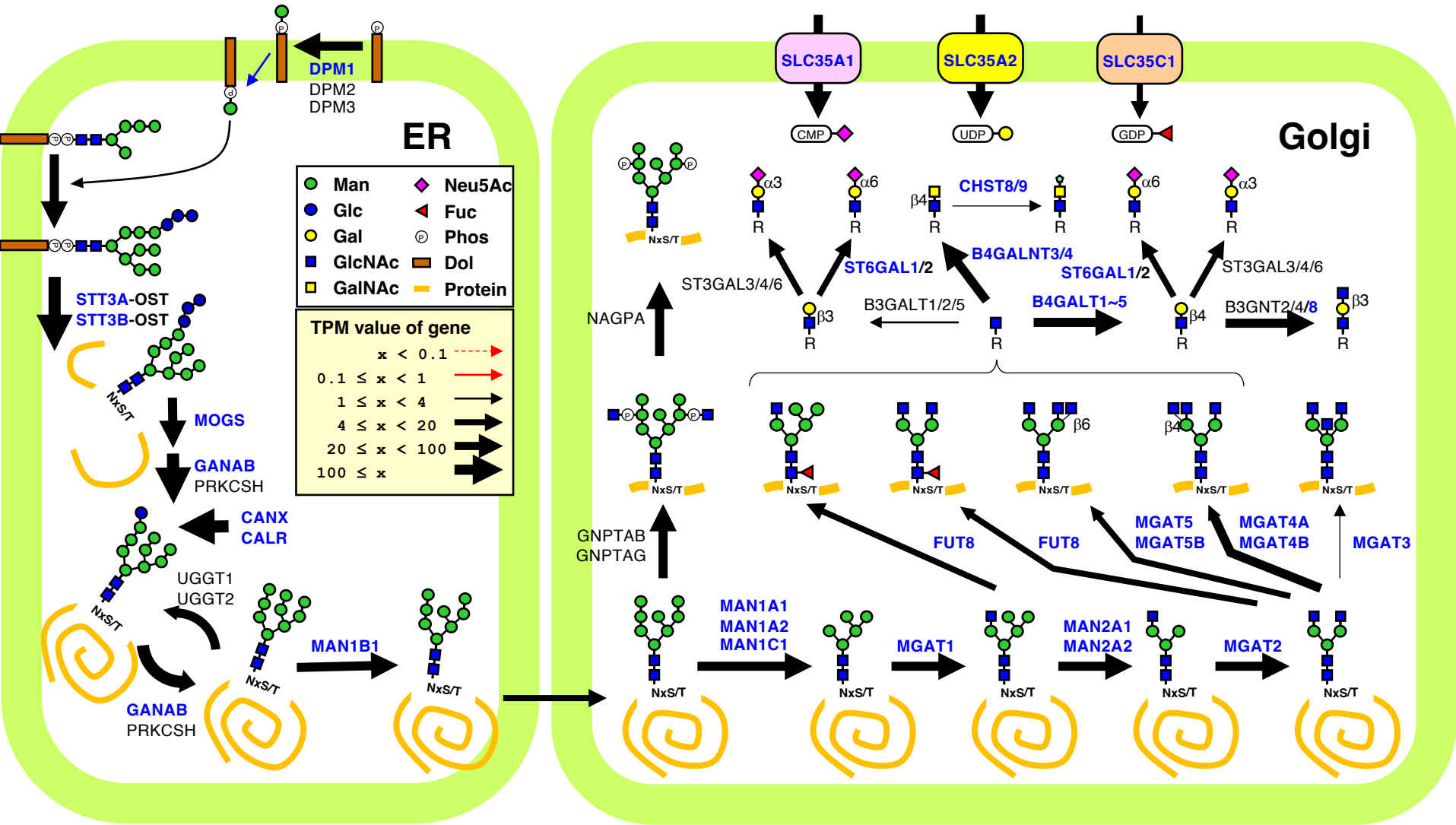
# Customization: Expression of gangliosides in HEK293



**GlycoMaple enables us to find rate-limiting steps and to customize glycosylation pathways.**

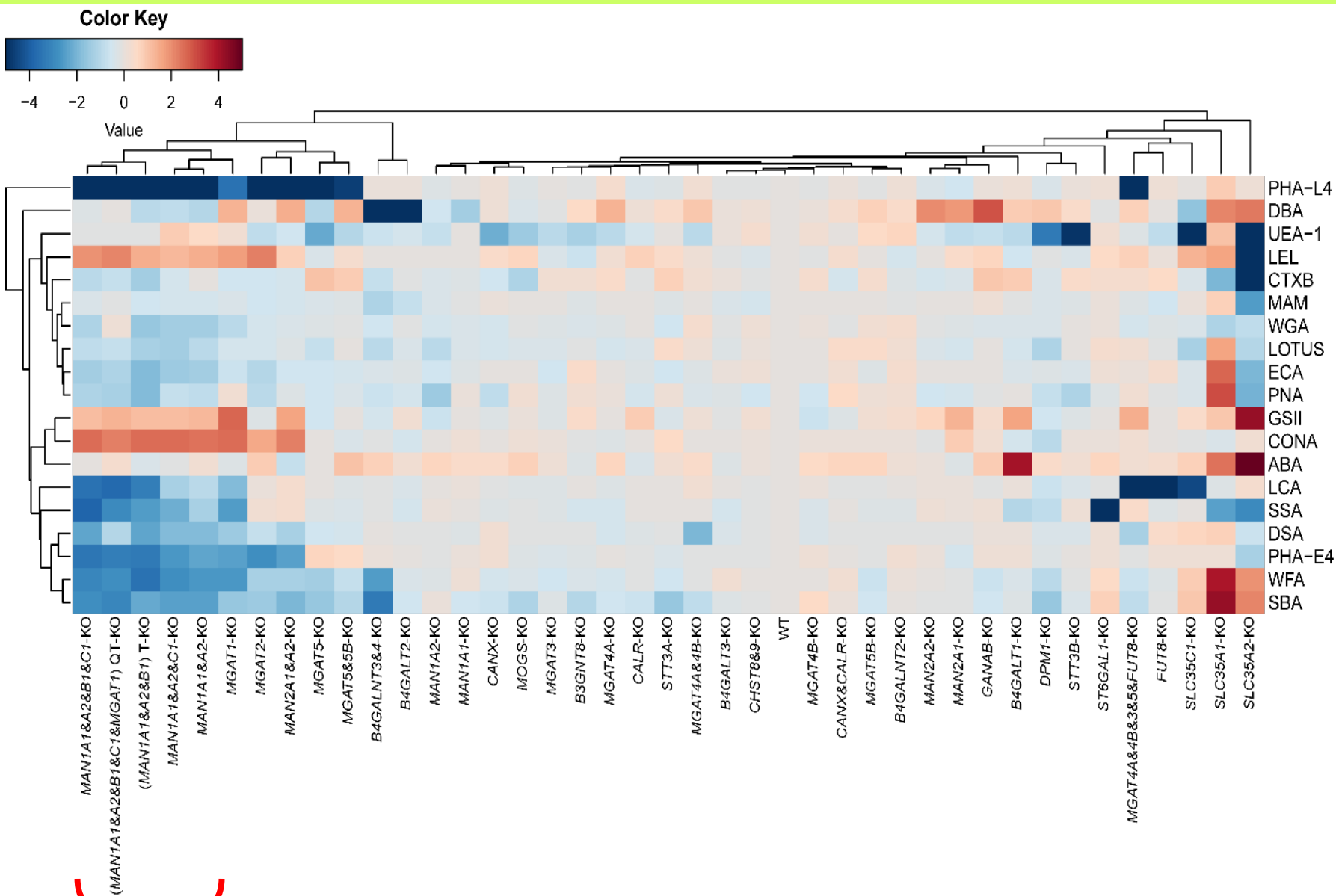


# Construction of Glyco-Gene KO library



Based on the gene expression profiles in HEK293 cells,  
 We selected genes required for N-glycan processing (Blue)  
**We constructed 40 different gene KO cell library.**

# Lectin staining of 40 Glyco-Gene KO cell library

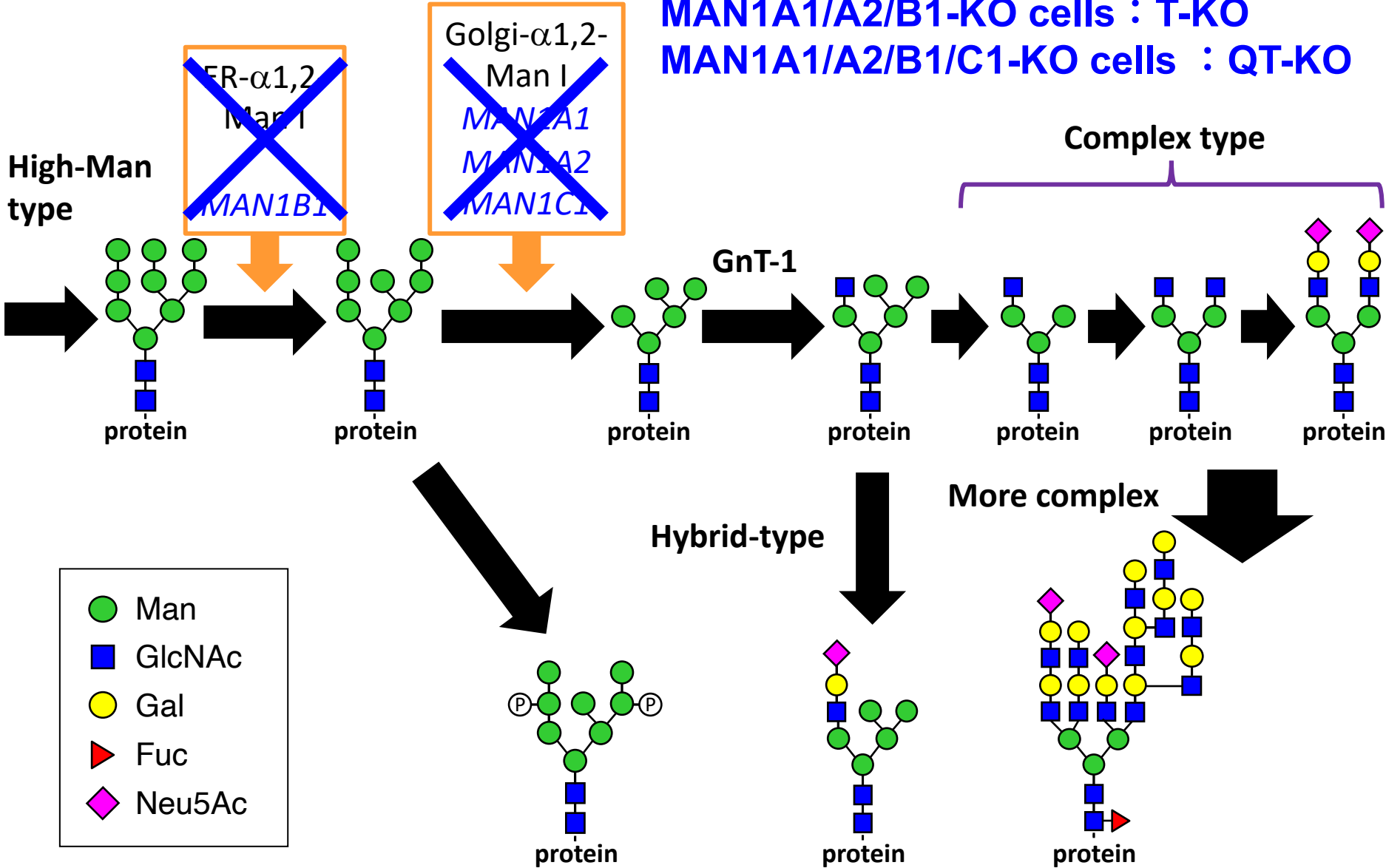


Mannosidase-I  
Gene KO cells

40 Glyco-Gene KO cells

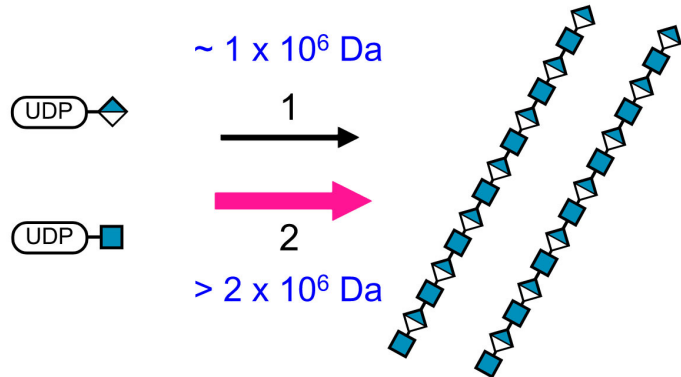
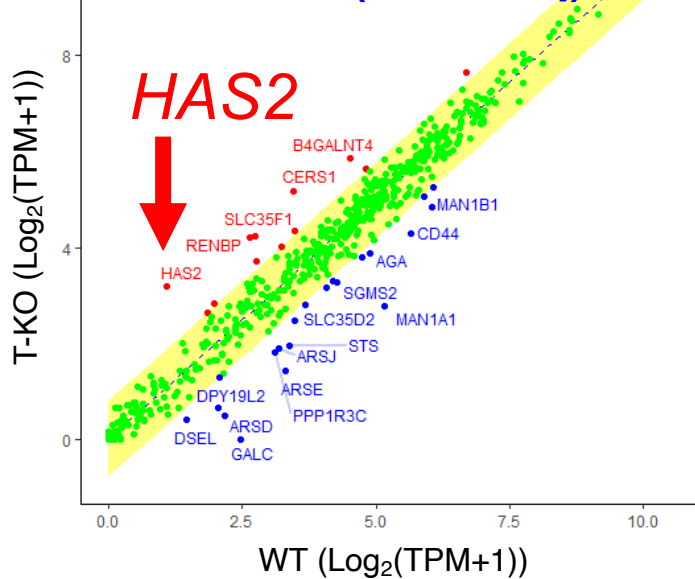
# Disruption of multiple mannosidase-genes in HEK2993

MAN1A1/A2/B1-KO cells : T-KO  
 MAN1A1/A2/B1/C1-KO cells : QT-KO



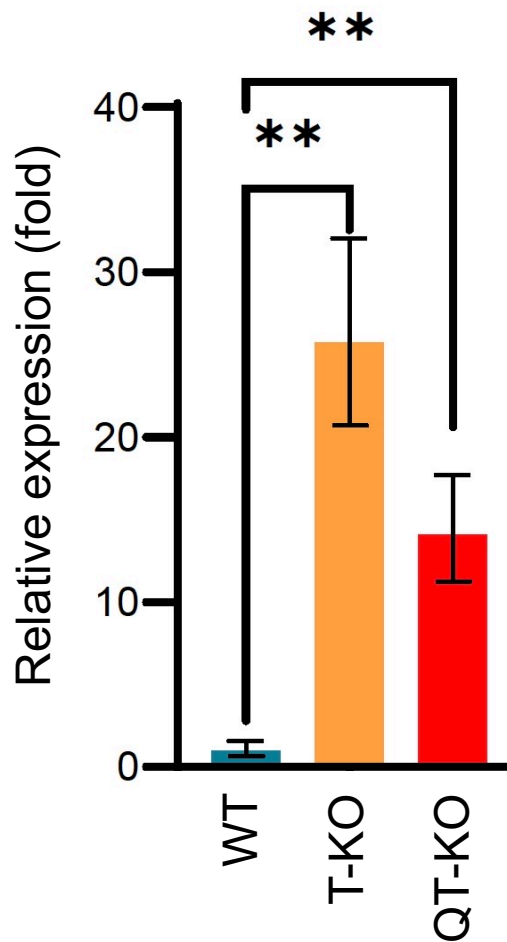
# Hyaluronan was increased in Mannosidase-KO cells

## Comparison of glycan-related gene expression between WT and T-KO (RNA-seq)

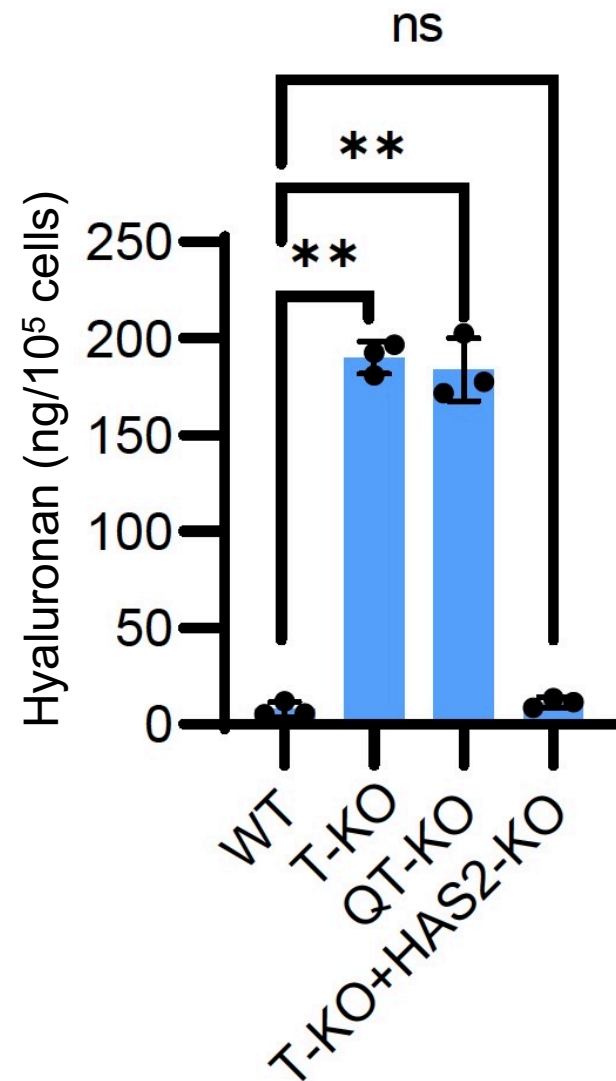


T-KO / WT : 4.3-fold ↑  
 QT-KO / WT : 4.0-fold ↑

## Expression of HAS2 mRNA (qPCR)

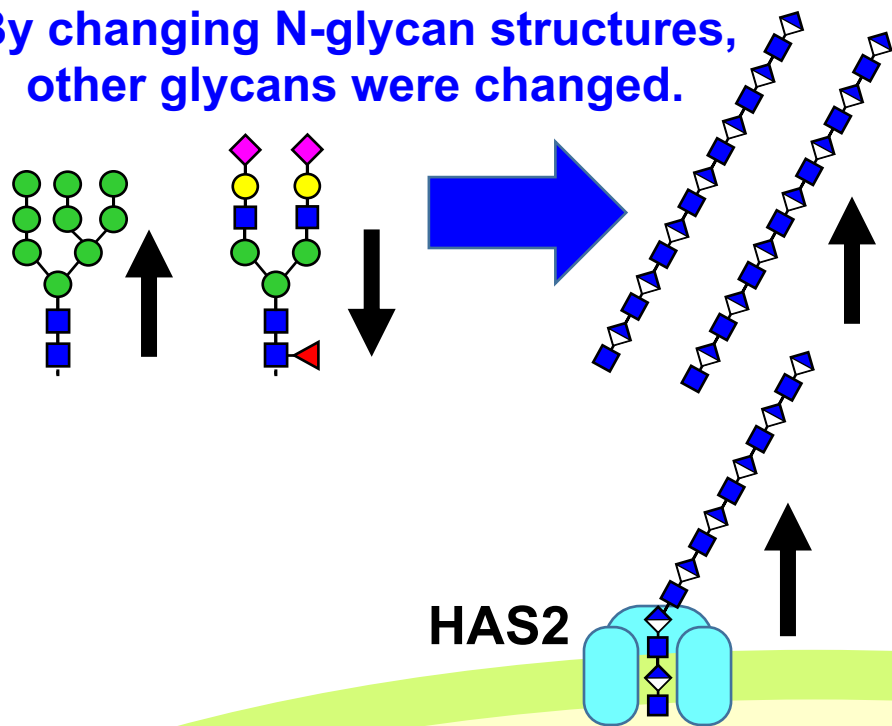


## Amounts of Hyaluronan

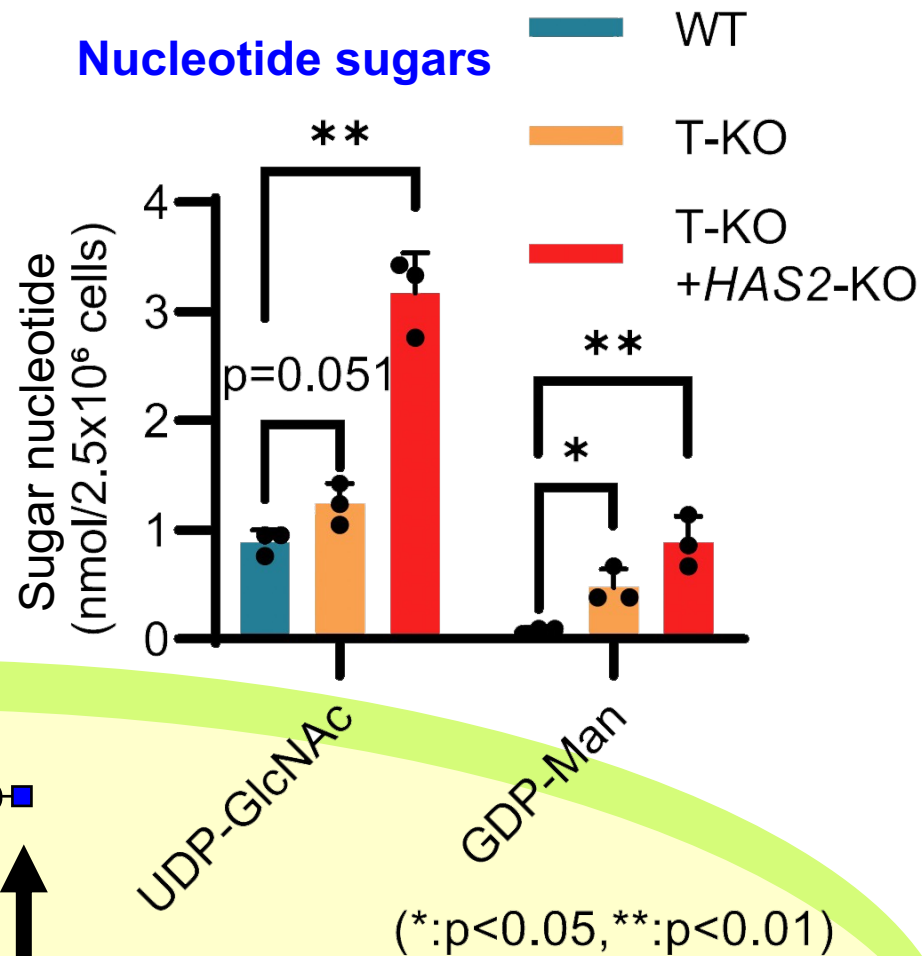


# Hyaluronan was increased in Mannosidase-KO cells

By changing N-glycan structures, other glycans were changed.



Nucleotide sugars



Man-I KO

HAS2 ↑

(\*: p < 0.05, \*\*: p < 0.01)

# GlycoMaple analysis using Kidney RNA-seq data in TCGA

TCGA (The Cancer Genome Atlas)

Comparison of normal and diseased tissues

## Kidney

Solid Tissue Normal (Normal) N = 140

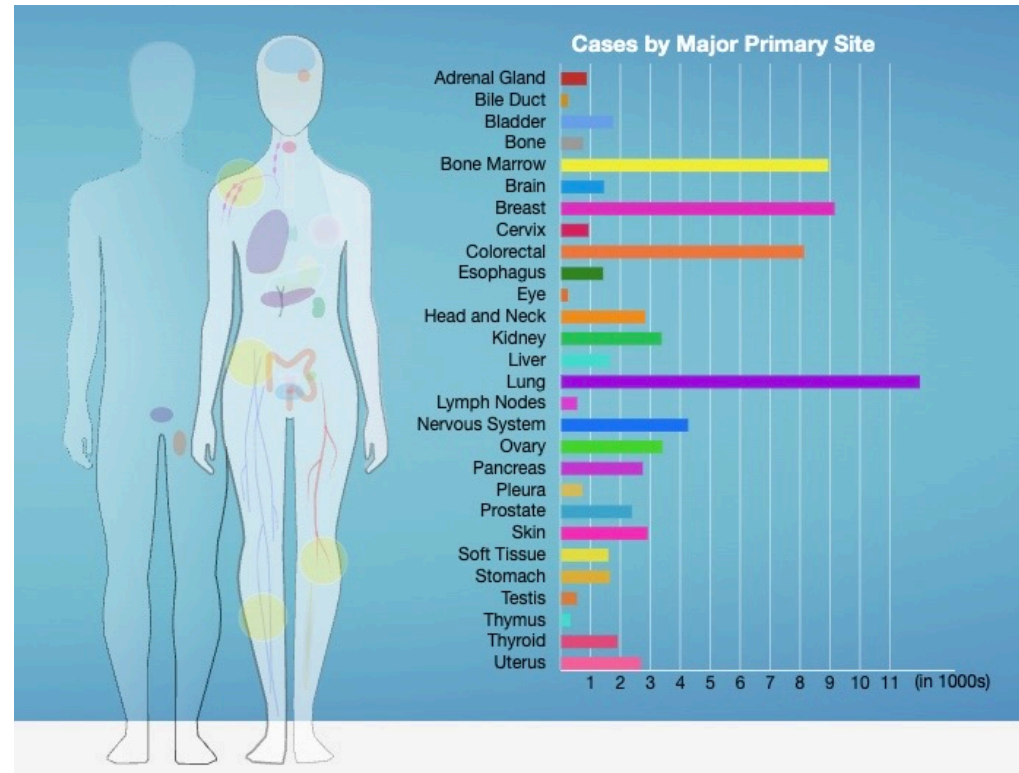
Clear cell carcinoma (ccRCC) N = 530

Papillary cell carcinoma (pRCC) N = 288



UCSC Xena

See the bigger picture



<https://portal.gdc.cancer.gov/>  
<https://xenabrowser.net/>

# Biosynthesis of capping structures in Kidney tumors

**Normal tissue**  
(N = 140; Median)

V.S.

ccRCC

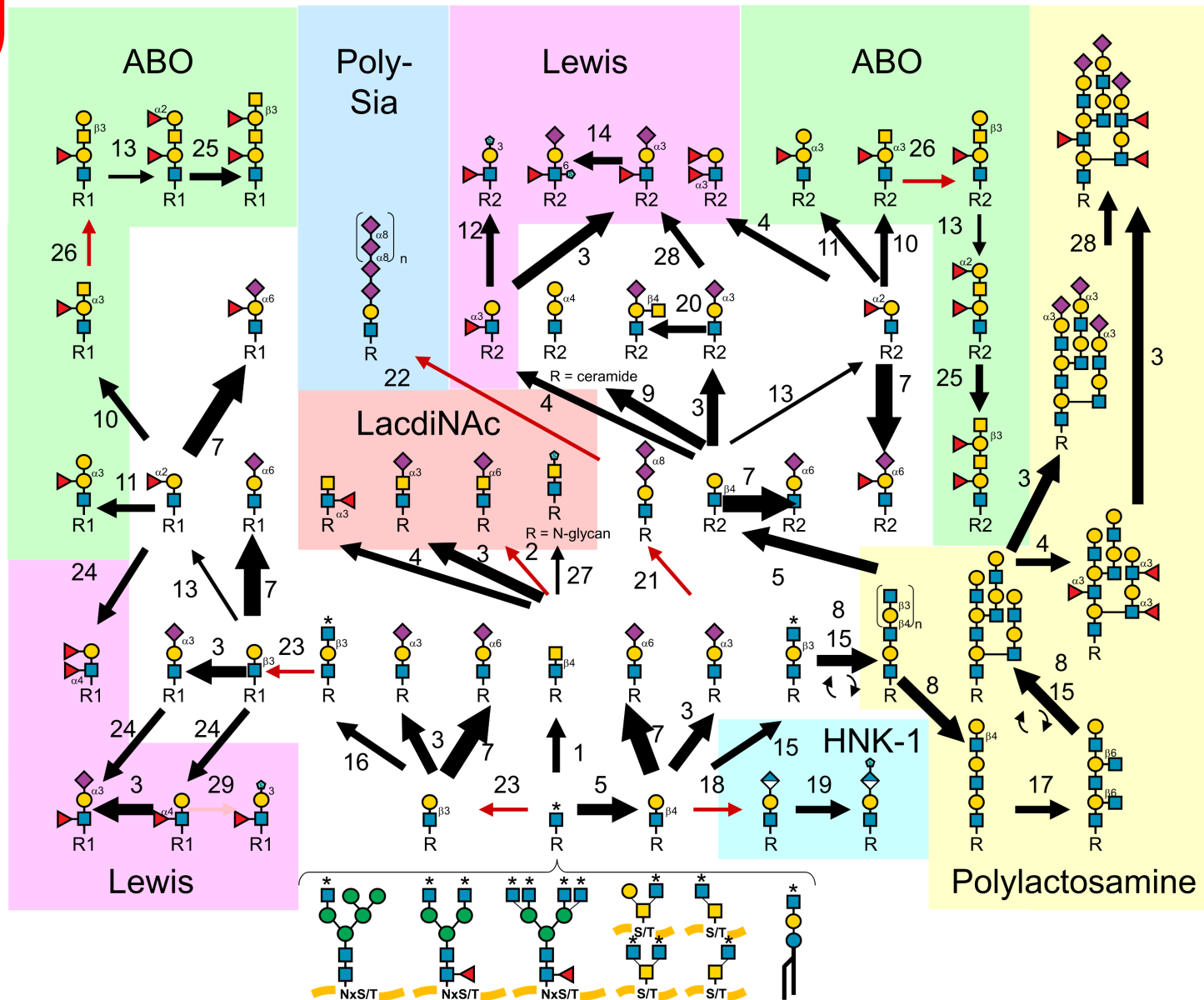
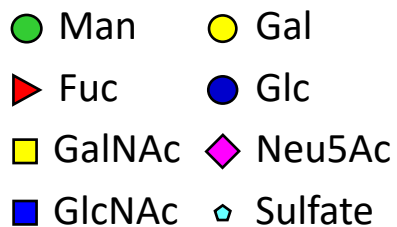
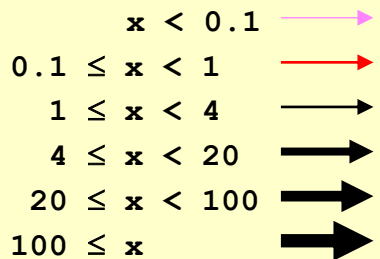
(N = 530; Median)

V.S.

pRCC

(N = 288; Median)

TPM value of gene



# Biosynthesis of capping structures in Kidney tumors

Normal tissue

(N = 140; Median)

V.S.

ccRCC

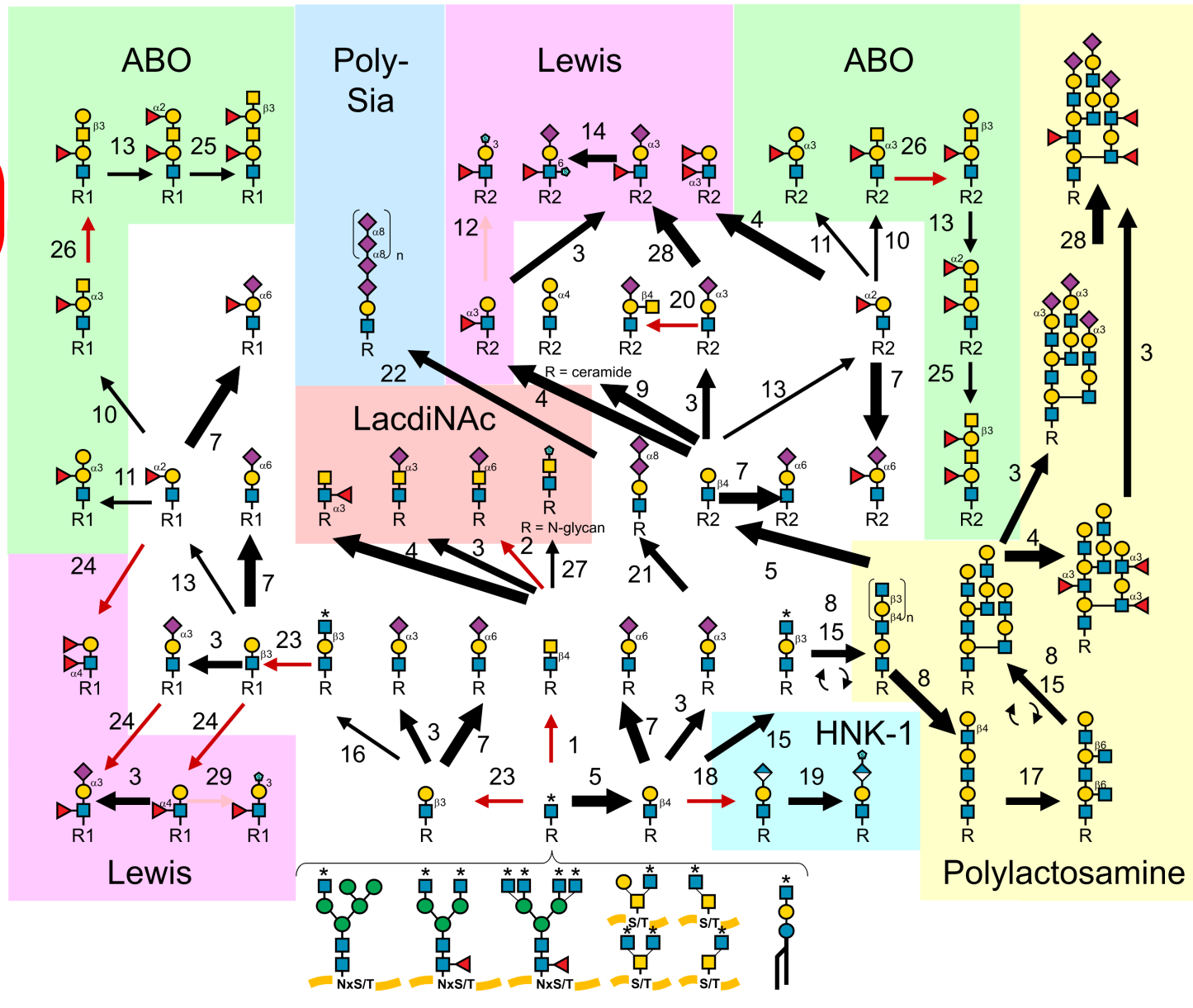
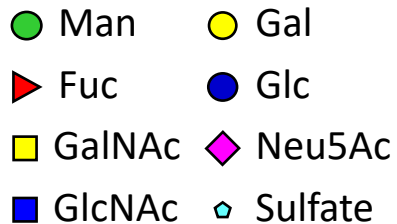
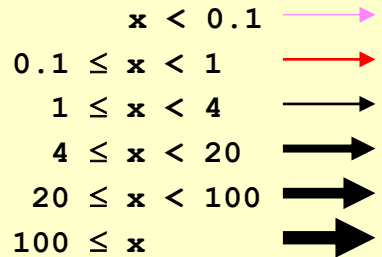
(N = 530; Median)

V.S.

pRCC

(N = 288; Median)

TPM value of gene





# Biosynthesis of capping structures in Kidney tumors

Normal tissue  
(N = 140; Median)

V.S.

ccRCC

(N = 530; Median)


V.S.

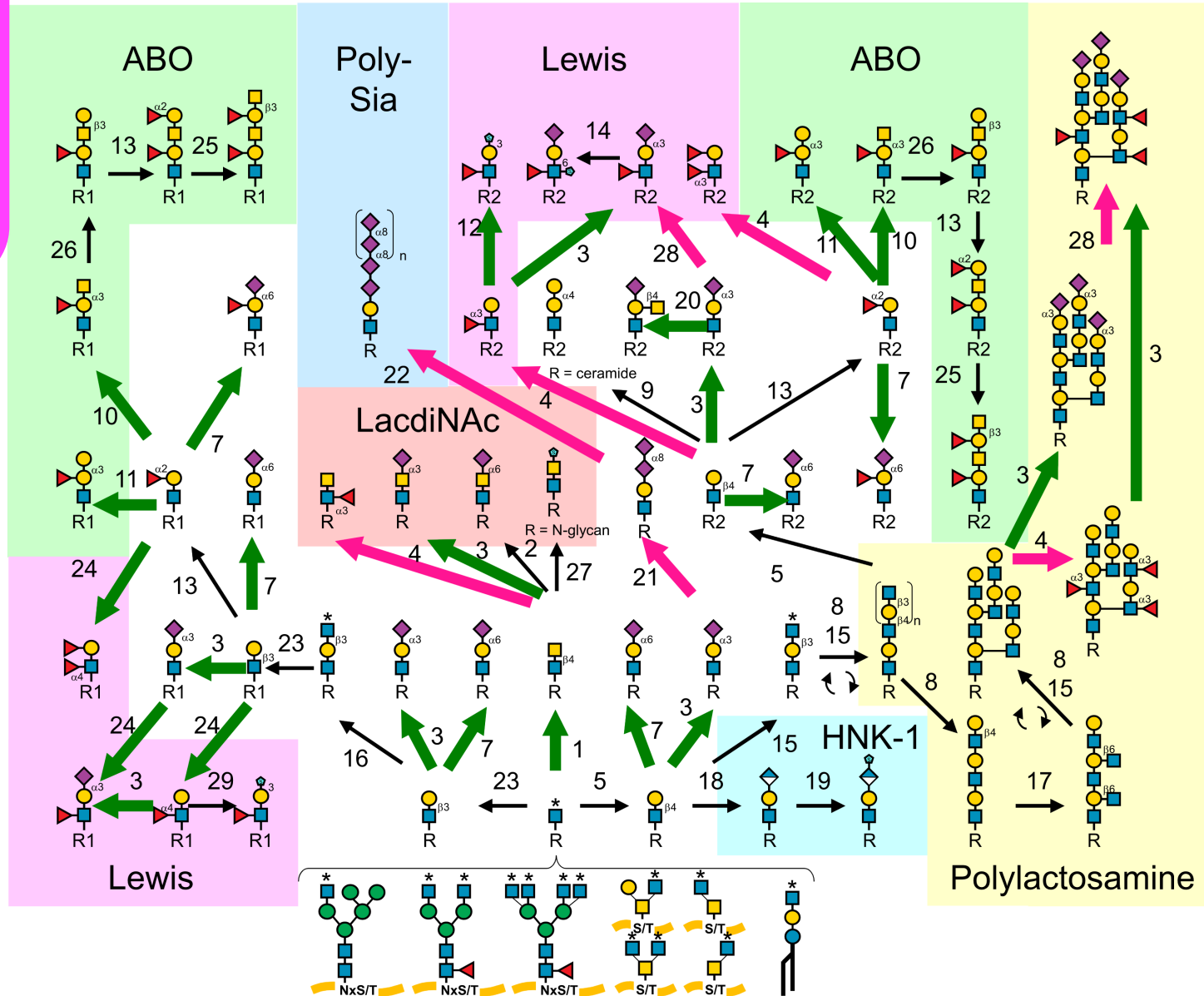
pRCC

(N = 288; Median)

$\frac{TPM_{tumor} + 1}{TPM_{normal} + 1}$

$\geq 2$  

$< 0.5$  



# Biosynthesis of capping structures in Kidney tumors

Normal tissue  
(N = 140; Median)

V.S.


ccRCC  
(N = 530; Median)

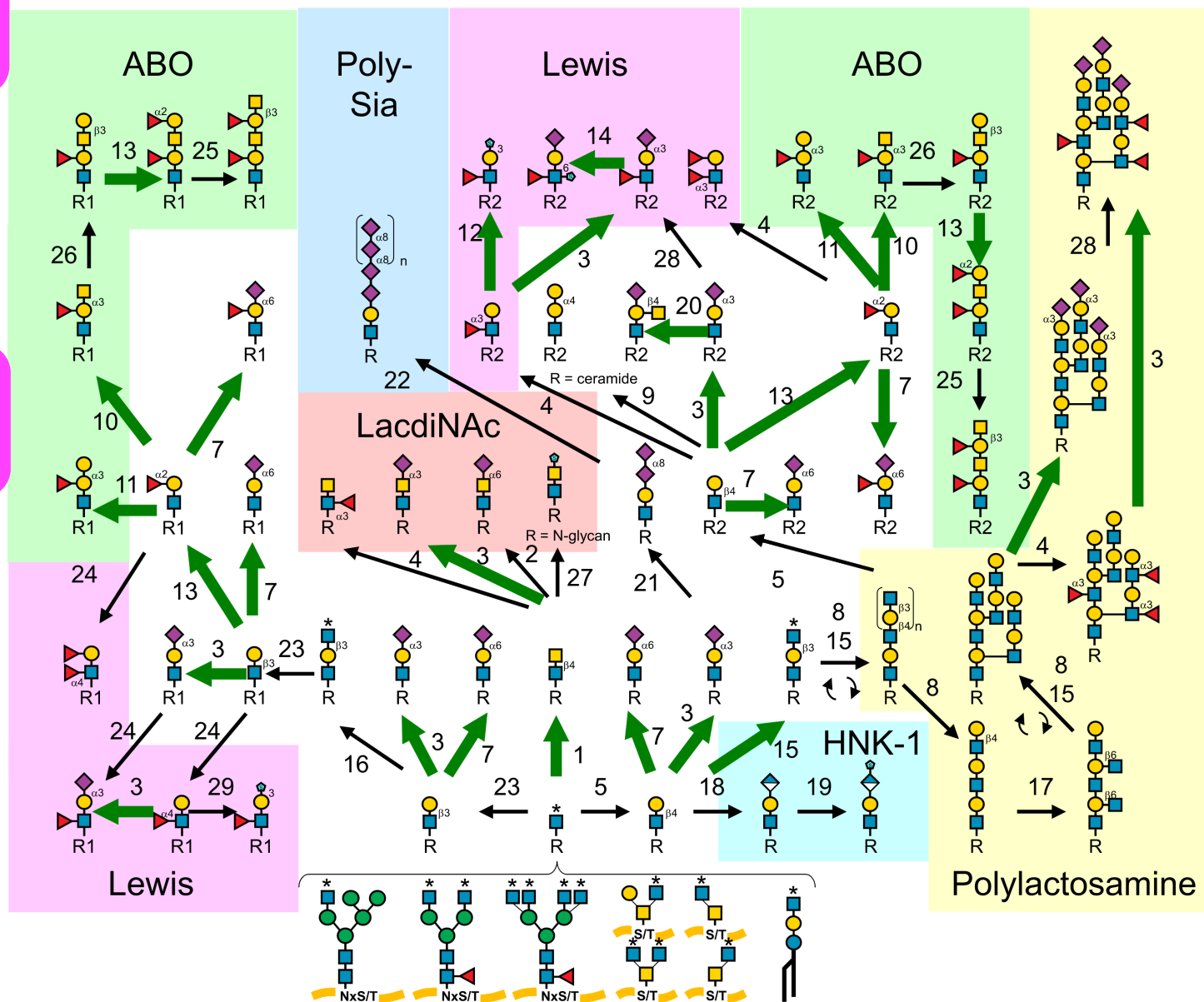
V.S.

pRCC  
(N = 288; Median)

$\frac{TPM_{tumor} + 1}{TPM_{normal} + 1}$

$\geq 2$  

$< 0.5$  



# Summary 2

We developed a mapping tool for glycosylation pathways.  
Based on gene expression, glycan structures were predicted.

## Support of glycomic analysis :

Identification of glycan structures smoothly.  
Estimation of isomers from gene expression patterns

## Engineering of glycosylation pathways :

Simplification and customization of glycans  
Based on gene expression patterns,

## Finding new insights of glycan regulation :

Complementation between N-glycans and glycolipids  
Glycan changes in normal and diseased tissues

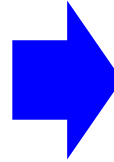
## Expansion of glycan researches :

Gene expression → Glycobiology

# Acknowledgements

**Key Laboratory of  
Carbohydrate Chemistry & Biotechnology  
Jiangnan University (2014 - 2022)**

Yi-Fan Huang



**Institute for Glyco-Core Research (iGCORE)  
Gifu University (2022 - )**



**Osaka University**

Taroh Kinoshita

**Soka University**

Kiyoko F. Aoki-Kinoshita

Sachiko Akase

**University of Georgia**

Kazuhiro Aoki (MCW, now)

Michael Tiemeyer

**AIST**

Yasunori Chiba

**Gifu University**

Yasuhiko Kizuka

**Meijo University**

Shuji Mizumoto

**Jiangnan University**

Xiao-Dong Gao



**Thank you for viewing the slides.**