

Understanding Hyaluronan-Protein Interactions

I [Introduction](#)

II [The Link Module Superfamily](#)

III [The Link Module Fold](#)

IV [The Molecular Basis of Hyaluronan-Protein Interactions](#)

V [Regulation of Hyaluronan-Protein Interactions](#)

VI [Evolution of Hyaluronan Binding Function](#)

VII [Concluding Remarks](#)

Authors' Profiles



Anthony J. Day: Tony Day is a structural biologist who has worked on hyaluronan-protein interactions for the last seven years. He obtained a B.A. in Chemistry from Oxford University in 1985 followed by a D.Phil. in Biochemistry in 1988. His doctoral research focused on the structural characterization of factor H, a control protein of the human complement system. From 1991 to 1998 he worked as an Arthritis Research Campaign Research Fellow in the laboratory of Professor Iain D. Campbell (Department of Biochemistry, University of Oxford), and it was during this time that he developed his interest in hyaluronan-binding proteins. This work led to the determination of the

first three-dimensional structure of a Link module, a domain found in many hyaladherins. Dr. Day is now a Senior Scientist at the Medical Research Council's Immunochemistry Unit (Oxford) and is engaged in research aimed at understanding the molecular basis of protein-hyaluronan interactions and the role of TSG-6 (an inflammation-associated hyaladherin) in extracellular matrix remodeling and leukocyte adhesion.

I Introduction Hyaluronan has diverse biological roles in vertebrates. For instance, it acts as a vital structural component of connective tissues,^a forms loose hydrated matrices that allow cells to divide and migrate (for example during development),^b mediates immune cell adhesion,^c and activates intracellular signalling.^{d,e} This wide range of functional activities may seem surprising given hyaluronan's simple structure, but results in fact from the large number of hyaluronan-binding proteins (often termed hyaladherins) that exhibit significant differences in their tissue expression, cellular localization, specificity, affinity and regulation. Characterization of the molecular basis of hyaluronan recognition by proteins, and how this is modulated *in vivo*, will be a key determinant in understanding the biology of this important polysaccharide.

^aSee review by Hardingham in this series.

^bSee review by Toole in this series.

^cSee review by Strong and de la Motte in this series.

^dSee review by Knudson and Knudson in this series.

^eSee review by Turley and Harrison in this series.



II The Link Module Superfamily

Many hyaluronan-binding proteins contain a common domain, termed a Link module, which is involved in ligand binding (**Figure 1**). The Link module (also referred to as a proteoglycan tandem repeat)¹ was first described in cartilage link protein, which is comprised of an immunoglobulin domain and two contiguous Link modules. This molecular arrangement is also found in the G1-domains of the chondroitin sulfate proteoglycans aggrecan, versican, neurocan and brevican. As shown in **Figure 2**, aggrecan forms huge link protein-stabilized complexes with hyaluronan that provide load-bearing function in articular cartilage.^a Similar aggregates, in which versican, neurocan or brevican substitute for aggrecan, are likely to contribute to the structural integrity of many other tissues, *e.g.*, skin, blood vessels and brain. The immunoglobulin domains are probably responsible for the link protein-proteoglycan interaction, while the Link modules mediate binding to hyaluronan.

^a See review by Hardingham in this series.

Link module superfamily

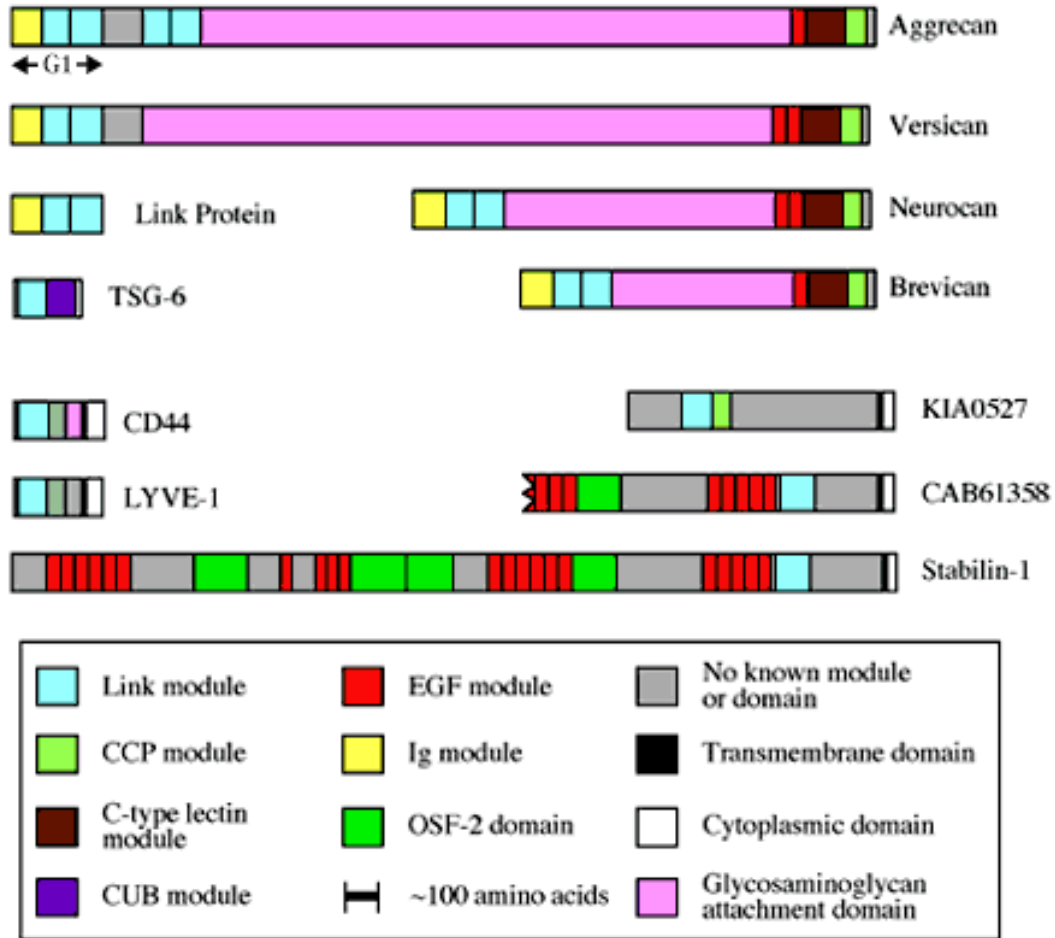


Figure 1.
The Link module superfamily.

The ubiquitous hyaluronan receptor CD44 has diverse functions including the attachment, organization and turnover of extracellular matrix at the cell surface (*e.g.*, in cartilage; [Figure 2](#)) and mediates the migration of lymphocytes during inflammation.^d CD44 can exist in numerous isoforms, due to alternative splicing of 10 variant exons in different combinations, all of which contain a single Link module close to the N-terminus of the protein. Apart from CD44, the only other member of the Link module superfamily ([Figure 1](#)) that has been clearly identified as a hyaluronan receptor is LYVE-1, a recently discovered protein that is restricted in its expression to lymph vessel endothelium and that may be involved in hyaluronan catabolism.²

^d See review by Knudson and Knudson in this series.

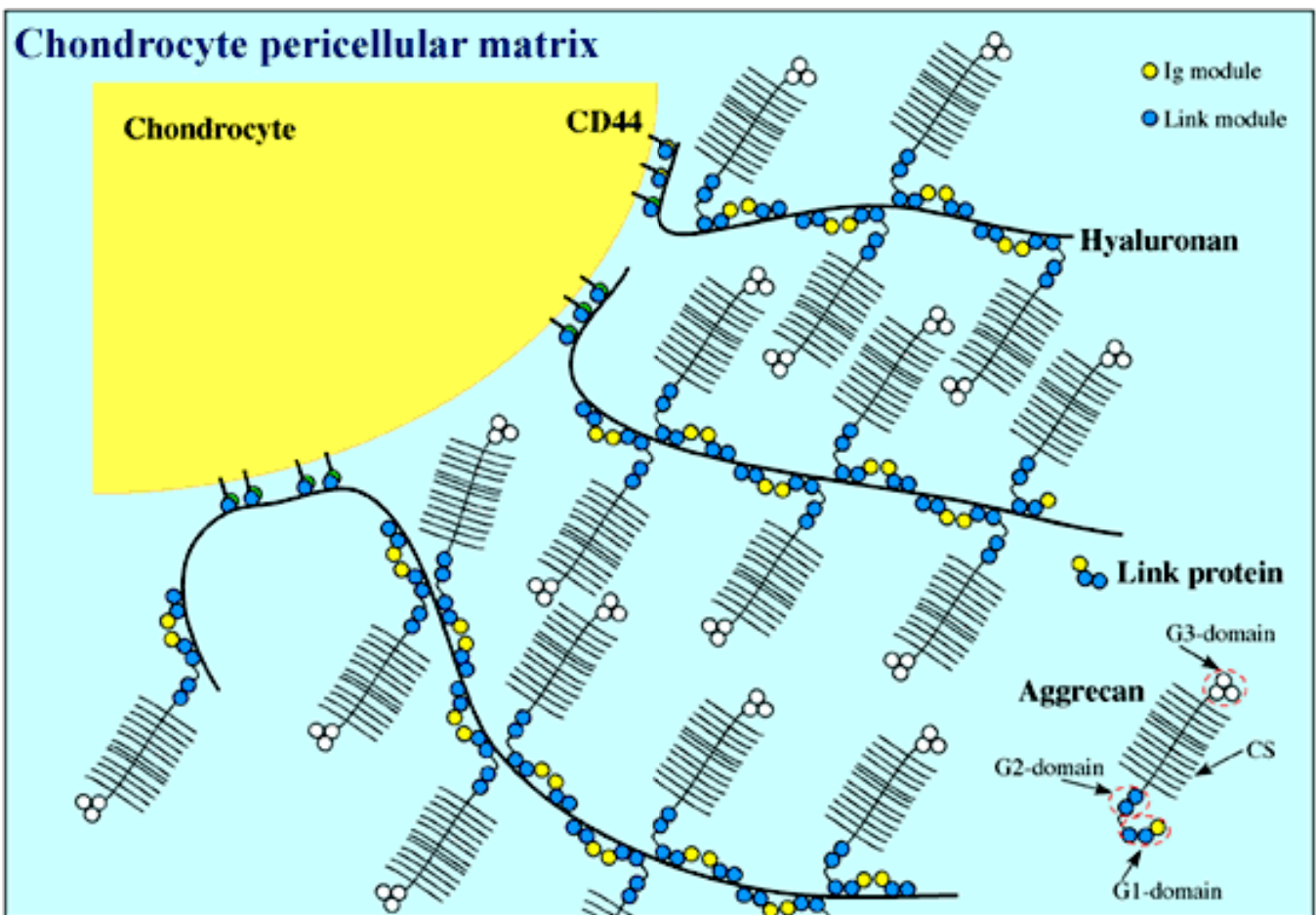


Figure 2. Chondrocyte extracellular matrix is composed of huge multi-molecular complexes consisting of an alternating array of link protein and aggrecan molecules along a central hyaluronan filament. These aggregates, some of which are anchored to the chondrocyte surface via the hyaluronan receptor CD44, provide cartilage with its load-bearing properties. Aggrecan is a proteoglycan made up of three globular domains (G1, G2 and G3) and a region where ~100 chondroitin sulfate (CS) chains are attached.

The protein product of tumor necrosis factor-stimulated gene-6 (TSG-6), which contains a single Link module, is secreted in response to inflammatory stimuli (*e.g.*, in the articular joints of arthritis patients).³ TSG-6 has been implicated in the regulation of leukocyte migration, and the pattern of its expression suggests that it is likely to be involved in extracellular matrix remodeling. For example, TSG-6 in cartilage may inhibit the formation of hyaluronan-protein aggregates, promote aggregate dissociation or modulate CD44 function (Figure 3).^{4,5} In certain tissue locations (*e.g.*, during the expansion of cumulus cell-oocyte complexes), TSG-6 can associate with the inter- α -inhibitor (I α I), and this TSG-6/I α I complex may be able to stabilize extracellular matrix by cross-linking of hyaluronan chains.^f Although the role of TSG-6 protein is poorly understood, its hyaluronan-binding properties, and their structural bases, are probably the best characterized of any hyaladherin (see below).

f See Salustri and Fülöp in this series; O Carrette, R V Nemade, A J Day, A Brickner and W J Larsen, manuscript submitted.

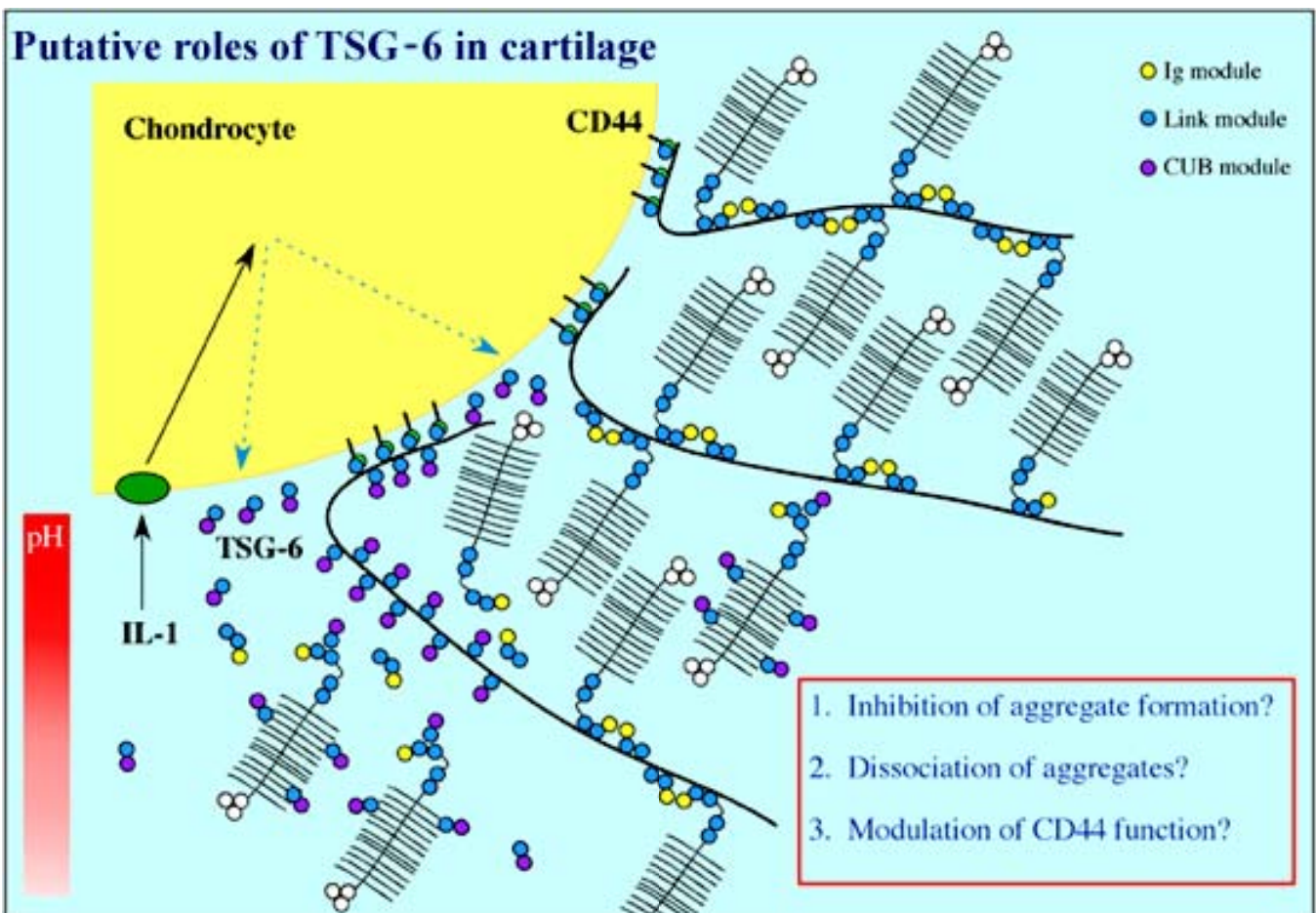


Figure 3. Putative roles of TSG-6 in cartilage. TSG-6, which is secreted by chondrocytes during inflammation (e.g., in rheumatoid arthritis), is likely to be involved in extracellular matrix remodeling, as it can interact with components of cartilage such as hyaluronan, chondroitin-4-sulfate and the G1-domain of aggrecan.^{4,5} For instance, TSG-6 may inhibit the formation of cartilage aggregates, blocking their assembly by binding to the G1-domain and the chondroitin-4-sulfate chains of *de novo* synthesized aggrecan. It could also be involved in the dissociation of existing aggregates, possibly by altering the structure of hyaluronan, and the regulation of CD44 function. The hyaluronan- and aggrecan-binding activities of TSG-6 are pH-dependent; this may result in regulation of its functions via pH gradients that exist in articular cartilage. In this regard, there is likely to be a significant drop in pH in the peri-chondrocytic region during inflammation.⁴

Figure 1 shows several new members of the Link module superfamily (*i.e.*, stabilin-1, CAB61358 and KIA0527) that are all likely to be cell surface proteins. Stabilin-1 (accession code AJ275213) and CAB61358 (which is the accession code for this incomplete sequence) have very similar domain organizations, and the Link modules of these proteins both show about 50% identity with that of TSG-6. However, it is unclear whether or not these are hyaladherins, as no data on their hyaluronan-binding properties are yet available. The same is true for KIA0527 (accession code AB011099), which was cloned from brain tissue.

As can be seen from **Figure 4**, there is a growing number of hyaluronan-binding proteins that do not contain a Link module. Most of these are unrelated to each other at the primary sequence level. There are few data on the positions of the hyaluronan-binding sites in these proteins, apart from the receptor RHAMM,^e and therefore they will not be covered in this review.

^e See review by Turley and Harrison in this series.

Other hyaluronan-binding proteins

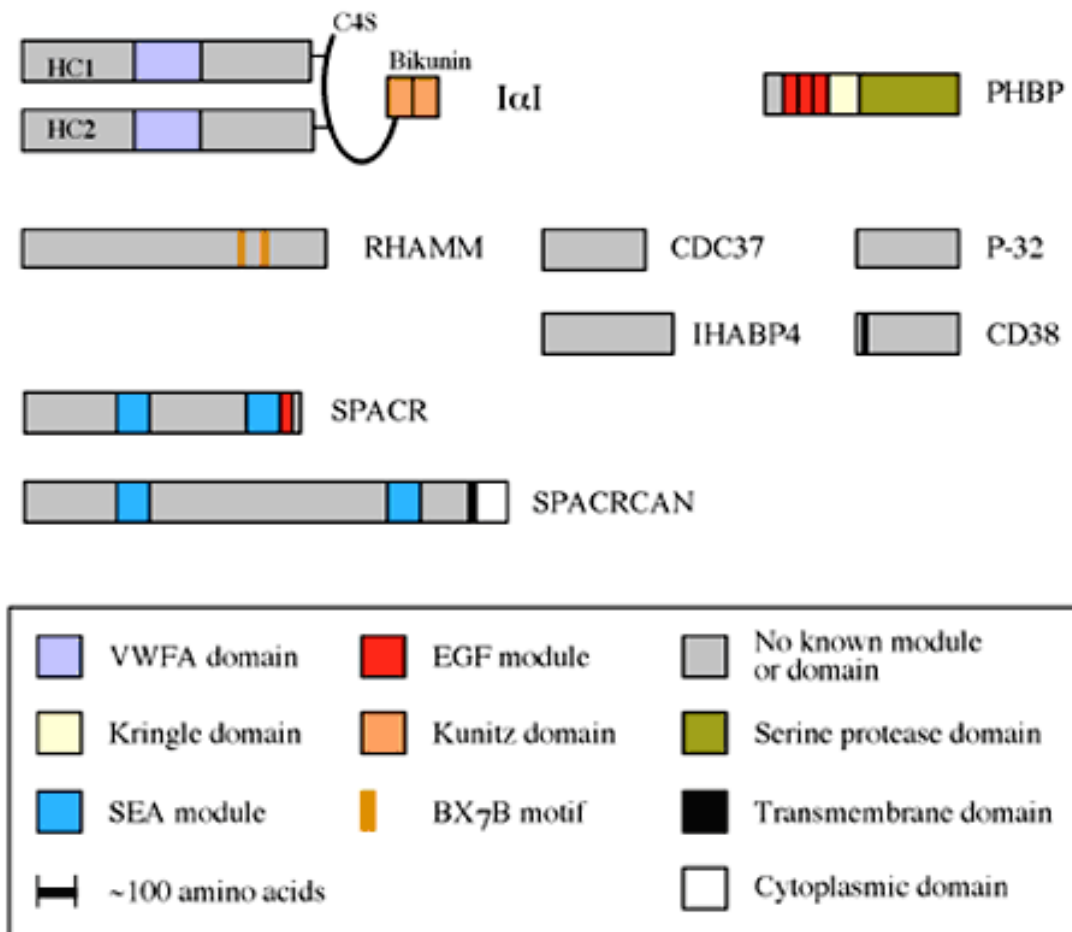


Figure 4.

Other hyaluronan binding proteins. The BX₇B motif is shown only for RHAMM, for which there is evidence of its involvement in hyaluronan binding.

III The Link Module Fold The Link module is a domain of approximately 100 amino acids with four cysteines disulfide-bonded in the pattern Cys1-Cys4 and Cys2-Cys3 (Figure 5). The three-dimensional structure of the Link module from human TSG-6 has been determined by nuclear magnetic resonance spectroscopy in solution.¹ It is comprised of two α -helices and two triple-stranded anti-parallel β -sheets (Figure 6). As can be seen from the alignment in Figure 5, many residues (including the four cysteines) are highly conserved across the Link module superfamily. There are no insertions or deletions present in regions corresponding to elements of secondary structure, apart from the bulge in the β 5 strand, and those that do occur would mainly affect the length of exposed loops. This structure therefore has defined the ‘consensus fold’ for the Link module superfamily as a whole. In practical terms, this means that the TSG-6 Link module co-ordinates can be used as a template for useful modeling of other superfamily members.^{6,7} A good test of this is the Link module from KIA0527, which despite having an unusual

arrangement of cysteine residues (Figure 5), was successfully modeled, as shown in Figure 7.

Alignment of Link module sequences

LP1	VVFYPPRLGRYNLNFHEAQQACLDDQDAVIASFQQLYDAWRG	GLDHCNAGHLSDGSVQYPIITKPREPCGGQN	TVPGVRNY	GFWDKDKSRYDVFCFTSNF									
LP2	GRFYVLIHPTKL TYDEAVQACLNDGAQIAKVGQIPAAWKIL	GYDRCDAGNLADGSVRYPISRPRRRCSPTE	AAVRFV	GFPDKKHKLYGVYCFRAYN									
A1	IVFHYRAISTRYTLDFDRAQRAQLQNSAIIATPEQLQAAYED	GFHQCDAGNLADQTVRYPIHTPREGCVGDKDEFPGVRTY		GIRDTN ETYDVYCFAEEM									
A3	VVFHYRPGPTRYSLTFFEEAQQACPGTGAVIASPEQLQAAYEA	GYEQCDAGHLRDQTVRYPIVSPRTPCVGDKDSPPGVRTY		GVRPST ETYDVYCFVDRL									
B1 (R)	VVFLYREGSARYAFSPAGAQAECARIGARIATPEQLYAAYLG	GYEQCDAGHLSQDQTVRYPIQNPRAECYGDMDGYPGVRNY		GVVGPD DLYDVYCYAEDL									
N1	VVFHYRSARDRYALTFAEAQAECRLSSAIIAAPRHLQAAFED	GFDCNCDAGHLSQRTVRYPIQSRPGCYGDRSSLPGVRSY		GRRNPQ ELYDVYCFAREL									
V1	VVFHYRAATSRYTILNFEEAQAACLDDVGAVIATPEQLPAAYED	GFEQCDAGHLADQTVRYPIRAPRVGCVGDKMGKAGVRTY		GFRSPQ ETYDVYCYVDHL									
A2	EVFYAT SPEKF TFQEAANECRRLGARLATTGHVYLAWQA	GMDHCSAGNLADRSVRYPIISKARPNCGGNL	LGVRTVYVHANQTGYDPDS	SRYDAICYTGED									
A4	EVFFAT RLEQF TFQEALEFCESHNAT ATTGQLYAAWSR	GLDKCYAGHLADGSLRYPIVTPRPACGGDK	PGVRTVYLYPNQTLGPDPL	SRHBACFRGIS									
B2 (R)	ELFLGA PPGKL TWEEARDYCLERGAQIASTGQLYAAWN	GLDRCSFGWADGSVRYPIITPSQRCSGGGL	PGVKTFLFFNQTFPPSKQ	NRFNVYCFRDSA									
N2	EVFYVG PARRL TLAGARAQCRROGAALASVQQLHLAWHE	GLDQCDPGLHLDGSVRYPIQTPRRRCGGPA	PGVRTVYRFANRTGFPSPA	ERFDAYCFRAHH									
V2	DVFFLT VPSKF TFEEAAKCEENQDARLATVGEQLQAAMRN	GFDCQDYGHLSDASVRHPVTVARAQC GGGL	LGVRTLYRFENQTFPPPD	SRFDAYCFKRRM									
CD44	GVFHVE KNGRYSISRTEAADLCKAFNSTLPTMAQNEKALS	IGFETCRYGFI EGHVVIPRIHPNSICAANN	TGVY	ILTSNTSQYDTCFNASA									
LYVE-1	GITLVS KRANQQLNFTEAKEACRLLGLSLAGKQVETALK	SPFETCSYGNVGDGFVVISRISPNPKCGKNG	VGVL	NKVPVSRQFAAYCYNSSD									
TSG-6	GVYHREARSGKYKLTIAEAKAVCFEFGGHLATYKQLEAARK	IGFHVCAAGNMAKGRVGYPIVKPGPNCGFGK	TGIIDY	GIRLNRSERWDAYCYNPHA									
STAB-1	GVFHLQATSGPYGLNPFSEAEAAACEAQAVLASFPQLSAAQQ	LGFHLCLMGNLANGSTAHVVFVADCGNGR	VGIVSL	GARKNLSERWDAYCFRVQD									
61358	GVFHLRSPLGQYKLTFDKAREACANEATMATYNQLSYAQA	KYHLCASAGHLETGRVAYPTAFASQNC GSGV	VGIVDY	GPRPNKSEMWDVFCYRMKG									
0527	KLFVLESQNGSQGLQLEAARLSCKSRGAHLASADELRVVVQDC	SFAVCTTGNLADGTLGTTVC SKSGSGEQQIM	RAVDVRIE	SNPVFGGTYSALCIKDEE									
SS	bbbbbb	aaaaaaaaaa	bbbb	aaaaaa	bbb	bbbbbb	bb^^	^bb	bbbbbb				
CON	VF	F EA	C	A A	QL	G	C	GML D V YPI P	C	GVR	G	D	YCF

Figure 5. Multiple sequence alignment of Link modules from the proteins in Figure 1. LP, link protein; A, aggrecan; B, brevican; V, versican; STAB-1, stabilin-1; 61358, CAB61358; 0527, KIA0527. All the sequences are from human except for brevican which is from rat (R). Cysteine residues are shown in yellow and other highly conserved residues (*i.e.*, identical in 2/3 of the sequences) are denoted in pink. These residues comprise the Link module consensus sequence (CON). The position of the secondary structure elements (SS), determined for TSG-6,¹ are shown below the alignment (a = α -helix, b = β -strand, ^ = bulge).

[Click here](#) enlarged Figure 5.

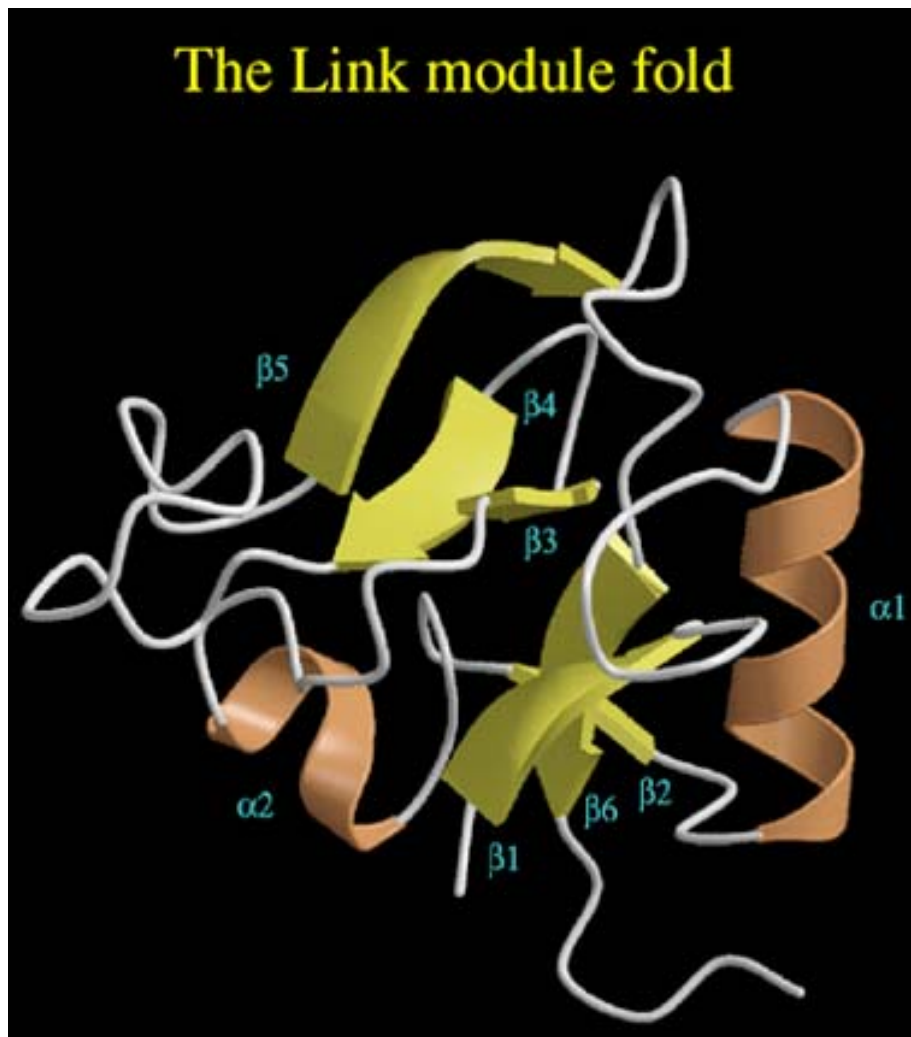


Figure 6.

The Link module fold. The structure consists of two α -helices (brown) and two triple-stranded β -sheets (yellow). Sheet I is comprised of strands $\beta 1$, $\beta 2$, $\beta 6$, while sheet II is made up from strands $\beta 3$, $\beta 4$ and $\beta 5$.

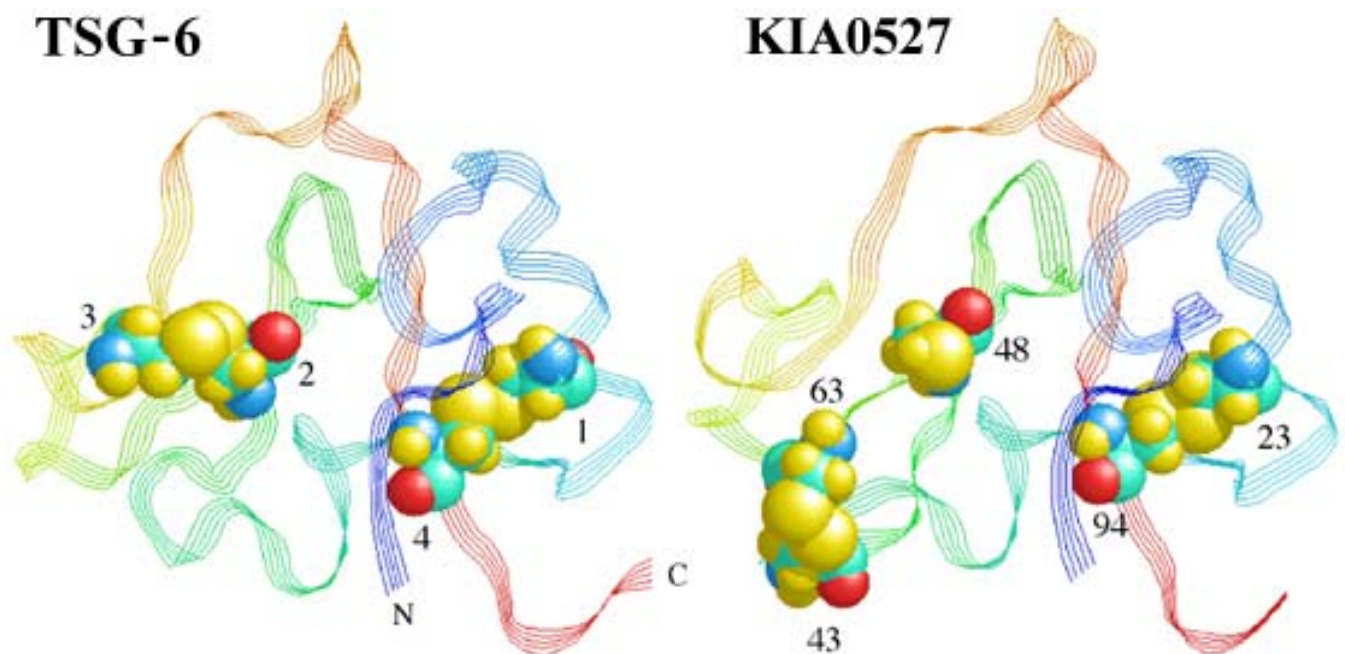


Figure 7.

Link modules of TSG-6 and KIA0527. The Link module of KIA0527 was modeled on the basis of the TSG-6 Link module co-ordinates¹ and the pairwise alignment of these proteins from Figure 5 (A J Day, unpublished work). TSG-6 is a typical member of the Link module superfamily with disulfide bridges between consensus Cys1-Cys4 and Cys2-Cys3. In the model of KIA0527, consensus Cys-2 (residue 48) is a free sulfhydryl, due to consensus Cys-3 being absent, while the two additional cysteines (residues 43 and 63) are able to form a disulfide bond. The fact that this unusual arrangement of cysteines can be accommodated into the TSG-6 structure is a good indicator that this does represent the consensus fold for the Link module.

The recombinant domain used to solve the tertiary structure was found to be functionally active,^{1,4,5,8} indicating that in TSG-6 a single Link module is sufficient for high-affinity hyaluronan binding. This is not the case for CD44, which has a hyaluronan-binding domain of ~160 amino acids made up of a single Link module with additional N- and C-terminal extensions linked by a disulfide bond. These extensions are required for correct folding and functional activity (Figure 8).^{9,10} A larger domain still is likely to be utilized by link protein, aggrecan and the other aggrecan-like proteoglycans, as these proteins all have a pair of contiguous Link modules in their hyaluronan-binding regions (Figure 1). In link protein and aggrecan, both Link modules participate in ligand binding.^{11,12} Therefore, the proteins of the Link module superfamily can be divided on the basis of the size of their hyaluronan-binding domains into three sub-groups,⁷ as illustrated in Figures 9 and 10. The sizes of these domains appear to correlate broadly with the length of hyaluronan recognized. For example, hexasaccharides (HA₆) and deca-saccharides (HA₁₀) are the minimum lengths of hyaluronan required for high-affinity binding to Type A and Type C domains, respectively.⁸ Hyaluronan binding to CD44 can be displaced by either HA₆ or HA₁₀, depending on the cell background on which it is expressed.^{13,14}

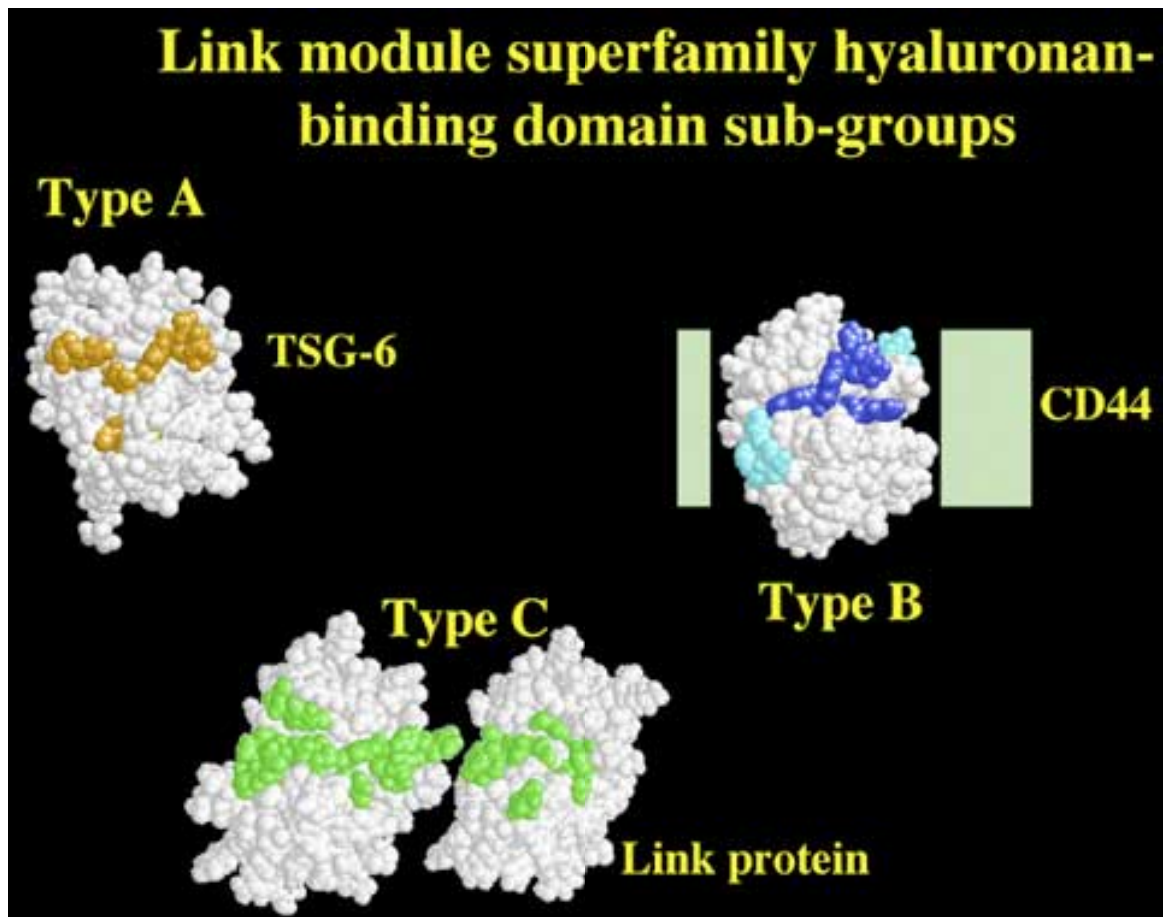


Figure 10.

The structures/models of the Type A, B and C hyaluronan-binding domains of TSG-6, CD44 and link protein. Only the tertiary structure of the Type A domain, a single Link module from TSG-6, has been determined to date. Residues that mediate the interaction with hyaluronan are colored in orange.⁸ The Link modules shown for CD44 and link protein were modeled on the basis of the TSG-6 co-ordinates. In CD44 the structures of the N- and C-terminal extensions (light green boxes) are currently unknown. However, these are likely to form a sub-domain in intimate contact with the Link module.⁷ Amino acids of the CD44 Link module that are either critical or important for ligand binding are depicted in dark blue or light blue, respectively.⁶ In the Type C domain, the relative orientations of the two Link modules remain to be determined, and only one possible configuration is illustrated. The residues of link protein predicted to be involved in the interaction with hyaluronan are colored green (A J Day, unpublished).

Residues of CD44 involved in hyaluronan binding have been determined by site-directed mutagenesis.^{6,9} As can be seen from **Figure 8**, functionally important amino acids are present both on the Link module and on the flanking sequences. The ligand-binding residues (in the Link module) form a continuous surface on one face of the molecule (**Figure 11**). The same face of the Link module is involved in hyaluronan binding in TSG-6, indicating that the position of the ligand interaction surface may be conserved across the superfamily.⁸ From these studies, it is clear that in Link module-containing proteins, binding residues are brought together from different parts of the sequence and a folded structure is necessary to generate a ligand-binding surface (**Figure 12**). Linear sequence motifs (*e.g.*, BX₇B peptides found in RHAMM)^e do not appear to be involved in Link module-hyaluronan interactions.⁸

^e See review by Turley and Harrison in this series.

^g DJ Mahoney and AJ Day, manuscript in preparation.

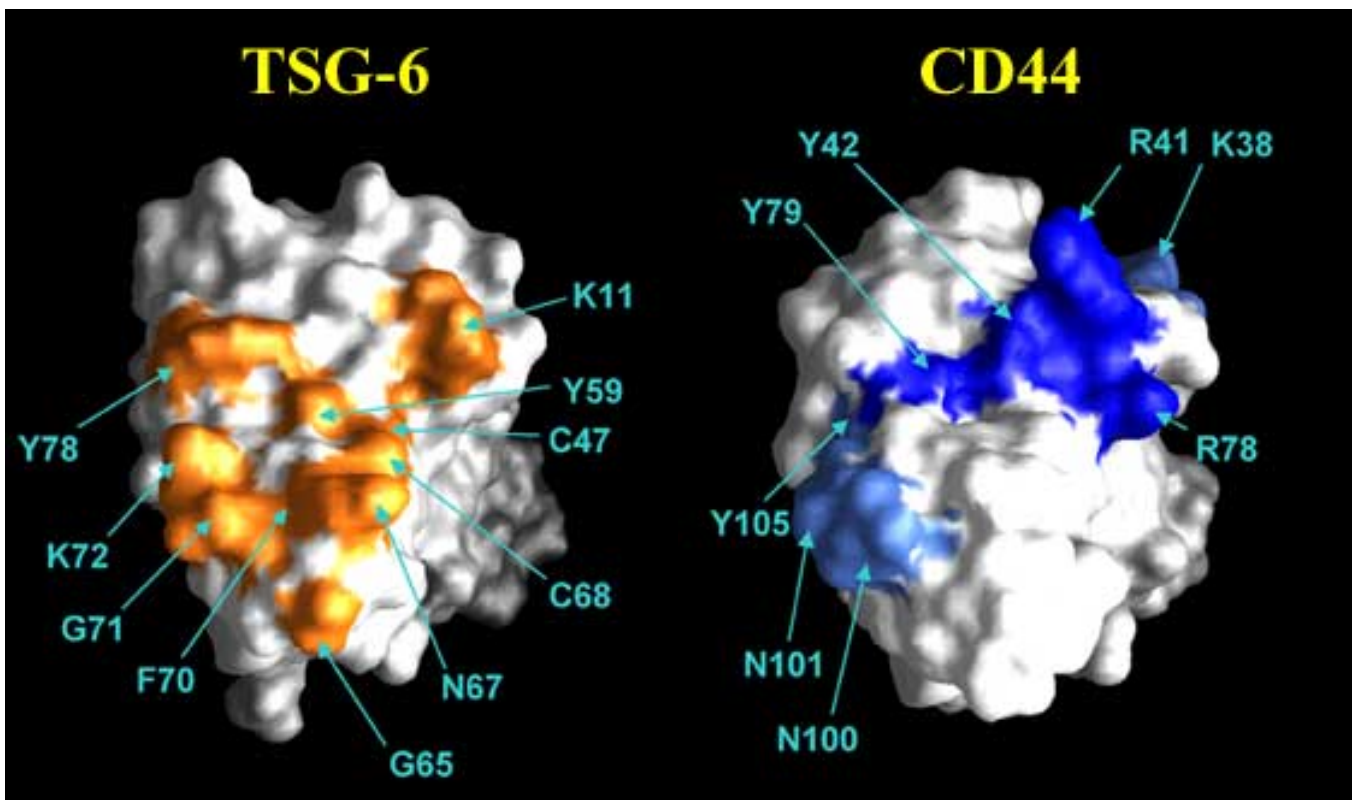


Figure 11.

Hyaluronan-binding sites of TSG-6 and CD44. The hyaluronan-binding sites of these proteins are on the same face of the Link module. Surface-exposed amino acids on the TSG-6 Link module that are perturbed on HA₈ binding are shown in orange.⁸

Site-directed mutagenesis has allowed residues directly involved in the interaction with hyaluronan to be distinguished from those affected due to ligand-induced conformation changes.^g Amino acids of the CD44 Link module that are either critical or important for ligand binding are depicted in dark blue or light blue, respectively.⁶

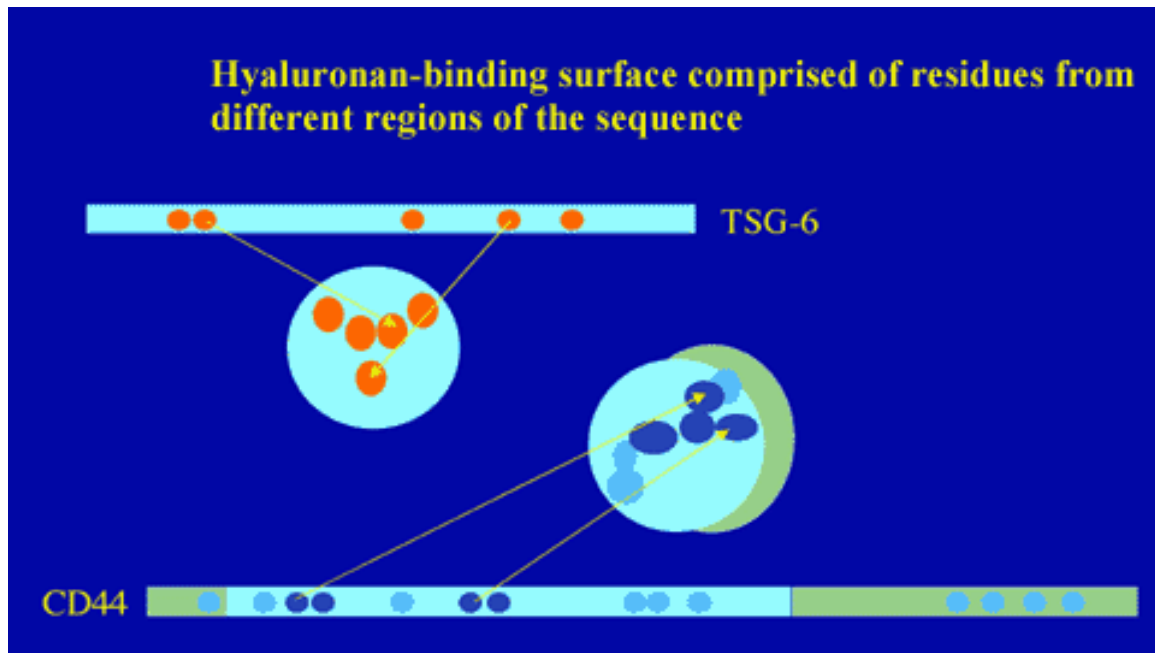


Figure 12.

The hyaluronan-binding surface is comprised of residues brought together from different parts of the protein sequence.

Although, the hyaluronan-binding sites in TSG-6 and CD44 have similar locations on the Link module surface (Figure 11), there appear to be some major differences in the details of the residues and sequence positions involved in binding in the two proteins. Comparison of the Link module sequences of CD44 and TSG-6 with all the other members of the superfamily indicates that the molecular details of hyaluronan-binding (*i.e.*, the interaction networks) are likely to be distinct in each protein. This is perhaps not surprising given the different sub-types of hyaluronan-binding domain (described above) as well as many other differences in specificity and regulation. For instance, TSG-6 binds to chondroitin-4-sulfate with high affinity, but this glycosaminoglycan is not recognized by aggrecan or link protein.⁵

IV *The Molecular Basis of Hyaluronan-Protein Interactions*

Chemical modification studies have indicated that basic amino acids are major determinants in the binding of aggrecan and link protein to hyaluronan by forming ionic bonds with the carboxylic acid groups of the glycosaminoglycan.⁸ This is also likely to be the case for CD44, for which two arginines (Arg41 and Arg78) play a critical role in ligand binding.^{6,9} In addition, two tyrosines (Tyr42, Tyr79) are essential for CD44 functional activity (Figure 11); mutation of Tyr79 to phenylalanine destroys functional activity, indicating that its hydroxyl group makes a crucial hydrogen bond to the sugar.⁶ Other amino acids of CD44, including three lysines, four arginines, two asparagines and a tyrosine (shown in orange on Figure 8), are also implicated in hyaluronan binding but have a less critical involvement. The large number of residues engaged in binding indicates that an extensive network of interactions (particular to CD44) maintains its association with hyaluronan. The energetics of this are clearly finely balanced, as loss of a single hydrogen bond or ionic interaction can be enough to abolish binding. This is also true for TSG-6 in which five amino acids have been identified as having an important functional role, including basic and aromatic residues.^g To date, the thermodynamics of hyaluronan-protein binding have been determined only for the TSG-6 Link module, in which the major energetic contribution driving the interaction is a large and exothermic enthalpy.⁸ The small change in heat capacity seen on binding is consistent with an intermolecular interface comprising a significant polar or charged component rather than the burial of a large hydrophobic surface area. While it is clear that ionic associations are important, non-ionic interactions (*e.g.*, hydrogen bonding and van der Waals forces) are also likely to make a significant contribution to the binding energy.¹⁵ This is consistent with data from site-directed mutagenesis showing that in addition to a lysine, 3 tyrosines and a phenylalanine have a critical role in binding.^g

g DJ Mahoney and AJ Day, manuscript in preparation.

Clusters of basic amino acids, referred to as BX₇B motifs (where B is either lysine or arginine and X can be any amino acid apart from acidic residues), have been implicated in the interaction between hyaluronan and RHAMM.^e Although BX₇B-like sequences are present in many other hyaladherins (*e.g.*, IαI, SPACR, SPACRCAN, CD38, CDC37, P-32 and IHABP4; Figure 4), there are no data to indicate that they mediate hyaluronan binding in these proteins. This spacing of basic amino acids is extremely common in protein sequences. Therefore, the presence of a BX₇B motif should not be interpreted as an indicator that a protein will bind hyaluronan, nor should it be assumed that this is necessarily the site of ligand-binding activity in a hyaladherin.

e See review by Turley and Harrison in this series.

V Regulation of Hyaluronan-Protein Interactions

Unlike other members of the Link module superfamily, the interaction of TSG-6 with hyaluronan is highly pH-dependent (Figure 13), with maximal binding at pH 6 and a dramatic reduction in activity as the pH is increased.⁴ This may serve to regulate the ligand-binding functions of this protein in tissues that have significant pH gradients, for example, in inflamed cartilage where TSG-6 is expressed (Figure 3). The structural basis of this unusual pH-dependency is not yet fully understood. However, it is clear that no detectable alteration in the conformation of the TSG-6 Link module fold occurs between pH 6 and 8.¹⁵ One possible explanation is that raising the pH changes the charge-state on a histidine residue, which leads to the loss of a critical binding interaction.

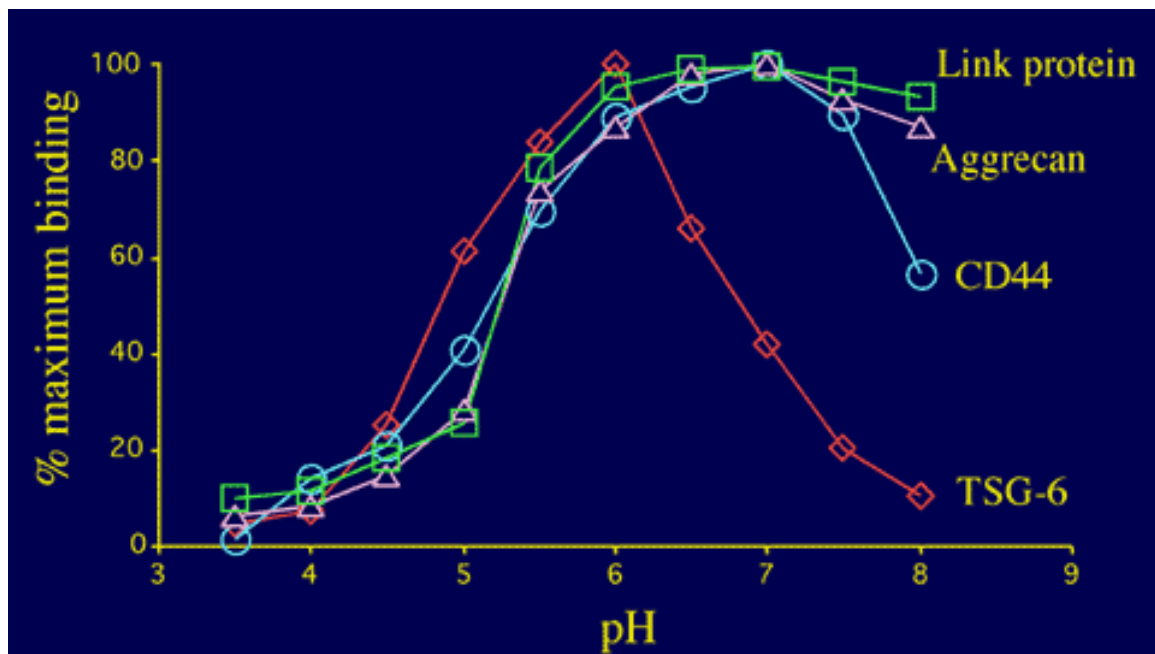


Figure 13.

Effect of pH on the hyaluronan-binding activity of four members of the Link module superfamily. The pH-dependency of the interaction between the TSG-6 Link module and hyaluronan is distinct from those of CD44 (A A Parker, S Banerji, D G Jackson & A J Day, unpublished data), aggrecan and link protein.⁴

CD44 is present on many cell types (*e.g.*, lymphocytes) in a non-functional form that requires some activation process to acquire hyaluronan-binding capabilities. The regulation of CD44 activity is a complex issue with many factors potentially involved. These include the cell background on which CD44 is expressed, post-translational modifications (*e.g.*, glycosylation), the splice isoform involved, cytoskeletal attachment, molecular clustering/receptor density on the cell surface and phosphorylation of the intra-cytoplasmic domain.^{d,14,16} Many of these have already been reviewed in this series,^d and therefore only the role of glycosylation will be discussed here. As shown in Figure 8, the hyaluronan-binding domain of human CD44 contains six potential sites for N-linked carbohydrate attachment. Expression of this region in *E. coli* in a non-glycosylated form¹⁰ and site-specific mutagenesis¹⁶ indicate that glycosylation is not required at any of these positions for either correct folding or functional activity. It is, however, one of the principal mechanisms

modulating the interaction between CD44 and hyaluronan, and there is differential glycosylation of the receptor depending on cell type and cell activation state. Removal of N-glycans by mutation of the first or fifth glycosylation sites, which are on the sequences flanking the Link module, can switch CD44 on an "inducible" cell background into a constitutively active form.¹⁶ This may indicate that particular carbohydrate structures are able to obscure the hyaluronan-binding site. Another possibility is that certain N-glycans prevent receptor clustering or lock the protein in a non-binding conformation. The molecular basis of CD44 regulation should become clearer once the tertiary structure of the hyaluronan-binding domain is known. For instance, this will reveal the positions of the glycosylation sites relative to the hyaluronan interaction surface.

^d See review by Knudson and Knudson in this series.



VI Evolution of Hyaluronan Binding Function

The Link module has a very similar fold to that of the C-type lectin domain.¹ From **Figure 14**, it can be seen that these domains have identical topologies, with similar secondary structural elements and many residues in common stabilizing the hydrophobic core. Although the mode of carbohydrate binding is clearly distinct in these domains (*i.e.*, the C-type lectin binds in a calcium-dependent manner, whilst the Link module-hyaluronan interaction is independent of metal ions), the positions of the ligand-binding surfaces are very similar.⁶ Both domains also mediate leukocyte rolling on the vascular endothelium during inflammation. Therefore, it seems very likely that they have a common evolutionary origin. The C-type lectin domain is present in many invertebrate proteins, whereas the Link module has only been identified in vertebrates, including sharks.¹⁷ This indicates that the Link module probably evolved from the C-type lectin domain sometime prior to the divergence of the cartilaginous fish and the bony fish/land vertebrates about 400 million years ago. Interestingly, Lee and Spicer¹⁸ have hypothesized that the ability to synthesize hyaluronan is a comparatively recent innovation in the evolution of metazoan organisms. For example, while a hyaluronan synthase has been found in the chordate *Amphioxus*, there are no such enzymes encoded in the *Drosophila* genome.^h Therefore, it is possible that the Link module and hyaluronan appeared at approximately the same time during evolution.

^h See review by Spicer and McDonald in this series.

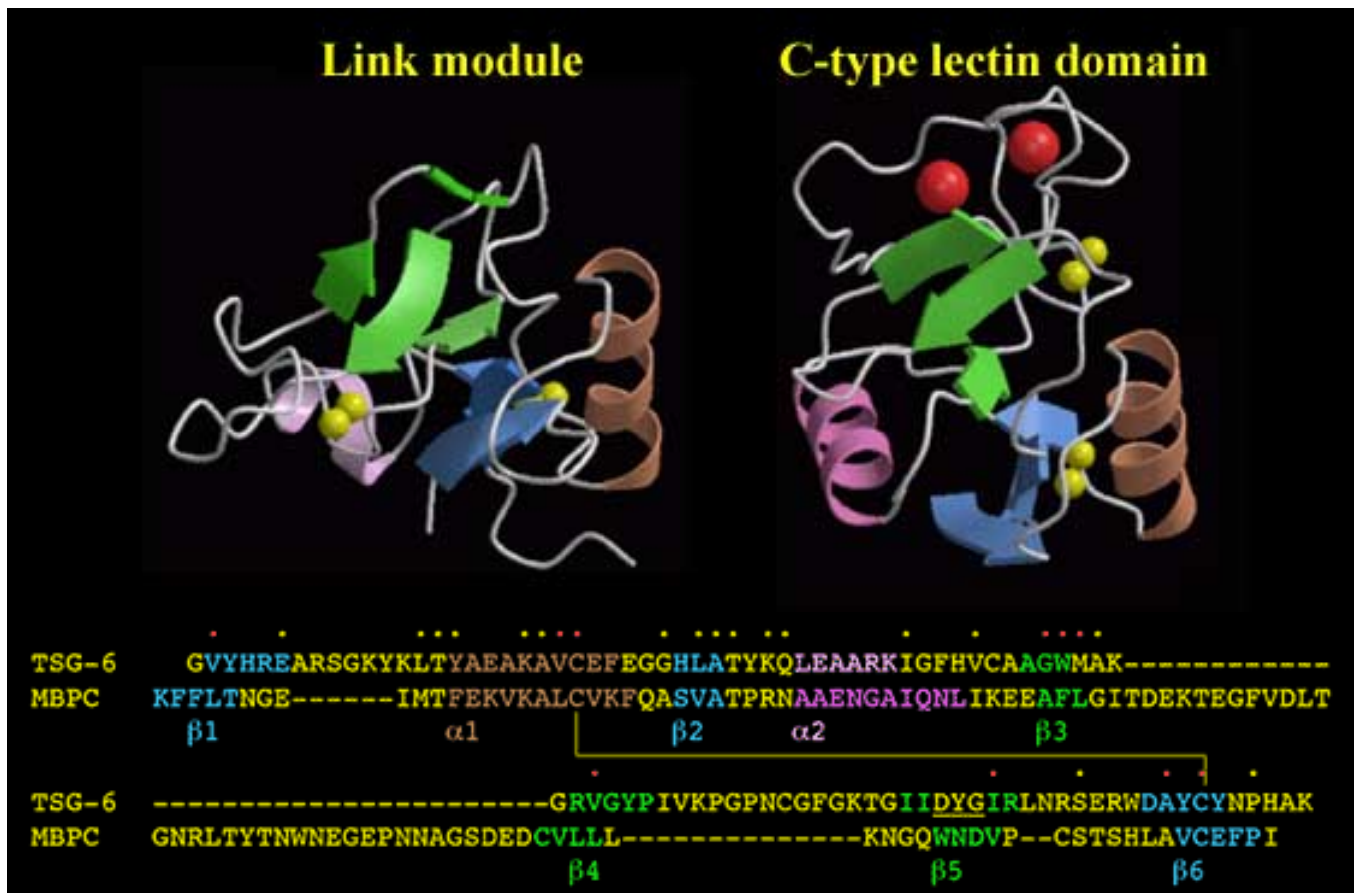


Figure 14.

The Link module and the C-type lectin domain. These modules have identical topologies and a very similar organization of secondary structure. An alignment of the Link module from human TSG-6 with the C-type lectin domain from human mannose-binding protein C-chain (MBPC) shows that the secondary structural elements are found in equivalent sequence positions. The yellow/red dots denote the positions of amino acid identities and conservative replacements. The hydrophobic interiors of the two domains are similar, with core residues in the C-type lectin domain found at equivalent sequence positions to those in the Link module (red dots). This includes consensus cysteines Cys1 and Cys4, which are linked by disulfide bonds in each module. The major difference between the structures is the length of two loops. The Link module has a short loop connecting the $\beta 3$ and $\beta 4$ strands, whereas in the C-type lectin domain, there is a region of ~35 amino acids containing residues responsible for chelating two Ca^{2+} ions (large red spheres). Similarly, the 14-residue loop between the $\beta 4$ and $\beta 5$ strands in the Link module is shorter in the lectin domain. The $\beta 5$ strand of the Link module is shown in two parts as it is interrupted by a bulge (underlined) as illustrated in Figure 5.

Contents

VIII Concluding Remarks Significant advances have been made in the last few years in our understanding of hyaluronan-protein interactions. However, this knowledge is far from complete, and further studies to determine the three-dimensional structures of different types of hyaluronan-binding domains, and their ligand complexes, are clearly essential if we are to unravel the molecular mechanisms underlying the biology of this amazing polysaccharide.

Contents

References

1. Kohda D, Morton CJ, Parkar AA, Hatanaka H, Inagaki FM, Campbell ID and Day AJ. Solution structure of the link module: a hyaluronan-binding domain involved in extracellular matrix stability and cell migration. *Cell* 86:767-775, 1996.
2. Banerji S, Ni J, Wang SX, Clasper S, Su J, Tammi R., Jones M and Jackson DG. LYVE-1, a new homologue of the CD44 glycoprotein, is a lymph-specific receptor for hyaluronan. *J Cell Biol* 144:789-801, 1999.
3. Wisniewski HG and Vilcek J. TSG-6: an IL-1/TNF-inducible protein with anti-inflammatory activity. *Cytokine Growth Factor Rev* 8:143-156, 1997.
4. Parkar AA, Kahmann JD, Howat SLT, Bayliss MT and Day AJ. TSG-6 interacts with hyaluronan and aggrecan in a pH-dependent manner via a common functional element: implications for its regulation in inflamed cartilage. *FEBS Lett* 428:171-176, 1998.
5. Parkar AA and Day AJ. Overlapping sites on the Link module of human TSG-6 mediate binding to hyaluronan and chondroitin-4-sulphate. *FEBS Lett* 410:413-417, 1997.
6. Bajorath J, Greenfield B, Munro SB, Day AJ and Aruffo A. Identification of CD44 residues important for hyaluronan binding and delineation of the binding site. *J Biol Chem* 273:338-343, 1998.
7. Day AJ. The structure and regulation of hyaluronan-binding proteins. *Biochem Soc Trans* 27:115-121, 1999.
8. Kahmann JD, O'Brien R, Werner JM, Heinegård D, Ladbury JE, Campbell ID and Day AJ. Localization and characterization of the hyaluronan-binding site on the link module from human TSG-6. *Structure* 8:763-774, 2000.
9. Peach RJ, Hollenbaugh D, Stamenkovic I and Aruffo A. Identification of hyaluronic acid binding sites in the extracellular domain of CD44. *J Cell Biol* 122:257-264, 1993.
10. Banerji S, Day AJ, Kahmann JD and Jackson DG. Characterization of a functional hyaluronan-binding domain from the human CD44 molecule expressed in *Escherichia coli*. *Protein Expr Purif* 14:371-381, 1998.
11. Grover J and Roughley PJ. The expression of functional link protein in a baculovirus system: analysis of mutants lacking the A, B and B' domains. *Biochem J* 300:317-324, 1994.
12. Watanabe H, Cheung SC, Itano N, Kimata K and Yamada Y. Identification of hyaluronan-binding domains of aggrecan. *J Biol Chem* 272:28057-28065, 1997.
13. Tammi R, MacCallum D, Hascall VC, Pienimäki JP, Hyttinen M and Tammi M. Hyaluronan bound to CD44 on keratinocytes is displaced by hyaluronan decasaccharides and not hexasaccharides. *J Biol Chem* 273:28878-28888, 1998.
14. Lesley J, Hascall VC, Tammi M and Hyman R. Hyaluronan binding by cell surface CD44. *J Biol Chem* 275:26967-26975, 2000.
15. Blundell CD, Kahmann JD, Perczel A, Mahoney DJ, Cordell MR, Teriete P, Campbell ID and Day AJ. Getting to grips with HA-protein interactions. In: *Hyaluronan 2000* (Kennedy JF, ed) Woodhead Publishing Ltd., Abington, Cambridge. 2001, in press.
16. English NM, Lesley JF and Hyman R. Site-specific de-N-glycosylation of CD44 can activate hyaluronan binding, and CD44 activation states show distinct threshold densities for hyaluronan binding. *Cancer Res* 58:3736-3742, 1998.
17. Schultz J, Copley RR, Doerks T, Ponting CP and Bork P. SMART: A Web-based tool for the study of genetically mobile domains. *Nucleic Acids Res* 28:231-234, 2000.
18. Lee JY and Spicer AP. Hyaluronan: a multifunctional, megaDalton, stealth molecule. *Curr Opin Cell Biology* 12:581-586, 2000.

Hyaluronan
INDEX

